

DOI: [10.55627/ppc.005.01.01321](https://doi.org/10.55627/ppc.005.01.01321)**Research Article****Effects of Phytochemicals in Attenuating Melanoma: A Comprehensive Literature Review**

Muhammad Mutayyab Javaid, Ahsan Ibrahim, Muhammad Haad Habib

Shifa College of Pharmaceutical Sciences, Shifa Tameer-e-Millat University, H-8, Islamabad, Pakistan

*Correspondence: ahsen.scps@stmu.edu.pk

© The Author(s) 2025. This article is licensed under a Creative Commons Attribution 4.0 International License. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

Abstract

Melanoma is an aggressive form of skin cancer associated with a low survival rate, high mortality, poor quality of life, and incidence that has risen significantly in the last few decades. While surgery, radiation, and chemotherapy drugs are available as treatment choices, melanoma still has an unfavorable clinical outcome. This can be related to the metastatic potential and heterogeneity of melanoma and to the multitude of adverse effects (AEs) that are caused by chemotherapy, the most common of which are alopecia, nausea, changes in taste and appetite, fatigue, vomiting, dry mouth, and constipation. The recent decade has seen a rise in the trend of studies using plant-derived phytochemicals in vivo and in vitro studies, which have demonstrated anti-tumor activity along with the added benefit of significantly lower AEs. These secondary metabolites of plants like chlorogenic acid, rosmarinic acid, palmatine, berberine, quercetin, myricadiol, and other phytochemicals found in natural sources like *Marrubium lutescens*, *Berberis pruinosa*, *Lansea coromandelica*, *Orbea variegata*, and *Galium verum*, among others, are now gaining considerable importance for use against melanoma. Related studies were collected through a literature review and are discussed in this article.

Keywords: Melanoma, phytochemicals, chlorogenic acid, rosmarinic acid, palmatine, berberine, quercetin, myricadiol

1. Introduction

Melanoma is the malignant neoplasm of melanocytes (Ali Khani et al. 2023). These are cells that secrete the pigment melanin, which gives the skin its color and protects it from the deteriorating effects of ultraviolet (UV) radiation, which is important as UV exposure is thought to be a major cause of skin cancer in many cases (Rashid, Shaughnessy, and Tsao 2023). Epidemiological data taken from published literature depict the global burden of melanoma, emphasizing its rising incidence and high mortality relative to other skin cancers. Melanoma has been responsible for roughly 57,000 deaths in 2020, accounting for 80% of all skin cancer deaths. In Europe, roughly 150,000 patients get diagnosed with melanoma each year, and roughly 25,000 of

them die of it (Nurla and Forsea 2024). In the US alone, roughly 106,000 new cases were seen in 2021, making it the 5th leading cancer diagnosis of that year. The US has seen a 320% rise in melanoma since 1975. The 5-year survival seen from 2011 to 2017 in the US was 30% for stage 4, 68% for stage 3, and 99.4% for stage 1 or 2 melanoma (Saginala et al. 2021). This high mortality and limited treatment response can be attributed to factors like metastatic potential and heterogeneity of the tumor (Ng, Simmons, and Boyle 2022).

The current management of melanoma involves the surgical excision of the primary tumor, and in invasive cases, sentinel lymph node biopsy for proper staging of the disease (Garbe et al. 2022). If the disease is resected and classified as high risk

