

## Review Article

The Role of Dietary Peptides and the Gut Bacteria in Maintaining  
Intestinal and Homeostatic BalanceAsif Iqbal Khan<sup>1,2\*</sup>, Yi Xin<sup>1</sup><sup>1</sup>Department of Biotechnology, College of Basic Medical Science, Dalian Medical University, Dalian 116044, China.<sup>2</sup>Institute of Medical Technology, Dow University of Health Sciences, Karachi, Pakistan.\*Correspondence: [asif.iqbal@duhs.edu.pk](mailto:asif.iqbal@duhs.edu.pk)

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## Abstract

The gut bacteria and the host have a mutually beneficial relationship. This complex interdependence plays a critical role in resource collection via prebiotic carbohydrate breakdown in the intestinal tract, the supply of important nutrients, vitamin synthesis, metabolic operations, and maintenance, as well as sheltering against colonization of pathogens, and opportunistic fungal infections. This leads to 'gut microbiota homeostasis' or 'gut microbial equilibrium', i.e., a stable and healthy gastrointestinal tract's microbial community. In this review, we discuss the gut micro biome's role in the development of the immune system, including oral tolerance and immunity. The therapeutic approach must begin with the interaction of functional foods and enterocytes. Furthermore, we discuss current knowledge and the promising application of functional foods that may stimulate the immune system to perform a further pro- or anti-inflammatory activity in the gastrointestinal system. When evaluating the immune system function of nutraceuticals, designs that modulate the membrane must be used whenever possible. Moreover, nutraceutical products' claim to be immunity boosters should have valid and accurate data to back it up. While assessing the stimulation of blood cell immunity through functional foods is more useful, it does not accurately reflect physical reality, and an investigation into the colonic immune response is a better way to understand it.

**Keywords:** Functional peptide, intestine, intestinal microbiota, protein hydrolysate, bioactive peptide, immunomodulation

## 1. Gut Microbiota

Microbiota refers to the hundreds of billions of microbes that live inside and outside the human body, fungi, viruses, protozoa, and archaea. Bacterial ecology encompasses a wide range of ecosystems (Costello et al. 2012). However, With over 100 trillion bacteria, the human intestinal microbiota has the most diverse microbial population (Khudojberdievich 2022). The gut bacteria and the host have a mutually beneficial

relationship. (Dinan and Cryan 2012, Sommer and Bäckhed 2015). Immunological, mechanical, biological, and chemical barriers are the four layers of the intestinal barrier system that work together to keep the gastrointestinal tract in a state of homeostasis (Viggiano et al. 2015, Zhang et al. 2020). The intestinal barrier integrity is critical for a normal healthy body, which is typically retained by the immune system and intestinal bacteria working together (Wells et al. 2011).

The intestinal bacteria population has an impact on inflammation and immunological interactions. The importance of gut bacteria for the growth and development of host defense by influencing its immune function has been further elaborated in experimental models (Chen et al. 2022).

The host's pattern of genetic inheritance, method of gene transfer, eating habits, lifestyle, immune response, exposure to chemicals, drugs, and severity of symptoms are all the factors that impact the intestinal bacterial patterns (Dominguez-Bello et al. 2010, Maurice, Haiser, and Turnbaugh 2013, David, Maurice, et al. 2014, Tulstrup et al. 2015). The proportion of the intestinal microbiota varies throughout an individual's life (Bäckhed et al. 2015). Although the characterization of the gut flora population is usually steady during whole of the adulthood, yet numerous intrinsic and extrinsic variables, such as host nature and nurture, could also have an impact on it (Faith et al. 2013, David, Materna, et al. 2014, David, Maurice, et al. 2014, Goodrich et al. 2014). Similarly, numerous links have been found between intestinal flora and innate immune system's nutrition, metabolism, and immune function (Lee and Mazmanian 2010, Abreu 2010, Frank et al. 2007). Immune response diseases, coronary artery disease, ulcerative colitis, colorectal cancer, prediabetes, renal failure, and psychological disorders have all been linked to microbial demographic shifts (Nishida et al. 2018, Perez-Lopez et al. 2016, Lane, Zisman, and Suskind 2017, Zhu et al. 2011, Wang et al. 2011, Vaziri, Yuan, and Norris 2013, Cenit, Sanz, and Codoñer-Franch 2017, Dwivedy and Aich 2011). Understanding the importance of gut flora in connection to human health has deepened significantly. Most studies determine the relative abundance profiles for estimating microbial diversity in

the human intestine with the use of longitudinal analysis and comparisons of fecal 16S ribosomal RNA (rRNA) (Talapko et al. 2022). The causes behind varying prevalence of particular microorganisms and their capacity to cohabit with or reject others are unknown. It has been estimated that about 30,000 microbial interactions take place at the same time (Goyal et al. 2021, Donaldson, Lee, and Mazmanian 2016). Furthermore, microorganisms have preferences for certain sites in the gut, which influences their population density and degrees of activity in different regions, adding to the complexity. The microbiota's makeup is highly impacted by complicated microbial interactions, yet our understanding of this process is still limited (Faust and Raes 2012, Abu-Ali et al. 2018). A comparative study between the microbiomes of healthier people with the sick ones could reveal new information about its contribution to a disease pathology and prognosis. However, the underlying mechanisms, and connection of gut bacteria with the pathological changes have yet to be determined.

## **2. The Microbiota Composition**

The microorganism can be found all over the body, but the intestinal mucosa has the highest number of microorganisms. The gut mucosa provides a warm, moist, and nutrient-rich environment for microbial growth and survival. It secretes mucus, which serves as a barrier and a source of nourishment for microorganisms (Gustafsson and Johansson 2022). The intestine has a distinct immune system that encourages the growth of helpful bacteria while suppressing the growth of harmful microbes (Linares, Ross, and Stanton 2016). In various areas of the body, the gut bacteria component differs (Ravel et al. 2011). Microbial clades in a given body part are

comparable across individuals, such as oral micro flora, which is similar to other body regions, and there is a significant variation in the bacteriological communities among humans (Robinson, Bohannan, and Young 2010). The worldwide population shares roughly 1150 microorganism species, (Tilg and Kaser 2011, Nam et al. 2011). Firmicutes and Bacteroidetes are perhaps the greatest mutual intestinal flora on the phylum basis, responsible for nearly 90% of all genetically diverse types (Thangaleela et al. 2022, Cheng, Xu, and Shao 2022). Verrucobacteria, Actinobacteria, Proteobacteria, and Fusobacteria constitute a substantially lower percentage of the microbial composition (Scanlan and Marchesi 2008, Thangaleela et al. 2022, Cheng, Xu, and Shao 2022). Bacteroidetes (genus *Prevotella*, *Porphyromonas*, and *Bacteroides*) are gram-negative bacteria, while Firmicutes, which comprise *Enterococcus*, *Lachnospireaceae*, *Clostridiales*, *Streptococcaceae*, *Helicobacterium*, and *Lactobacillus*, contains roughly 50–60 percent of the bacteria (Ghoshal et al. 2018, Vickers 2017).

Fusobacteria, and Proteobacteria, are among the other phyla, Actinobacteria (genus *Collinsella*, *Bifidobacterium*) are gram-positive bacteria (Qin et al. 2010, Eckburg et al. 2005, Ghoshal et al. 2018). These main phyla contribute to the structural system of the intestinal population, which aids in protection of the host's health. The characterization of microbes throughout most people depends on the most prevailing and diverse species of the genera such as *Prevotella*, *Bacteroides*, or *Ruminococcus*, according to previous findings (Arumugam et al. 2011). Additionally, microbial diversity and proportions vary across different parts of the body (Goodman et al. 2011, Frank et al. 2007). The large intestine has an enormous microbial diversity; on the other hand, the small intestine has less

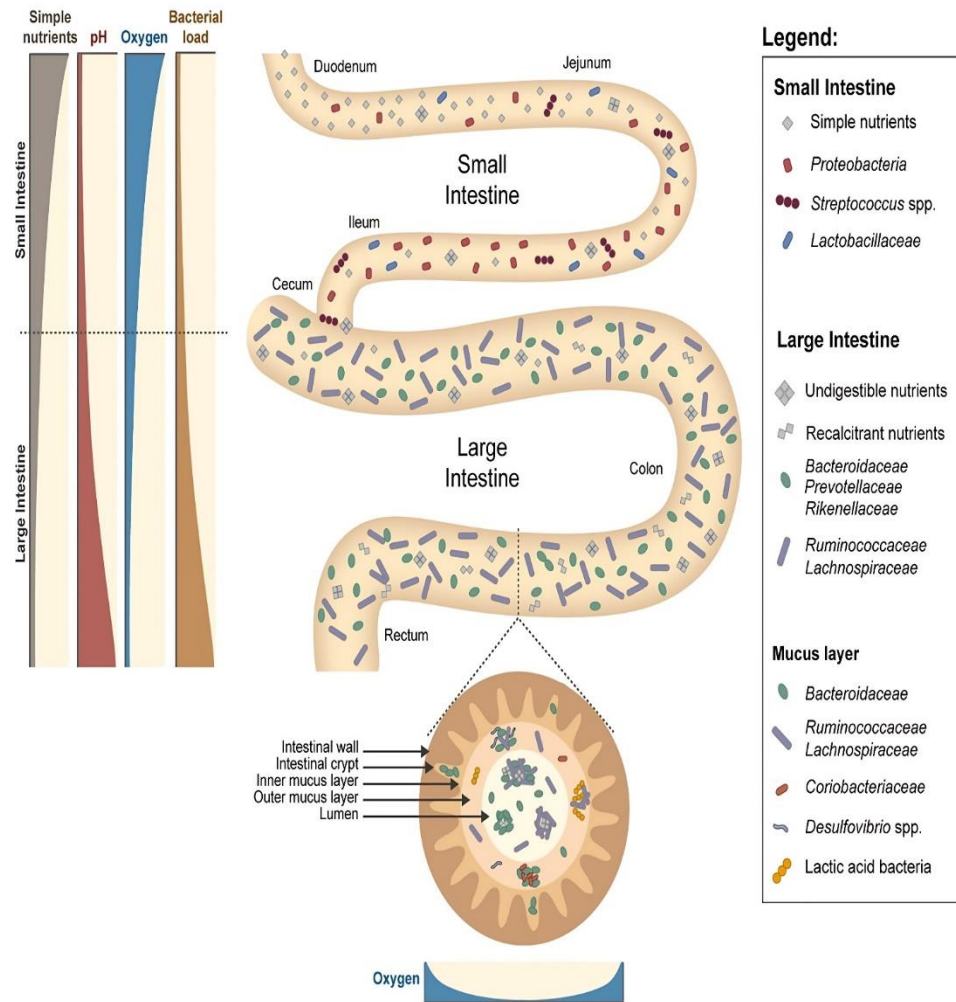
microbial distribution and abundance. Predominantly nutritional status affects enteral microbial bio-density (Frank et al. 2007, Wu et al. 2011).