(stage IIB-IV), adjuvant systemic therapy will be offered, which includes anti-PD-1 immunotherapy (nivolumab or pembrolizumab) or targeted therapy (BRAF/MEK inhibitors dabrafenib plus trametinib) for BRAF V600 tumors (Marquez-Rodas et al. 2024; Eggermont et al. 2022). Recently, in resectable clinical stage IIIB-IV melanoma, neoadjuvant systemic therapy was recommended with immune therapy (pembrolizumab or nivolumab/ipilimumab) before surgical intervention (Swetter et al. 2021; Blank et al. 2024). In the case of unresectable or metastatic disease, the first line systemic therapy will incorporate strong immune therapies, either anti-PD-1 as a monotherapy, or anti-PD-1 plus anti-CTLA-4 or anti-LAG-3 combination therapy as the initial first line management (Garbe et al. 2025). Targeted therapy with BRAF/MEK inhibitors is pursued as either a primary first line option or a subsequent option for BRAF V600 mutant disease (Haist et al. 2022). In melanoma, multiple drugs such as ipilimumab (anti-CTLA-4), nivolumab (anti-PD-1), Tebentafusp (target gp100), Dabrafenib (target BRAF), Trametinib (target MEK1 and MEK2), Relatlimab (target LAG-3) are used which are mentioned in standard guidelines (Almoussalam and Zhu 2016; Faghfuri et al. 2015; Staeger et al. 2025; Hoffner and Benchich 2018). However, such options give low response rates and median overall survival especially in advanced stages of the disease as observed in published literature (Gaughan and Horton 2022; Galus et al. 2025; Goldinger et al. 2022). Many chemotherapeutic drugs such as Temozolomide and Dacarbazine are limited by their poor physicochemical and pharmacokinetic properties such as low stability, low water solubility and rapid metabolism (Lopes et al. 2022; Ahad et al. 2022; Almoussalam and Zhu 2016). These drugs in general cause a multitude of adverse effects (AEs) which in most cases are alopecia, constipation, changes in taste and appetite, fatigue, vomiting, and nausea but also include other AEs specific to the drug or

combination used (Seth et al. 2020; Boutros et al. 2024; Altun and Sonkaya 2018).

Phytochemicals, which are secondary metabolites obtained from plant material, have proven anti-cancer, anti-proliferative, and anti-inflammatory properties and have shown great potential in reducing the progression of skin cancer (Penta et al. 2018). Phytochemicals are gaining considerable importance for use against melanoma, which is evident from the rise in number of research articles in recent years (Tabolacci et al. 2023). The benefit of phytochemicals is that they have significantly lower AEs when compared with conventional chemotherapy. Herbal medicine, however, can also cause AEs especially when they are not prepared for the standards of allopathic medicine. Furthermore, phytochemicals, due to their varied lipophilic and hydrophilic properties, contribute to rapid degradation, low bioavailability, metabolic instability, and insufficient absorption, which ultimately lowers therapeutic efficacy. In addition, limited mechanistic understanding restricts patient stratification, and limited preclinical and clinical testing restricts the translation of phytochemicals into effective therapies for humans (Kim et al. 2025). The primary objective of this review is to explore the anti-neoplastic potential of such biological constituents for the purpose of managing skin cancer. In pursuit of this objective, a comprehensive review of the existing literature was conducted to evaluate the effects of phytochemicals on melanoma in vitro and in vivo. Furthermore, the review provides a discussion on the etiological and pathophysiological insights of melanoma, evaluates the limitations of current therapeutic options, and presents the selected studies in a tabulated format.

2. Classification, Etiology, and Pathophysiological Insights of Melanoma

2.1. Classification

WHO classifies melanoma on the basis of sun damage with emphasis on genetic hallmarks and divides melanoma into nine distinct groups,

which include low cumulative sun damage (CSD) melanoma, high-CSD melanoma, desmoplastic melanoma, melanoma arising in congenital nevus, malignant Spitz tumor, acral melanoma, nodular melanoma, mucosal melanoma, which develops in blue nevus, and uveal melanoma (Ferrara and Argenziano 2021). Genetic mutations identified in melanoma include those in *BRAF*, *NF1* and *NRAS* (Samlowski 2024), *NFKBIE* promoter mutations, activating mutations in the *MAPK* pathway like *MAP2K1* (Zob et al. 2022), kinase fusions in *NTRK3*, *NTRK1*, *RET*, *MET*, *ALK* and *ROS1* (Olmedo et al. 2022), *MAP3K8* truncating/fusions mutation (Sibira et al. 2024), *CDKN2A* homozygous deletion (Kreuger et al. 2023), *TERT* promoter mutation, *TERT*, *KIT* and *CCND1* amplification (Broit et al. 2022), *EIF1AX*, *SF3B1* and *BAP1* mutations (Skrehot, Alsoudi, and Schefler 2025), mutations initiating in the *Gαq* signaling pathway like *PLCB4*, *GNA11*, *CYSLTR2* and *GNAQ* (Akin-Bali 2021) and chromosome 8q gain (Delgado-Bellido et al. 2025).