### 3. Population of the Intestinal Tract

The intestine is a large digestive, absorbing, and immune organ in which gut microbiota and immunity are continually interacting to achieve immune intestinal integrity and hemostasis (Ying et al. 2020, Shi et al. 2017). The gastrointestinal tract has an acidic environment that is rich in a variety of bacterial communities (Elinav et al. 2011, Viviji, Aertsen, and Michiels 2016, Hillman et al. 2017). *Bacteroidetes*, *Proteobacteria*, *Firmicutes*, *Fusobacteria* and *Actinobacteria* are the most abundant taxa in the intestinal community, while *Deferribacteres*, TM7, and other taxa account for a smaller proportion (Zwiehler et al. 2011, Xu and Zhang 2015, Bik et al. 2006, Andersson et al. 2008). The bacterial composition of the esophageal and oral regions is very similar, with *Gemella*, *Prevotella*, *Actinomyces*, and *Streptococcus* (Andersson et al. 2008). The microbial variation occurs from the small intestine towards the colon, with indispensable anaerobic bacteria being the prominent taxa (Hayashi et al. 2005, Wang et al. 2003). The microbial density is greater in the gut, comprised of approximately 5000 microbiological taxa. Despite such diversity, only a few taxa are studied, with Firmicutes and Bacteroidetes among the most common phyla (Ley, Peterson, and Gordon 2006, Eckburg et al. 2005, Dethlefsen et al. 2008). In its first two years of age, the layout of pediatric gut bacteria differs from that of an adult's. *Enterococci*, *Staphylococci*, *Escherichia coli*, *Clostridia*, *Streptococci*, *Bacteroides*, *Enterobacteria*, and *Bifidobacteria* are by far the most prevalent taxa during infant stages

(Favier et al. 2002, Palmer et al. 2007, Mackie, Sghir, and Gaskins 1999). The mode of transmission, feeding, and environmental factors all have an impact on the microbial population of infants (Mackie, Sghir, and

Gaskins 1999). Throughout adulthood, there is less variation, but minor interferences do take place in some groups, such as *Bifidobacteria*, and *Lactobacillus* groups (Scanlan et al. 2006, Rajilić-Stojanović et al. 2009).



**Figure 1. The gut microbiota's compositional diversity throughout the intestinal system. The proximal parts of the small intestine have low bacterial loads due to pH and oxygen gradients along the longitudinal axis, whereas the large intestine has high number of bacterial loads (Pereira and Berry 2017).**

#### 4. Dysbiosis of the Gut Intestinal Microbiota

The intestinal microbes promote healthy intestinal function and regulate cellular metabolism (Jandhyala et al. 2015). The 'intestinal barrier system' consists of four layers that work together to keep the gastrointestinal tract in a state of homeostasis:

immunological, mechanical, biological, and chemical barriers (Viggiano et al. 2015, Zhang et al. 2020). Dysbiosis contributes to the growth of the gut microenvironment as well as in many external systemic diseases and metabolic syndromes (De Santis et al. 2015, Hansson 2012, Sommer and Bäckhed 2013).

The composition of the gut microbiota changes dynamically throughout infancy and stabilizes during adulthood. However, compositional changes can occur as a result of extrinsic and intrinsic factors. "Dysregulation" or "Dysbacteriosis" refers to the microbial change or alteration in the intestinal bacteria proportions. The characterization of the gut microbial group is influenced by a variety of elements. These include diet, host genetics, age, antibiotic use, geographic location, environment, disease and health status,

medications and supplements usage (Greenhalgh et al. 2016, Santoro et al. 2018). Microbiological dysbiosis has been associated with multiple factors such as inflammatory conditions (CD & UC), metabolic and immune disorders (diabetes, obesity-related diseases), among others. If the hemostatic imbalance occurs, dysbiosis of gut microbial communities will trigger a variety of diseases and abnormal immune responses (Berg, Clemente, and Colombel 2015).

**Table 1. Microbial metabolites or components that are implicated in disease (Rooks and Garrett 2016).**

Human diseases and preclinical models	Microbial metabolites and components	Refs
<b>Allergic and Immune Disorders</b>		
Asthma	SCFAs	40,45
Inflammatory Bowel Disease	SCFAs	42-44, 53
	B Vitamins	53
<b>Cancer</b>		
Colorectal cancer	SCFAs	53,118,119
	B Vitamins	53
	N <sup>1</sup> , N <sup>12</sup> -diacetylspermine	80
<b>Gynaecological and Reproductive Disorders</b>		
Bacterial vaginosis and other sexually transmitted infections	Polyamine	120
	HBP	100
Preterm labour	SCFAs	50
<b>Metabolic Disorders</b>		
Cardiovascular disease	TMAO	121
Kidney disease	SCFAs	122
	p-Cresol	123
Obesity and metabolic syndrome	TMAO	121
Type 2 diabetes	TMAO	121
<b>Neurological Disorders</b>		
Autism spectrum disorder	4-EPS	124
Central nervous system dysfunction	SCFAs	52,125
<b>Other Gastrointestinal Disorders</b>		
Infectious colitis ( <i>Clostridium difficile</i> )	Bile acids	126

4-EPS, 4-ethyl phenol sulfate; HBP, D-glycero-β-D-manno-heptose-1,7-biphosphate; SCFAs, short-chain fatty acids; TMAO, trimethylamine N-oxide.

## 5. Immune System

The immune system is the body's capacity to mount rapid, precise, and protective responses against a diverse range of potentially harmful microorganisms, including viruses, parasites, bacteria, and fungi that are prevalent in our surroundings (Paul 2013, Williams 2011a). The immune system is divided into two components that work together effectively despite their differences. The host immune system comes first, followed by the adaptive immune system. When confronted with a pathogen, the innate immune system responds quickly, whereas adaptive immunity is highly specialized and capable of recognizing a pathogenic organisms. However, adaptive immunity develops slower than innate immunity and occurs in specified organs within the lymph glands (Aderem and Ulevitch 2000, Lamb 2012). The Immune System, both Innate and Adaptive is made up of Secondary lymphatic tissues, main lymphoid organs, and several immune cells. The body's innate immune response consists of chemical barriers (reactive oxygen and antimicrobial peptides), soluble mediators (complement system), innate immune cells, innate antibodies, physical barriers (skin and mucous membranes), and associated cytokines (Koenderman, Buurman, and Daha 2014, Williams 2011b). Adaptive immunity is more extensive, and highly attuned to variety of self, non-self-antigen recognition, T-cell receptors, and clone-specific antibodies, modulated in reconfiguring Ig gene superfamily genes, direct the immune system (Bonilla and Oettgen 2010, Medzhitov and Janeway Jr 1997). Chemical and physical barriers prevent pathogen entry. Complement system and other humoral factors inhibit infection through phagocytosis, removal by cytotoxicity mechanism, activate antigen-presenting cells for T and B lymphocytes, and

cytokine synthesis of adaptive immune response (Williams 2011b, Beutler 2004, Tosi 2005). This results in immunological effector pathways unique to pathogens, immunologic memory formation, and immune homeostasis modulation in the host (Bonilla and Oettgen 2010).

## 6. The microbiome of gut and mucosal immune system

The mucous membrane plays a crucial function in the immune system because it contains specialized chemical and mechanical boundaries that keeps foreign contaminants from entering by disintegrating them through a wide variety of chemical processes, the majority of which are yet to be understood (Ogra et al. 2012). The mucosal immune system is unique because it contains cells that aren't even present in the circulation and have a distinct immunological response, as well as the secretory antibody IgA (Dwivedy and Aich 2011, Phalipon et al. 2002). Most infections aim to enter host body through the digestive system. Fortunately, it has the biggest mucosal barrier of almost any molecular mechanisms involved. The most important portion of intestinal epithelium, intestinal epithelial cells, regulates variety of gastrointestinal activities including metabolism, secretions, absorption, and protection (Phalipon et al. 2002, Strober, Fuss, and Blumberg 2002).

Moreover, it plays a prominent part in antigen presentation, with the opportunity to demonstrate two very different Class I and Class II antigens along with releasing immunologically active substances in responses to pro-inflammatory mediators, protecting the gastrointestinal mucosa and lumen from injury (Bland 1998). T helper (Th) cells are critical in autoimmunity even though they eliminate infections even during the host defense system and cause inflammatory

responses, ultimately contributing to internal necrosis. T regulatory cell plays an important role in immunological tolerance and inflammatory regulation. As just a result, gastrointestinal autoimmunity is linked to the imbalance of T regulatory as well as pro-inflammatory T helper lymphocytes inside the intestine (Colombo et al. 2015, Shi et al. 2017, Johansson, DeNardo, and Coussens 2008). When the equilibrium is disrupted, the gastrointestinal immune response malfunctions, resulting in disorders including inflammatory bowel disease (IBD). IBD is a complicated disorder caused by inherited, ecological, and microbiological reasons which leads to inflammatory response by activating an abnormal immune response. IBDs such as ulcerative colitis (UC) and Crohn's disease (CD) are common (Berg, Clemente, and Colombel 2015).

## 7. Immunoregulatory Pathways

The gut microbiota has a strong influence on the health of the intestines and their host. Intestinal epithelial cells (IECs) create a physiological and immunologic shield to prevent the intestinal flora from pattern recognition receptor-mediated degradation (Carvalho et al. 2012). Innate immune processes, such as microbial substance ligation of targets dedicated to the identification of microbe-associated pattern recognition receptors, is most likely used to regulate colonic microorganisms in the intestinal epithelium, macrophages of spleen, lymph nodes, and bone marrow (Daddaoua et al. 2013). It included Toll-like receptors (TLRs), which may help with maintaining intestinal homeostasis by improving the epithelial intestinal barrier and innate immune expression of genes (Daddaoua et al. 2013, Zhao et al. 2020). Toll-like receptor 4 (TLR4) is a transmembrane protein, member of the toll-

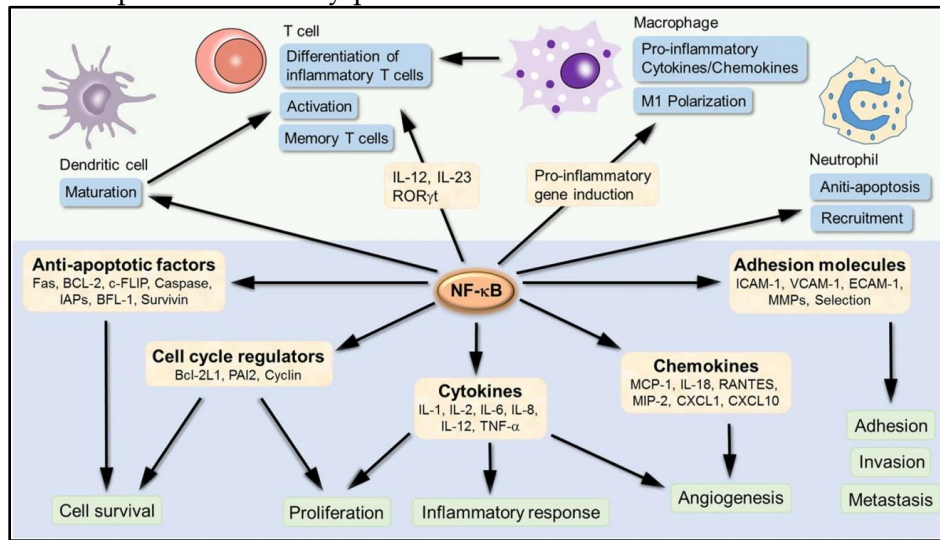
like receptor family, which belongs to the pattern recognition receptor (PRR) family. It plays a significant role in boosting our natural immune responses and triggering the production of certain proteins called cytokines. This process occurs when a molecule called MyD88 binds to TLR4 inside the cell, which then sets off a series of chemical reactions and signals within the cell. These signals help activate various important substances and pathways within the cell. (Li et al. 2020, Liu et al. 2021, Xiang et al. 2021). MyD88 activates the IKK complex, which releases substances like IL-1 $\beta$  and TNF- $\alpha$ . These substances activate the immune system through the NF- $\kappa$ B pathway. In short, MyD88 triggers a process that boosts immune response. (Chen, Chen, et al. 2019, Mitchell, Vargas, and Hoffmann 2016).

NF- $\kappa$ B (nuclear factor- $\kappa$ B) is a critical transcriptional factor that is composed of Rel family heterodimer proteins p65 and p50, NF- $\kappa$ B is a transcription factor that affects the expression of several genes as well as immunological activities. It also is implicated in immunological and inflammatory functions (Gilmore, Siggers, and Gerondakis 2015, Liang, Zhou, and Shen 2004). The I $\kappa$ B kinase (IKK) protein kinase is a key part of the NF- $\kappa$ B signaling pathway and is crucial for modulating NF- $\kappa$ B stimulation and gene regulation. It stimulates the phosphorylated and ubiquitination breakdown mechanism of the I $\kappa$ B protein, resulting in the release of the NF- $\kappa$ B dimer (Li et al. 2019, Zhang et al. 2019). Even amongst the I $\kappa$ B family proteins, I $\kappa$ B- $\alpha$  is primarily responsible for the baseline suppression of NF- $\kappa$ B activity. In resting cells, the inhibitory protein I $\kappa$ B adheres to the NF- $\kappa$ B dimer and adsorb it in the cytoplasm in an inactive form. When cells are stimulated by numerous stimulating substances activated I $\kappa$ B kinase (IKK). The active NF- $\kappa$ B dimer

enters the nucleus and attaches to genes include TNF- $\alpha$ , IL-8, ICAM-1, COX-2, and IL-1 $\beta$ , (Laveti et al. 2013, Lee and Yang 2012, Agarwal and Shanmugam 2022) that provide an NF- $\kappa$ B-binding receptor, causing the transcription and production of inflammatory and pro-inflammatory mediators to activate. The nuclear transcription factor NF- $\kappa$ B is required for the control of oxidative stress, pro-inflammatory, and immunological responses (Li et al. 2019, DiDonato, Mercurio, and Karin 2012, Zhang et al. 2019).which determines immunological and inflammation processes regulates the genes involved to modulate innate and acquired immunity, B cell development, inflammatory processes, lymphoid organ synthesis, and stress reaction, among other cellular mechanisms (Hayden and Ghosh 2004, Zhao et al. 2020, Wang et al. 2019).

Furthermore, IL-6-induced intracellular signaling increases the production of associated genes like iNOS. Nitric oxide is a free nitrogen oxide radical that is a crucial constituent of a cell for stimulating macrophages and plays a vital role in the non-specific immune response. It not only protects

against extracellular microbes and cancer cells but that also responsible for regulating immunity-related biological functions (Guzik, Korbust, and Adamek-Guzik 2003, Li et al. 2019, Zhang et al. 2019). NO synthase (NOS) in mammals is divided into three categories: endothelial (eNOS), neuronal (nNOS), and inducible (iNOS). The most essential enzyme in NO synthesis, iNOS, is quickly stimulated and demonstrated in activated macrophages (Li et al. 2019, Vannini, Kashfi, and Nath 2015, Sessa 1994). The isoenzyme iNOS is responsible for the majority of NO synthesis. In macrophages, cytokines such as TNF- $\alpha$ , interferon- $\gamma$  (IFN- $\gamma$ ), and lipopolysaccharide stimulate the interpretation of iNOS mRNA and protein. Through NADPH and oxygen-dependent mechanism, iNOS catalyzes the synthesis of NO and L-citrulline from L-arginine (Hu et al. , Zhang et al. 2019). The NF- $\kappa$  B is one of the most essential signaling pathways for generating iNOS expression in cells, which eventually leads to an increased in NO production, and NO plays a key role in macrophage activation (Tao et al. 2018, Vannini, Kashfi, and Nath 2015).



**Figure 2.** NF $\kappa$ B has an important role in the generation of inflammatory markers, chemokines, and cell adhesion, as well as in the cell cycle regulation, apoptosis, morphogenesis, and differentiation. (Liu et al. 2017).

## 8. Dietary Modification of Gut Microbiota

There has been implementation and evaluation of the gut microbiota in recent years, with the logical use of probiotics as a functional food being particularly well established (Shi et al. 2017, Delcenserie et al. 2008, Gamallat et al. 2019). Pathogens cause infections and are closely associated with illness onset or progression. While commensal gut microbiota influences both genetic and epigenetic factors that directly or indirectly modulates host health (Kurashima, Goto, and Kiyono 2013, Gamallat et al. 2019). Functional foods are non-digestible dietary products/natural ingredients that foster healthy microorganisms in the large gastrointestinal system (Roberfroid 2007). Protective benefits have been discovered in a portion from casein hydrolysate for about the first time in 1984 (Kiewiet, Faas, and De Vos 2018).

## 9. Protein Hydrolysates

Protein hydrolysates are commonly used in commercial applications as an important food supplement and are comprised of two different proteins and peptides that are formed when complete proteins hydrolysis occurs (Kiewiet, Faas, and De Vos 2018). Several studies on functional foods, marine protein hydrolysate, and bioactive peptides for immune system enhancement have been published in recent years (Ambigaipalan and Shahidi 2017). A bioactive peptide is a polypeptide group that has been degraded by enzymatic digestion of natural and marine dietary protein. The most frequent method is the enzymatic hydrolysis of the whole protein to create bioactive peptides using enzymes such as chymotrypsin, trypsin, pepsin, alcalases, thermolysin, and pancreatin (Bhat et al. 2015, Chen, Su, et al. 2019). These peptides are made up of amino acids that are arranged

in different ways. It contains more than 2 -20 amino acid residues structurally because of its hydrophobic characteristics and small dimensions. Bioactive chemicals are much easier to absorb (Hartmann and Meisel 2007). Peptide 6 kDa has a high molecular weight, good absorption, and various health benefits, including antimicrobial, antihypertensive, antimicrobial, antioxidative, antithrombotic, anticancer efficacies, and immunomodulatory effects on the adaptive and innate immune response. Bioactive peptides in the form of short peptides, dipeptides, and tripeptides are absorbed faster than the same composition of the free amino acid (Chen, Su, et al. 2019).