Melanoma is usually seen on the skin but is found on mucosal surfaces like the uveal tract of the eye, oral cavity, upper gastrointestinal (GI) tract, leptomeninges, and genital mucosa. Melanoma that spreads to the skin, lymphatic vessels, or lymph nodes is referred to as regional metastasis. It includes satellite metastases that are seen within 2 cm of the original tumor, in-transit metastasis showing spread of tumor to lymphatic vessels or blood capillaries, and nodal metastasis showing the invasion of tumor to the lymph nodes (Zbytek et al. 2008). Cutaneous melanomas are grouped in accordance with their growth patterns. Superficial spreading melanoma, seen in 57% of patients, is the most prevalent. Within the epidermis, melanocytes with pagetoid spread are confined in an irregular pattern. This is seen on the skin as a network of brown dots having pseudopods with a blue/red coloration or depigmentation. Nodular melanoma is found in 21% of patients. It is seen as individual grey/blue colored globules containing asymmetric vessels and white streaks. Lentigo maligna melanoma is found in 9% of cases. It

contains irregular follicular hyperpigmented openings outsized by asymmetric colored dots. Elastosis and epidermal atrophy are observed on the skin due to the growth of individual tumor cells within the adnexal structures of epithelium and epidermis. The least common acral lentiginous melanoma is observed in 4% of individuals. Multiple hypopigmented regions are seen in brown to grey colored asymmetric polygonal shapes. Melanocytes are seen alone or in groups spreading within the basal layer of hyperplastic dermis (Caraban et al. 2024). Non-cutaneous melanomas are less prevalent and can be present in areas harboring melanocytes. These include uveal melanoma, which is seen in the choroid, ciliary body, or iris of the eye's uvea, and mucosal melanoma seen in the mucosal surfaces of the nasal cavity, oral cavity, sinuses, and genitals (Vergara et al. 2022; Kibbi, Kluger, and Choi 2016). Mucosal melanoma is grouped into genomic subtypes such as *NF1*, *NRAS*, and *BRAF*, which arise from benign lesions after mutations such as loss of *CDKN2A*, *PTEN*, and *TP53*. In contrast, *BRAF*, *SPRED1*, *NF1*, *KIT*, *SF3B1*, and *NRAS* mutations are observed in uveal melanoma (Ma et al. 2021).

2.2. Etiology

Numerous risk factors contribute to the development of melanoma. Studies have revealed that excessive and prolonged exposure to sunlight or indoor tanning leads to inflammation, oxidative stress, and DNA damage in the skin. People with a family history of melanoma, those with lighter skin tones (which indicate lower levels of melanin), and those who frequently suntan are at much greater risk of UV skin damage and melanoma (Tang et al. 2024). Melanoma is prevalent in fair-skinned individuals due to the interplay of *MC1R* signaling and cysteine, which regulate the production of dark brown eumelanin and red to orange pheomelanin, two forms of melanin produced by melanocytes (Flanagan et al. 2000). Pheomelanin induces oxidative stress via the production of reactive oxygen species (ROS) and depletion of antioxidants (Morgan, Lo, and

Fisher 2013). The presence of an unusual number of dysplastic nevi has been associated with melanoma. In nevus-associated melanomas, a higher nevus count and thickness of sporadic dysplastic nevi are linked to age and intermittent exposure to UV radiation (Dessinioti et al. 2022).