## 10. The Impact of Protein Hydrolysates on Gastric Barrier and Immunity

In the intestinal lumen, epithelial cells serve as the first line of defense against dangerous bacteria and chemicals. A coating of mucus protects these cells, so the luminal composition and the rest of the body are separated by this physical barrier (Farhadi et al. 2003). Protein hydrolysates and bioactive peptides fortify the epithelial barrier and increase mucus production to defend against harmful microorganisms (De Santis et al. 2015). Peptides may play a significant role in the intestine by regulating digestive enzymes, stabilizing the intestinal epithelial tight junctions, and modulating dietary absorption (Shimizu 2004, Korhonen and Pihlanto 2006). They also increase mucus production and the number of goblet and Paneth cells, which regulate mucus and anti-bacterial peptide production (Elshaer and Begun 2017). Dairy-derived peptides, such as those found in yoghurt, can boost the production of mucus-related genes and anti-bacterial factors (Plaisancié et al. 2013). Epithelial cells regulate immune cell differentiation by secreting cytokines. Consumption of egg yolk peptide,

for example, enhanced IL-6 production in epithelial cells, can influence innate and adaptive immunological responses in mice (Nelson et al. 2007). However, the overall consequence of elevated IL-6 is complex and context-dependent. The concentration of IgA in the colon following hydrolysate delivery in the animal study is among the most researched immunological responses, As proven in the case of a common carp egg hydrolysate, hydrolysate consumption might produce an increase in IgA level (Chalamaiah et al. 2015). Changes in gene expression in target cells consist of the cellular responses to most cytokines, numerous cytokine-induced changes in gene expression result in T and B lymphocyte differentiation and effector cells activation such as macrophages. IL-2 becomes cytokine-producing by T-helper cells and is a key component of immune response to trigger T cell proliferation and promote B cell growth, acting on the natural killer, glioma, and Th1 cell. IL-4 is a cytokine that can be produced by activated Th2 cells, stimulate B cells growth, differentiation of B cells, and effects on T, B cells, endothelial cells, fibroblasts and mast cells, and macrophages (Hou et al. 2016, Abraha 2020). IL-10 is an inflammatory cytokine that inhibits the host immune system and regulates immune-mediated inflammation. It particularly inhibits the responses involving activated macrophages and dendritic cells. Protein hydrolysates, particularly those generated from cow's milk, offer anti-inflammatory properties that prevent intestinal damage and weight loss in colitis and ileitis models. Furthermore, they increase IL-10 and other regulatory cytokines while decreasing pro-inflammatory cytokines (Fernandez-Tome et al. 2019, Khan, Rehman, Farooqui, Siddiqui, Ayub, Ramzan, Zexu, et al. 2022). IFN- $\gamma$  is an important cytokine and is critical for its immune-stimulatory and

immunomodulatory effects, which activate the macrophages, regulate the cells of Th1, Th2 and induce the generation of T cells (Hou et al. 2016, Abraha 2020). Protein hydrolysates have been demonstrated to upregulate genes involved in innate immunity and host defense in gut-associated lymphoid tissue, specifically the Peyer's patches. In another investigation on adaptive responses, sensitization with partially hydrolyzed whey protein inhibited the growth of specific T cell types in the Peyer's patches following a challenge with intact whey (Kiewiet et al. 2015, Adolfsson, Meydani, and Russell 2004). These findings suggest that protein hydrolysates can modulate various aspects of the immune system in the intestine, impacting IgA levels, cytokine production, inflammation, and adaptive immune responses.

The mesenteric lymph nodes (MLN) are responsible for tolerance induction and antigen presentation processes (Rescigno 2011). Dendritic cells carrying antigens from the Peyer's patches and lamina propria migrate to the MLN, where they activate T and B cells, leading to immune response or tolerance induction (Pabst and Mowat 2012). Studies using bovine milk hydrolysates or peptides in murine sensitization models have examined the cell types present in the MLN after hydrolysate consumption (Kiewiet, Faas, and De Vos 2018). Milk peptides alone or in combination with indigestible oligosaccharides fed to mice prior to sensitization with intact whey enhanced Treg population in the MLN (Meulenbroek et al. 2013). Consumption of casein-derived glycomacropeptide enhanced tolerogenic IL-10 production in MLN cells of a mouse model with colitis, indicating a potential increase in Treg (Lozano-Ojalvo and López-Fandiño 2018).

In addition to T cell responses, B cells can be activated in the MLN, leading to follicle expansion. Sensitization with partially hydrolyzed whey protein increased the number of IgA+ B cells (Kiewiet et al. 2017). Protein hydrolysates affect the immune system both locally and systemically. Peptides produced from hydrolysates have been found in the blood after consuming dairy and soy products (Chalamaiah et al. 2014). Different protein hydrolysates, such as those from milk, oyster, salmon, and fish, enhance macrophage phagocytosis in the peritoneal cavity and increase NK cell activity in the spleen (Martínez-Medina et al. 2022). Administration of specific hydrolysates, including oyster, tuna cooking drip, salmon, and common carp eggs, promotes splenocyte proliferation in mice (Chalamaiah et al. 2014). These hydrolysates can modulate the populations of CD4+ and

CD8+ cells, induce a mixed Th1/Th2 response, and impact the production of cytokines (Martínez-Medina et al. 2022). Whey hydrolysate increases Treg and regulatory B cells (Breg) in the spleen, while casein hydrolysate induces IL-10-producing Breg. Protein hydrolysate consumption also leads to increased antibody levels, such as IgG and IgA (Khan, Rehman, Farooqui, Siddiqui, Ayub, Ramzan, Wang, et al. 2022). Studies on humans show similar immunomodulatory effects, including altered leukocyte numbers, increased CD11b+ and CD56+ cells, and enhanced NK cell activity (Bernardini et al. 2016, Yimit et al. 2012). However, further well-designed studies are necessary to comprehensively understand the effects of protein hydrolysates on human immune responses.

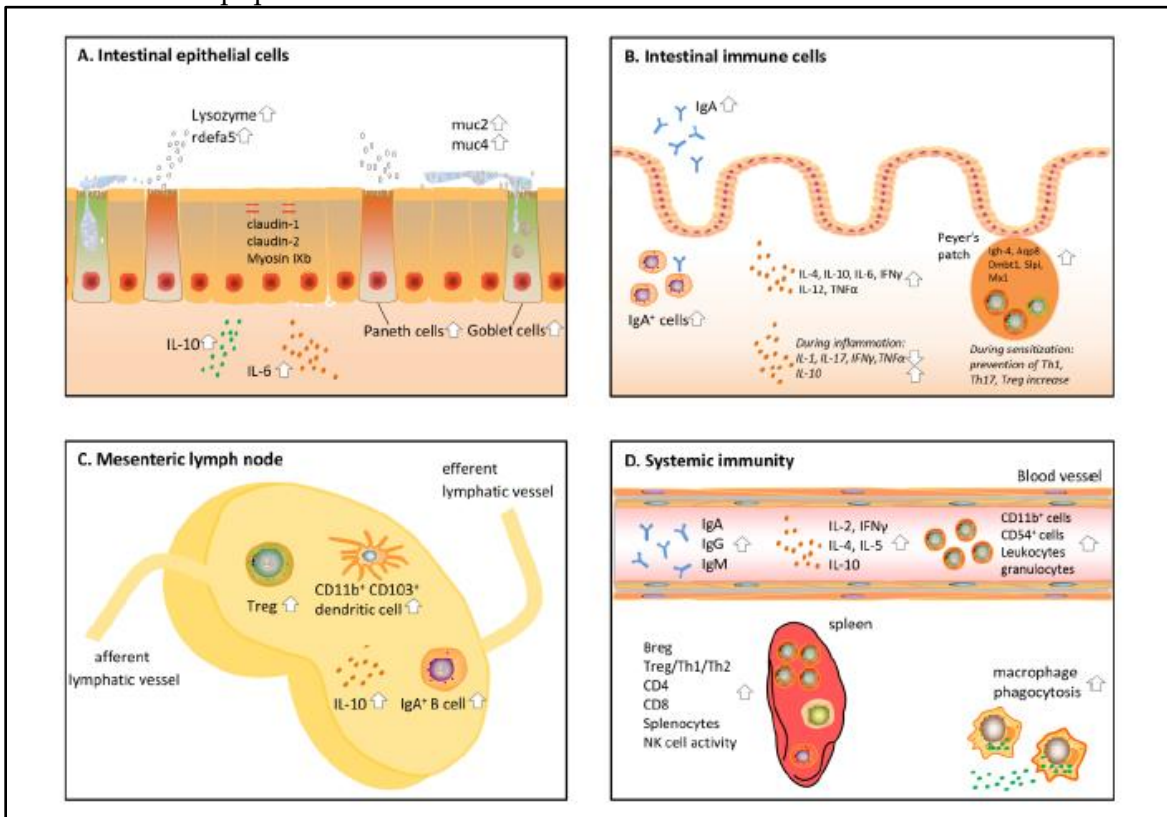


Figure 3. A summary of the immunological activities of protein hydrolysates (Kiewiet, Faas, and De Vos 2018).

## 11. Conclusions

The gut microbiota, consisting of diverse microorganisms, plays a vital role in maintaining intestinal function, and protecting against pathogens. The immune system, including innate and adaptive components, defends the body against harmful microorganisms. The gut microbiota and immune system have a complex relationship, and dysbiosis of the gut microbiota can lead to various diseases and abnormal immune responses. The mucosal immune system, present in the gastrointestinal tract, plays a crucial role in preventing the entry of pathogens. Functional foods, such as protein hydrolysates and bioactive peptides, have been found to modulate the immune system and offer potential benefits for intestinal health. Further research is needed to fully understand the effects of protein hydrolysates on human immune responses.

## Conflict of Interest

All contributing authors declare no conflicts of interest.

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## Ethics Approval

NA

## Consent Forms

NA

## Data Availability

NA

## Authors Contribution

AIK conceptualized the study and wrote the final manuscript, AIK and YX did the literature analysis, AIK and YX contributed to

manuscript writing and AIK supervised the whole project and wrote the final manuscript.

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## References

- Abraha, Rigbe. 2020. "Review on the role and Biology of Cytokines in Adaptive and Innate Immune System." *Archives of Veterinary and Animal Sciences* no. 2 (2).
- Abreu, Maria T. 2010. "Toll-like receptor signalling in the intestinal epithelium: how bacterial recognition shapes intestinal function." *Nature Reviews Immunology* no. 10 (2):131-144.
- Abu-Ali, Galeb S, Raaj S Mehta, Jason Lloyd-Price, Himel Mallick, Toby Branck, Kerry L Ivey, David A Drew, Casey DuLong, Eric Rimm, and Jacques Izard. 2018. "Metatranscriptome of human faecal microbial communities in a cohort of adult men." *Nature microbiology* no. 3 (3):356-366.
- Aderem, Alan, and Richard J Ulevitch. 2000. "Toll-like receptors in the induction of the innate immune response." *Nature* no. 406 (6797):782-787.
- Adolfsson, Oskar, Simin Nikbin Meydani, and Robert M Russell. 2004. "Yogurt and gut function." *The American journal of clinical nutrition* no. 80 (2):245-256.
- Agarwal, Happy, and Venkat Kumar Shanmugam. 2022. "Mechanism-based approach of medicinal plants mediated treatment of inflammatory disorders: A review." *South African Journal of Botany* no. 147:380-390.
- Ambigaipalan, Priyatharini, and Fereidoon %J Journal of Functional Foods Shahidi.

2017. "Bioactive peptides from shrimp shell processing discards: Antioxidant and biological activities." no. 34:7-17.
- Andersson, Anders F, Mathilda Lindberg, Hedvig Jakobsson, Fredrik Bäckhed, Pål Nyrén, and Lars Engstrand. 2008. "Comparative analysis of human gut microbiota by barcoded pyrosequencing." *PloS one* no. 3 (7):e2836.
- Arumugam, Manimozhiyan, Jeroen Raes, Eric Pelletier, Denis Le Paslier, Takuji Yamada, Daniel R Mende, Gabriel R Fernandes, Julien Tap, Thomas Bruls, and Jean Michel Batto. 2011. "Erratum: Enterotypes of the human gut microbiome (Nature (2011) 473 (174-180))." *Nature* no. 474 (7353):666.
- Bäckhed, Fredrik, Josefine Roswall, Yangqing Peng, Qiang Feng, Huijue Jia, Petia Kovatcheva-Datchary, Yin Li, Yan Xia, Hailiang Xie, and Huanzi Zhong. 2015. "Dynamics and stabilization of the human gut microbiome during the first year of life." *Cell host & microbe* no. 17 (5):690-703.
- Berg, Dana, Jose C Clemente, and Jean-Frederic Colombel. 2015. "Can inflammatory bowel disease be permanently treated with short-term interventions on the microbiome?" *Expert review of gastroenterology & hepatology* no. 9 (6):781-795.
- Bernardini, Giovanni, Fabrizio Antonangeli, Valentina Bonanni, and Angela Santoni. 2016. "Dysregulation of chemokine/chemokine receptor axes and NK cell tissue localization during diseases." *Frontiers in immunology* no. 7:402.
- Beutler, Bruce. 2004. "Innate immunity: an overview." *Molecular immunology* no. 40 (12):845-859.
- Bhat, ZF, Sunil Kumar, Hina Fayaz %J Journal of Food Science Bhat, and Technology. 2015. "Bioactive peptides of animal origin: a review." no. 52 (9):5377-5392.
- Bik, Elisabeth M, Paul B Eckburg, Steven R Gill, Karen E Nelson, Elizabeth A Purdom, Fritz Francois, Guillermo Perez-Perez, Martin J Blaser, and David A Relman. 2006. "Molecular analysis of the bacterial microbiota in the human stomach." *Proceedings of the National Academy of Sciences* no. 103 (3):732-737.
- Bland, Paul W. 1998. "Mucosal T cell-epithelial cell interactions." *Mucosal T Cells* no. 71:40-63.
- Bonilla, Francisco A, and Hans C Oettgen. 2010. "Adaptive immunity." *Journal of Allergy and Clinical Immunology* no. 125 (2):S33-S40.
- Carvalho, Frederic A, Jesse D Aitken, Matam Vijay-Kumar, and Andrew T Gewirtz. 2012. "Toll-like receptor-gut microbiota interactions: perturb at your own risk!" *Annual review of physiology* no. 74:177-198.
- Cenit, María Carmen, Yolanda Sanz, and Pilar Codoñer-Franch. 2017. "Influence of gut microbiota on neuropsychiatric disorders." *World journal of gastroenterology* no. 23 (30):5486.
- Chalamaiah, M, R Hemalatha, T Jyothirmayi, Prakash V Diwan, K Bhaskarachary, A Vajreswari, R Ramesh Kumar, and B Dinesh Kumar. 2015. "Chemical composition and immunomodulatory effects of enzymatic protein hydrolysates from common carp (*Cyprinus carpio*) egg." *Nutrition* no. 31 (2):388-398.
- Chalamaiah, M, R Hemalatha, T Jyothirmayi, Prakash V Diwan, P Uday Kumar, Chetan Nimgulkar, and B Dinesh

- Kumar. 2014. "Immunomodulatory effects of protein hydrolysates from rohu (*Labeo rohita*) egg (roe) in BALB/c mice." *Food Research International* no. 62:1054-1061.
- Chen, Caixia, Xiulan Su, Zhiwei %J Experimental Hu, and therapeutic medicine. 2019. "Immune promotive effect of bioactive peptides may be mediated by regulating the expression of SOCS1/miR-155." no. 18 (3):1850-1862.
- Chen, Dan, Guijie Chen, Yu Ding, Peng Wan, Yujia Peng, Chunxu Chen, Hong Ye, Xiaoxiong Zeng, and Linwu Ran. 2019. "Polysaccharides from the flowers of tea (*Camellia sinensis* L.) modulate gut health and ameliorate cyclophosphamide-induced immunosuppression." *Journal of Functional Foods* no. 61:103470.
- Chen, Yanfei, Jin Lin, Lanlan Xiao, Xuan Zhang, Lidan Zhao, Min Wang, and Lanjuan Li. 2022. "Gut microbiota in systemic lupus erythematosus: A fuse and a solution." *Journal of Autoimmunity* no. 132:102867.
- Cheng, Ting, Chen Xu, and Jing Shao. 2022. "Updated immunomodulatory roles of gut flora and microRNAs in inflammatory bowel diseases." *Clinical and Experimental Medicine*:1-17.
- Colombo, Bruno M, Thibault Scalvenzi, Sarah Benlamara, and Nicolas Pollet. 2015. "Microbiota and mucosal immunity in amphibians." *Frontiers in immunology* no. 6:111.
- Costello, Elizabeth K, Keaton Stagaman, Les Dethlefsen, Brendan JM Bohannon, and David A Relman. 2012. "The application of ecological theory toward an understanding of the human microbiome." *Science* no. 336 (6086):1255-1262.
- Daddaoua, Abdelali, Enrique Martínez-Plata, Mercedes Ortega-González, Borja Ocón, Carlos J Aranda, Antonio Zarzuelo, María D Suárez, Fermín Sánchez de Medina, and Olga Martínez-Augustin. 2013. "The nutritional supplement Active Hexose Correlated Compound (AHCC) has direct immunomodulatory actions on intestinal epithelial cells and macrophages involving TLR/MyD88 and NF- $\kappa$ B/MAPK activation." *Food chemistry* no. 136 (3-4):1288-1295.
- David, Lawrence A, Arne C Materna, Jonathan Friedman, Maria I Campos-Baptista, Matthew C Blackburn, Allison Perrotta, Susan E Erdman, and Eric J Alm. 2014. "Host lifestyle affects human microbiota on daily timescales." *Genome biology* no. 15 (7):1-15.
- David, Lawrence A, Corinne F Maurice, Rachel N Carmody, David B Gootenberg, Julie E Button, Benjamin E Wolfe, Alisha V Ling, A Sloan Devlin, Yug Varma, and Michael A Fischbach. 2014. "Diet rapidly and reproducibly alters the human gut microbiome." *Nature* no. 505 (7484):559-563.
- De Santis, Stefania, Elisabetta Cavalcanti, Mauro Mastronardi, Emilio Jirillo, and Marcello Chieppa. 2015. "Nutritional keys for intestinal barrier modulation." *Frontiers in immunology* no. 6:612.
- Delcenserie, Véronique, D Martel, M Lamoureux, J Amiot, Y Boutin, and D Roy. 2008. "Immunomodulatory effects of probiotics in the intestinal tract." *Current issues in molecular biology* no. 10 (1-2):37-54.

- Dethlefsen, Les, Sue Huse, Mitchell L Sogin, and David A Relman. 2008. "The pervasive effects of an antibiotic on the human gut microbiota, as revealed by deep 16S rRNA sequencing." *PLoS biology* no. 6 (11):e280.
- DiDonato, Joseph A, Frank Mercurio, and Michael Karin. 2012. "NF- $\kappa$ B and the link between inflammation and cancer." *Immunological reviews* no. 246 (1):379-400.
- Dinan, Timothy G, and John F Cryan. 2012. "Regulation of the stress response by the gut microbiota: implications for psychoneuroendocrinology." *Psychoneuroendocrinology* no. 37 (9):1369-1378.
- Dominguez-Bello, Maria G, Elizabeth K Costello, Monica Contreras, Magda Magris, Glida Hidalgo, Noah Fierer, and Rob Knight. 2010. "Delivery mode shapes the acquisition and structure of the initial microbiota across multiple body habitats in newborns." *Proceedings of the National Academy of Sciences* no. 107 (26):11971-11975.
- Donaldson, Gregory P, S Melanie Lee, and Sarkis K Mazmanian. 2016. "Gut biogeography of the bacterial microbiota." *Nature Reviews Microbiology* no. 14 (1):20-32.
- Dwivedy, Abhisek, and Palok Aich. 2011. "Importance of innate mucosal immunity and the promises it holds." *International journal of general medicine* no. 4:299.
- Eckburg, Paul B, Elisabeth M Bik, Charles N Bernstein, Elizabeth Purdom, Les Dethlefsen, Michael Sargent, Steven R Gill, Karen E Nelson, and David A Relman. 2005. "Diversity of the human intestinal microbial flora." *science* no. 308 (5728):1635-1638.
- Elinav, Eran, Till Strowig, Andrew L Kau, Jorge Henao-Mejia, Christoph A Thaiss, Carmen J Booth, David R Peaper, John Bertin, Stephanie C Eisenbarth, and Jeffrey I Gordon. 2011. "NLRP6 inflammasome regulates colonic microbial ecology and risk for colitis." *Cell* no. 145 (5):745-757.
- Elshaer, Dana, and Jakob Begun. 2017. The role of barrier function, autophagy, and cytokines in maintaining intestinal homeostasis. Paper read at Seminars in cell & developmental biology.
- Faith, Jeremiah J, Janaki L Guruge, Mark Charbonneau, Sathish Subramanian, Henning Seedorf, Andrew L Goodman, Jose C Clemente, Rob Knight, Andrew C Heath, and Rudolph L Leibel. 2013. "The long-term stability of the human gut microbiota." *Science* no. 341 (6141).
- Farhadi, Ashkan, ALI Banan, Jeremy Fields, and ALI Keshavarzian. 2003. "Intestinal barrier: an interface between health and disease." *Journal of gastroenterology and hepatology* no. 18 (5):479-497.
- Faust, Karoline, and Jeroen Raes. 2012. "Microbial interactions: from networks to models." *Nature Reviews Microbiology* no. 10 (8):538-550.
- Favier, Christine F, Elaine E Vaughan, Willem M De Vos, and Antoon DL Akkermans. 2002. "Molecular monitoring of succession of bacterial communities in human neonates." *Applied and environmental microbiology* no. 68 (1):219-226.
- Fernandez-Tome, Samuel, Blanca Hernandez-Ledesma, Maria Chaparro, Pedro Indiano-Romacho, David Bernardo, and Javier P Gisbert. 2019. "Role of food proteins and bioactive peptides in

- inflammatory bowel disease." *Trends in Food Science & Technology* no. 88:194-206.
- Frank, Daniel N, Allison L St Amand, Robert A Feldman, Edgar C Boedeker, Noam Harpaz, and Norman R Pace. 2007. "Molecular-phylogenetic characterization of microbial community imbalances in human inflammatory bowel diseases." *Proceedings of the national academy of sciences* no. 104 (34):13780-13785.
- Gamallat, Yaser, Xiaomeng Ren, Abdo Meyiah, Meiqi Li, Xinxiu Ren, Yazeed Jamal, Siyuan Song, Luhan Xie, Bashir Ahmad, and Abdullah Shopit. 2019. "The immune-modulation and gut microbiome structure modification associated with long-term dietary supplementation of *Lactobacillus rhamnosus* using 16S rRNA sequencing analysis." *Journal of Functional Foods* no. 53:227-236.
- Ghoshal, Uday C, Kok-Ann Gwee, Gerald Holtmann, Yanmei Li, Soo Jung Park, Marcellus Simadibrata, Kentaro Sugano, Kaichun Wu, Eamonn MM Quigley, and Henry Cohen. 2018. "The role of the microbiome and the use of probiotics in gastrointestinal disorders in adults in the Asia-Pacific region-background and recommendations of a regional consensus meeting." *Journal of gastroenterology and hepatology* no. 33 (1):57-69.
- Gilmore, Thomas D, TW Siggers, and S Gerondakis. 2015. "NF-kappaB and the Immune System." In *Encyclopedia of Cell Biology*, 580-587. Academic Press.
- Goodman, Andrew L, George Kallstrom, Jeremiah J Faith, Alejandro Reyes, Aimee Moore, Gautam Dantas, and Jeffrey I Gordon. 2011. "Extensive personal human gut microbiota culture collections characterized and manipulated in gnotobiotic mice." *Proceedings of the National Academy of Sciences* no. 108 (15):6252-6257.
- Goodrich, Julia K, Jillian L Waters, Angela C Poole, Jessica L Sutter, Omry Koren, Ran Blekhman, Michelle Beaumont, William Van Treuren, Rob Knight, and Jordana T Bell. 2014. "Human genetics shape the gut microbiome." *Cell* no. 159 (4):789-799.
- Goyal, Akshit, Tong Wang, Veronika Dubinkina, and Sergei Maslov. 2021. "Ecology-guided prediction of cross-feeding interactions in the human gut microbiome." *Nature communications* no. 12 (1):1335.
- Greenhalgh, Kacy, Kristen M Meyer, Kjersti M Aagaard, and Paul Wilmes. 2016. "The human gut microbiome in health: establishment and resilience of microbiota over a lifetime." *Environmental microbiology* no. 18 (7):2103-2116.
- Gustafsson, Jenny K, and Malin EV Johansson. 2022. "The role of goblet cells and mucus in intestinal homeostasis." *Nature Reviews Gastroenterology & Hepatology* no. 19 (12):785-803.
- Guzik, T, R Korbut, and T Adamek-Guzik. 2003. "Nitric oxide and superoxide in inflammation." *J physiol pharmacol* no. 54 (4):469-487.
- Hansson, Gunnar C. 2012. "Role of mucus layers in gut infection and inflammation." *Current opinion in microbiology* no. 15 (1):57-62.
- Hartmann, Rainer, and Hans Meisel. 2007. "Food-derived peptides with biological activity: from research to food applications." *Current opinion in biotechnology* no. 18 (2):163-169.

- Hayashi, Hidenori, Rei Takahashi, Takahiro Nishi, Mitsuo Sakamoto, and Yoshimi Benno. 2005. "Molecular analysis of jejunal, ileal, caecal and recto-sigmoidal human colonic microbiota using 16S rRNA gene libraries and terminal restriction fragment length polymorphism." *Journal of medical microbiology* no. 54 (11):1093-1101.
- Hayden, Matthew S, and Sankar Ghosh. 2004. "Signaling to NF- $\kappa$ B." *Genes & development* no. 18 (18):2195-2224.
- Hillman, Ethan T, Hang Lu, Tianming Yao, and Cindy H Nakatsu. 2017. "Microbial ecology along the gastrointestinal tract." *Microbes and environments*:ME17017.
- Hou, Hu, Yan Fan, Shikai Wang, Leilei Si, and Bafang %J Journal of functional foods Li. 2016. "Immunomodulatory activity of Alaska pollock hydrolysates obtained by glutamic acid biosensor–Artificial neural network and the identification of its active central fragment." no. 24:37-47.
- Hu, Qian, Jingyu Shi, Jiao Zhang, Yi Wang, Yuanyuan Guo, and Zhiping Zhang. "Progress and Prospects of Regulatory Functions Mediated by Nitric Oxide on Immunity and Immunotherapy." *Advanced Therapeutics*:2100032.
- Jandhyala, Sai Manasa, Rupjyoti Talukdar, Chivkula Subramanyam, Harish Vuyyuru, Mitnala Sasikala, and D Nageshwar Reddy. 2015. "Role of the normal gut microbiota." *World journal of gastroenterology: WJG* no. 21 (29):8787.
- Johansson, Magnus, David G DeNardo, and Lisa M Coussens. 2008. "Polarized immune responses differentially regulate cancer development." *Immunological reviews* no. 222 (1):145-154.
- Khan, Asif Iqbal, Ata Ur Rehman, Nabeel Ahmed Farooqui, Nimra Zafar Siddiqui, Qamar Ayub, Muhammad Noman Ramzan, Liang Wang, and Yi Xin. 2022. "Effects of shrimp peptide hydrolysate on intestinal microbiota restoration and immune modulation in cyclophosphamide-treated mice." *Molecules* no. 27 (5):1720.
- Khan, Asif Iqbal, Ata Ur Rehman, Nabeel Ahmed Farooqui, Nimra Zafar Siddiqui, Qamar Ayub, Muhammad Noman Ramzan, Wang Zexu, Xiaoxiao Zhang, Yingshuo Yu, and Yi Xin. 2022. "Shrimp peptide hydrolysate modulates the immune response in cyclophosphamide immunosuppressed mice model." *Journal of Food Biochemistry* no. 46 (9):e14251.
- Khudojberdievich, Ziyadullaev Shukhrat. 2022. "THE ROLE OF THE GUT MICROBIOTA IN NUTRITION AND HEALTH." *World Bulletin of Public Health* no. 10:109-114.
- Kiewiet, MBG, M Gros, RJJ van Neerven, MM Faas, and P de Vos. 2015. "Immunomodulating properties of protein hydrolysates for application in cow's milk allergy." *Pediatric Allergy and Immunology* no. 26 (3):206-217.
- Kiewiet, Mensiena B Gea, Betty CAM van Esch, Johan Garssen, Marijke M Faas, and Paul de Vos. 2017. "Partially hydrolyzed whey proteins prevent clinical symptoms in a cow's milk allergy mouse model and enhance regulatory T and B cell frequencies." *Molecular nutrition & food research* no. 61 (11):1700340.
- Kiewiet, Mensiena BG, Marijke M Faas, and Paul De Vos. 2018. "Immunomodulatory protein

- hydrolysates and their application." *Nutrients* no. 10 (7):904.
- Koenderman, Leo, Wim Buurman, and Mohamed R Daha. 2014. "The innate immune response." *Immunology letters* no. 162 (2):95-102.
- Korhonen, Hannu, and Anne Pihlanto. 2006. "Bioactive peptides: production and functionality." *International dairy journal* no. 16 (9):945-960.
- Kurashima, Yosuke, Yoshiyuki Goto, and Hiroshi Kiyono. 2013. "Mucosal innate immune cells regulate both gut homeostasis and intestinal inflammation." *European journal of immunology* no. 43 (12):3108-3115.
- Lamb, Tracey J. 2012. "Notes on the immune system." *Immunity to Parasitic Infection*:13-57.
- Lane, Erin R, Timothy L Zisman, and David L Suskind. 2017. "The microbiota in inflammatory bowel disease: current and therapeutic insights." *Journal of inflammation research* no. 10:63.
- Laveti, Durgaprasad, Manoj Kumar, R Hemalatha, Ramakrishna Sistla, V Gm Naidu, Venu Talla, Vinod Verma, Navrinder Kaur, and Ravinder Nagpal. 2013. "Anti-inflammatory treatments for chronic diseases: a review." *Inflammation & Allergy-Drug Targets (Formerly Current Drug Targets-Inflammation & Allergy)(Discontinued)* no. 12 (5):349-361.
- Lee, I-Ta, and Chuen-Mao Yang. 2012. "Role of NADPH oxidase/ROS in pro-inflammatory mediators-induced airway and pulmonary diseases." *Biochemical pharmacology* no. 84 (5):581-590.
- Lee, Yun Kyung, and Sarkis K Mazmanian. 2010. "Has the microbiota played a critical role in the evolution of the adaptive immune system?" *science* no. 330 (6012):1768-1773.
- Ley, Ruth E, Daniel A Peterson, and Jeffrey I Gordon. 2006. "Ecological and evolutionary forces shaping microbial diversity in the human intestine." *Cell* no. 124 (4):837-848.
- Li, Wei, Shengwang Ye, Zhuangwei Zhang, Jiachao Tang, Huoxi Jin, Fangfang Huang, Zuisu Yang, Yunping Tang, Yan Chen, and Guofang Ding. 2019. "Purification and characterization of a novel pentadecapeptide from protein hydrolysates of *Cyclina sinensis* and its immunomodulatory effects on RAW264. 7 cells." *Marine drugs* no. 17 (1):30.
- Li, Xujiao, Rui Guo, Xuejiao Wu, Xin Liu, Lianzhong Ai, Yi Sheng, Zibo Song, and Yan Wu. 2020. "Dynamic digestion of tamarind seed polysaccharide: Indigestibility in gastrointestinal simulations and gut microbiota changes in vitro." *Carbohydrate polymers* no. 239:116194.
- Liang, Yan, Yang Zhou, and Pingping Shen. 2004. "NF-kappaB and its regulation on the immune system." *Cell Mol Immunol* no. 1 (5):343-350.
- Linares, Daniel M, Paul Ross, and Catherine Stanton. 2016. "Beneficial microbes: the pharmacy in the gut." *Bioengineered* no. 7 (1):11-20.
- Liu, Feng, Lijia Zhang, Xi Feng, Salam A Ibrahim, Wen Huang, and Ying Liu. 2021. "Immunomodulatory Activity of Carboxymethyl Pachymaran on Immunosuppressed Mice Induced by Cyclophosphamide." *Molecules* no. 26 (19):5733.
- Liu, Ting, Lingyun Zhang, Donghyun Joo, and Shao-Cong Sun. 2017. "NF-κB signaling in inflammation." *Signal*

- transduction and targeted therapy* no. 2 (1):1-9.
- Lozano-Ojalvo, Daniel, and Rosina López-Fandiño. 2018. "Immunomodulating peptides for food allergy prevention and treatment." *Critical reviews in food science and nutrition* no. 58 (10):1629-1649.
- Mackie, Roderick I, Abdelghani Sghir, and H Rex Gaskins. 1999. "Developmental microbial ecology of the neonatal gastrointestinal tract." *The American journal of clinical nutrition* no. 69 (5):1035s-1045s.
- Martínez-Medina, Gloria A, Mónica L Chávez-González, J Yajaira Méndez-Carmona, Orlando de la Rosa, Rocío Carranza-Méndez, Dora Elisa Cruz-Casas, Pilar Espitia-Hernández, Daisy P Amaya-Chantaca, and Cristobal N Aguilar. 2022. "Immunomodulatory Properties of Proteins and Peptides: Food Derivatives Approach." In *Immunomodulators and Human Health*, 415-438. Springer.
- Maurice, Corinne Ferrier, Henry Joseph Haiser, and Peter James Turnbaugh. 2013. "Xenobiotics shape the physiology and gene expression of the active human gut microbiome." *Cell* no. 152 (1-2):39-50.
- Medzhitov, Ruslan, and Charles A Janeway Jr. 1997. "Innate immunity: impact on the adaptive immune response." *Current opinion in immunology* no. 9 (1):4-9.
- Meulenbroek, Laura APM, Betty CAM van Esch, Gerard A Hofman, Constance F den Hartog Jager, Alma J Nauta, Linette EM Willemsen, Carla AFM Bruijnzeel-Koomen, Johan Garssen, Els van Hoffen, and Léon MJ Knippels. 2013. "Oral treatment with  $\beta$ -lactoglobulin peptides prevents clinical symptoms in a mouse model for cow's milk allergy." *Pediatric Allergy and Immunology* no. 24 (7):656-664.
- Mitchell, Simon, Jesse Vargas, and Alexander Hoffmann. 2016. "Signaling via the NF $\kappa$ B system." *Wiley Interdisciplinary Reviews: Systems Biology and Medicine* no. 8 (3):227-241.
- Nam, Young-Do, Mi-Ja Jung, Seong Woon Roh, Min-Soo Kim, and Jin-Woo Bae. 2011. "Comparative analysis of Korean human gut microbiota by barcoded pyrosequencing." *PloS one* no. 6 (7):e22109.
- Nelson, Ralph, Shigeru Katayama, Yoshimuri Mine, Jairo Duarte, and Chantal Matar. 2007. "Immunomodulating effects of egg yolk low lipid peptic digests in a murine model." *Food and Agricultural Immunology* no. 18 (1):1-15.
- Nishida, Atsushi, Ryo Inoue, Osamu Inatomi, Shigeki Bamba, Yuji Naito, and Akira Andoh. 2018. "Gut microbiota in the pathogenesis of inflammatory bowel disease." *Clinical journal of gastroenterology* no. 11 (1):1-10.
- Ogra, Pearay L, Jiri Mestecky, Michael E Lamm, Warren Strober, Jerry R McGhee, and John Bienenstock. 2012. *Handbook of mucosal immunology*: Academic Press.
- Pabst, O, and AM Mowat. 2012. "Oral tolerance to food protein." *Mucosal immunology* no. 5 (3):232-239.
- Palmer, Chana, Elisabeth M Bik, Daniel B DiGiulio, David A Relman, and Patrick O Brown. 2007. "Development of the human infant intestinal microbiota." *PLoS biology* no. 5 (7):e177.
- Paul, William E. 2013. *Fundamental Immunology*. (William E. Paul, ed.). United States: Wolters Kluwer Health/Lippincott Williams & Wilkins.

- Pereira, Fátima C, and David Berry. 2017. "Microbial nutrient niches in the gut." *Environmental microbiology* no. 19 (4):1366-1378.
- Perez-Lopez, Araceli, Judith Behnsen, Sean-Paul Nuccio, and Manuela Raffatellu. 2016. "Mucosal immunity to pathogenic intestinal bacteria." *Nature Reviews Immunology* no. 16 (3):135-148.
- Phalipon, Armelle, Ana Cardona, Jean-Pierre Kraehenbuhl, Léna Edelman, Philippe J Sansonetti, and Blaise Corthésy. 2002. "Secretory component: a new role in secretory IgA-mediated immune exclusion in vivo." *Immunity* no. 17 (1):107-115.
- Plaisancié, Pascale, Jean Claustre, Monique Estienne, Gwenaele Henry, Rachel Boutrou, Armelle Paquet, and Joelle Léonil. 2013. "A novel bioactive peptide from yoghurts modulates expression of the gel-forming MUC2 mucin as well as population of goblet cells and Paneth cells along the small intestine." *The Journal of nutritional biochemistry* no. 24 (1):213-221.
- Qin, Junjie, Ruiqiang Li, Jeroen Raes, Manimozhayan Arumugam, Kristoffer Solvsten Burgdorf, Chaysavanh Manichanh, Trine Nielsen, Nicolas Pons, Florence Levenez, and Takuji Yamada. 2010. "A human gut microbial gene catalogue established by metagenomic sequencing." *nature* no. 464 (7285):59-65.
- Rajilić-Stojanović, Mirjana, Hans GHJ Heilig, Douwe Molenaar, Kajsa Kajander, Anu Surakka, Hauke Smidt, and Willem M De Vos. 2009. "Development and application of the human intestinal tract chip, a phylogenetic microarray: analysis of universally conserved phylotypes in the abundant microbiota of young and elderly adults." *Environmental microbiology* no. 11 (7):1736-1751.
- Ravel, Jacques, Pawel Gajer, Zaid Abdo, G Maria Schneider, Sara SK Koenig, Stacey L McCulle, Shara Karlebach, Reshma Gorle, Jennifer Russell, and Carol O Tacket. 2011. "Vaginal microbiome of reproductive-age women." *Proceedings of the National Academy of Sciences* no. 108 (Supplement 1):4680-4687.
- Rescigno, Maria. 2011. "Dendritic cells in oral tolerance in the gut." *Cellular microbiology* no. 13 (9):1312-1318.
- Roberfroid, Marcel. 2007. "Prebiotics: the concept revisited." *The Journal of nutrition* no. 137 (3):830S-837S.
- Robinson, Courtney J, Brendan JM Bohannon, and Vincent B Young. 2010. "From structure to function: the ecology of host-associated microbial communities." *Microbiology and Molecular Biology Reviews* no. 74 (3):453-476.
- Rooks, Michelle G, and Wendy S Garrett. 2016. "Gut microbiota, metabolites and host immunity." *Nature reviews immunology* no. 16 (6):341-352.
- Santoro, Aurelia, Rita Ostan, Marco Candela, Elena Biagi, Patrizia Brigidi, Miriam Capri, and Claudio Franceschi. 2018. "Gut microbiota changes in the extreme decades of human life: a focus on centenarians." *Cellular and Molecular Life Sciences* no. 75:129-148.
- Scanlan, Pauline D, and Julian R Marchesi. 2008. "Micro-eukaryotic diversity of the human distal gut microbiota: qualitative assessment using culture-dependent and-independent analysis of faeces." *The ISME journal* no. 2 (12):1183-1193.

- Scanlan, Pauline D, Fergus Shanahan, Caitlin O'Mahony, and Julian R Marchesi. 2006. "Culture-independent analyses of temporal variation of the dominant fecal microbiota and targeted bacterial subgroups in Crohn's disease." *Journal of clinical microbiology* no. 44 (11):3980-3988.
- Sessa, William C. 1994. "The nitric oxide synthase family of proteins." *Journal of vascular research* no. 31 (3):131-143.
- Shi, Na, Na Li, Xinwang Duan, and Haitao Niu. 2017. "Interaction between the gut microbiome and mucosal immune system." *Military Medical Research* no. 4 (1):1-7.
- Shimizu, Makoto. 2004. "Food-derived peptides and intestinal functions." *Biofactors* no. 21 (1-4):43-47.
- Sommer, Felix, and Fredrik Bäckhed. 2013. "The gut microbiota—masters of host development and physiology." *Nature reviews microbiology* no. 11 (4):227-238.
- Sommer, Felix, and Fredrik Bäckhed. 2015. "The gut microbiota engages different signaling pathways to induce Duox2 expression in the ileum and colon epithelium." *Mucosal immunology* no. 8 (2):372-379.
- Strober, Warren, Ivan J Fuss, and Richard S Blumberg. 2002. "The immunology of mucosal models of inflammation." *Annual review of immunology* no. 20 (1):495-549.
- Talapko, Jasminka, Aleksandar Včev, Tomislav Meštrović, Emina Pustijanac, Melita Jukić, and Ivana Škrlec. 2022. "Homeostasis and Dysbiosis of the Intestinal Microbiota: Comparing Hallmarks of a Healthy State with Changes in Inflammatory Bowel Disease." *Microorganisms* no. 10 (12):2405.
- Tao, Jin-Hua, Jin-Ao Duan, Wei Zhang, Shu Jiang, Jian-Ming Guo, and Dan-Dan Wei. 2018. "Polysaccharides from *Chrysanthemum morifolium* ramat ameliorate colitis rats via regulation of the metabolic profiling and NF- $\kappa$ B/TLR4 and IL-6/JAK2/STAT3 signaling pathways." *Frontiers in pharmacology* no. 9:746.
- Thangaleela, Subramanian, Bhagavathi Sundaram Sivamaruthi, Periyanaina Kesika, Muruganantham Bharathi, and Chaiyavat Chaiyasut. 2022. "Nasal Microbiota, Olfactory Health, Neurological Disorders and Aging—A Review." *Microorganisms* no. 10 (7):1405.
- Tilg, Herbert, and Arthur Kaser. 2011. "Gut microbiome, obesity, and metabolic dysfunction." *The Journal of clinical investigation* no. 121 (6):2126-2132.
- Tosi, Michael F. 2005. "Innate immune responses to infection." *Journal of Allergy and Clinical Immunology* no. 116 (2):241-249.
- Tulstrup, Monica Vera-Lise, Ellen Gerd Christensen, Vera Carvalho, Caroline Linninge, Siv Ahrné, Ole Højberg, Tine Rask Licht, and Martin Iain Bahl. 2015. "Antibiotic treatment affects intestinal permeability and gut microbial composition in Wistar rats dependent on antibiotic class." *PloS one* no. 10 (12):e0144854.
- Vannini, Federica, Khosrow Kashfi, and Niharika Nath. 2015. "The dual role of iNOS in cancer." *Redox biology* no. 6:334-343.
- Vaziri, Nosratola D, Jun Yuan, and Keith Norris. 2013. "Role of urea in intestinal barrier dysfunction and disruption of epithelial tight junction in chronic

- kidney disease." *American journal of nephrology* no. 37 (1):1-6.
- Vickers, Neil J. 2017. "Animal communication: when i'm calling you, will you answer too?" *Current biology* no. 27 (14):R713-R715.
- Viggiano, D, G Ianiro, G Vanella, S Bibbò, G Bruno, G Simeone, and G Mele. 2015. "Gut barrier in health and disease: focus on childhood." *Eur Rev Med Pharmacol Sci* no. 19 (6):1077-85.
- Vivijis, Bram, Abram Aertsen, and Chris W Michiels. 2016. "Identification of genes required for growth of *Escherichia coli* MG1655 at moderately low pH." *Frontiers in microbiology* no. 7:1672.
- Wang, Han, Lei Xu, Mingming Yu, Yuanhong Wang, Tingfu Jiang, Shuang Yang, and Zhihua Lv. 2019. "Glycosaminoglycan from *Apostichopus japonicus* induces immunomodulatory activity in cyclophosphamide-treated mice and in macrophages." *International journal of biological macromolecules* no. 130:229-237.
- Wang, X, SP Heazlewood, DO Krause, and THJ Florin. 2003. "Molecular characterization of the microbial species that colonize human ileal and colonic mucosa by using 16S rDNA sequence analysis." *Journal of applied microbiology* no. 95 (3):508-520.
- Wang, Zeneng, Elizabeth Klipfell, Brian J Bennett, Robert Koeth, Bruce S Levison, Brandon DuGar, Ariel E Feldstein, Earl B Britt, Xiaoming Fu, and Yoon-Mi Chung. 2011. "Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease." *Nature* no. 472 (7341):57-63.
- Wells, Jerry M, Oriana Rossi, Marjolein Meijerink, and Peter van Baarlen. 2011. "Epithelial crosstalk at the microbiota-mucosal interface." *Proceedings of the national academy of sciences* no. 108 (Supplement 1):4607-4614.
- Williams, AE. 2011a. "Basic concepts in immunology." *Immunology: mucosal and body surface defences*. Chichester, UK: John Wiley & Sons, Ltd:1-19.
- Williams, AE. 2011b. "The innate immune system." *Immunology: mucosal and body surface defences*. Chichester, UK: John Wiley & Sons, Ltd:20-40.
- Wu, Gary D, Jun Chen, Christian Hoffmann, Kyle Bittinger, Ying-Yu Chen, Sue A Keilbaugh, Meenakshi Bewtra, Dan Knights, William A Walters, and Rob Knight. 2011. "Linking long-term dietary patterns with gut microbial enterotypes." *Science* no. 334 (6052):105-108.
- Xiang, Xing-Wei, Hui-Zhen Zheng, Rui Wang, Hui Chen, Jin-Xing Xiao, Bin Zheng, Shu-Lai Liu, and Yu-Ting Ding. 2021. "Ameliorative Effects of Peptides Derived from Oyster (*Crassostrea gigas*) on Immunomodulatory Function and Gut Microbiota Structure in Cyclophosphamide-Treated Mice." *Marine Drugs* no. 19 (8):456.
- Xu, Xiaofei, and Xuewu Zhang. 2015. "Effects of cyclophosphamide on immune system and gut microbiota in mice." *Microbiological research* no. 171:97-106.
- Yimit, Dilshat, Parida Hoxur, Nurmuhmat Amat, Kimono Uchikawa, and Nobuo Yamaguchi. 2012. "Effects of soybean peptide on immune function, brain function, and neurochemistry in healthy volunteers." *Nutrition* no. 28 (2):154-159.
- Ying, Mengxi, Bing Zheng, Qiang Yu, Kunyou Hou, Hui Wang, Mingming Zhao, Yi Chen, Jianhua Xie, Shaoping Nie, and Mingyong Xie. 2020. "Ganoderma

- atrum polysaccharide ameliorates intestinal mucosal dysfunction associated with autophagy in immunosuppressed mice." *Food and Chemical Toxicology* no. 138:111244.
- Zhang, Lirong, Wanxiu Cao, Yuan Gao, Ruili Yang, Xu Zhang, Jie Xu, and Qingjuan Tang. 2020. "Astaxanthin (ATX) enhances the intestinal mucosal functions in immunodeficient mice." *Food & function* no. 11 (4):3371-3381.
- Zhang, Zhuangwei, Xuyang Hu, Lin Lin, Guofang Ding, and Fangmiao Yu. 2019. "Immunomodulatory activity of low molecular-weight peptides from *Nibea japonica* in RAW264. 7 cells via NF- $\kappa$ B pathway." *Marine drugs* no. 17 (7):404.
- Zhao, Ya, Yamei Yan, Wangting Zhou, Dan Chen, Kaiyin Huang, Shijie Yu, Jia Mi, Lu Lu, Xiaoxiong Zeng, and Youlong Cao. 2020. "Effects of polysaccharides from bee collected pollen of Chinese wolfberry on immune response and gut microbiota composition in cyclophosphamide-treated mice." *Journal of Functional Foods* no. 72:104057.
- Zhu, Yuanmin, T Michelle Luo, Christian Jobin, and Howard A Young. 2011. "Gut microbiota and probiotics in colon tumorigenesis." *Cancer letters* no. 309 (2):119-127.
- Zwielehner, Jutta, Cornelia Lassl, Berit Hippe, Angelika Pointner, Olivier J Switzeny, Marlene Remely, Elvira Kitzweger, Reinhard Ruckser, and Alexander G Haslberger. 2011. "Changes in human fecal microbiota due to chemotherapy analyzed by TaqMan-PCR, 454 sequencing and PCR-DGGE fingerprinting." *PloS one* no. 6 (12):e28654.