2.3. Pathophysiology

Our current understanding of genetic predisposition and molecular basis has helped us identify relevant genes responsible for melanoma, such as RB1, CDK4/6, and CDKN2A encoding for two proteins, p14/ARF and p16/INK4a (Nelson and Tsao 2009). Histologic changes in melanoma progression and their link to specific genetic mutations, showing their impact on molecular signalling, can be seen using the Clark model (Scolyer, Long, and Thompson 2011). This model offers a structure for understanding the multi-stage development of non-chronic sun-damaged melanomas primarily found in Caucasian populations, from a benign lesion to invasive cancer. Using this model, specific histopathological stages are correlated to the described sequential genetic and molecular changes, focusing on pathways of importance that could become therapeutic targets, including phytochemical approaches.

2.3.1. Formation of Benign Nevus

Normal melanocytes in the skin proliferate, forming benign nevi. These are dark brownish, elevated or flat lesions in which the color is either uniformly distributed or present in the form of pigmented dots. Histological examination shows multiple nested melanocytes in the basal layer of these lesions (Miller and Mihm Jr 2006). This phase generally starts with the acquisition of an oncogenic driver mutation, typically a BRAF V600E, which is an important first step to melanocytic neoplasia. However, continued BRAF V600E expression in normal melanocytes induces oncogene-induced senescence, a cell-cycle arrest mechanism. This is associated with the induction of p16^{INK4a} and acidic β -galactosidase activity that helps to maintain tumor size and its benign behavior (Yu et al. 2009; Mackiewicz-Wysocka et

al. 2017). Most melanocytic nevi resulting from BRAF V600 mutations do not progress to melanoma. This is attributed to tumor-suppressive mechanisms, most notably, oncogene-induced senescence, which in melanocytic nevi leads to growth arrest and suppression of tumor (Damsky and Bosenberg 2017).

2.3.2. Development of Dysplastic Nevi

Next begins the abnormal development, plus a benign nevus at a prior or new area. These nevi develop into thicker, multicolored lesions with irregular or asymmetric borders. Irregular, sporadic cytologic atypia are seen in histological studies. At this phase, the melanocyte must move out of senescence (Goldstein and Tucker 2013). The main molecular abnormality that allows the cell to escape senescence is a change or inactivation of the p16^{INK4a}-retinoblastoma (Rb) pathway. This change frequently includes the inactivation of the CDKN2A gene, which encodes p16^{INK4a} and p19^{ARF}. Evidence for the mutation includes frequent loss of heterozygosity at the CDKN2A locus or rare mutations in the CDKN2A gene in sporadic dysplastic nevus (Gray-Schopfer et al. 2006; Shain et al. 2015).

2.3.3. Radial-Growth Phase

Following intraepidermal proliferation of cells, the lesions in the radial-growth phase (RGP) become raised, and continuous atypia is presented as cytomorphic cancer. Tumor cells can further proliferate either alone or in a group into the papillary dermis (Paluncic et al. 2016). To transition into this stage, the progressively proliferating melanocytes must become immortalized to escape replicative senescence, which is conferred by telomerase reverse transcriptase (TERT) activation. It is presumed that external factors (likely keratinocyte products) are required for RGP cells to survive, and thus their growth is limited to the upper layers of skin, adjacent to the epidermis (Takata, Murata, and Saida 2010). Aberrant TERT activation can often be attributed to TERT promoter mutations that generate binding motifs for transcription factors

Table 1: Comparison of efficacy outcomes, toxicity profiles, and specific biomarker requirements across targeted therapies, immune checkpoint inhibitors, and bispecific molecules used to treat different subtypes of advanced melanoma.

Drug	Target/Class	Specific Biomarker	Efficacy	Major Toxicities	Key Limitations	Ref.
Ipilimumab Monotherapy	Fully human mAb blocking CTLA-4	None required for enrollment; studies ongoing to identify predictive biomarkers.	Phase II study (CA184-008). ORR: 5.8%. Median OS: 10.2 months. 2-year survival rate: 32.8%.	AEs were largely immune related, primarily in the skin and GI tract. 19% grade 3 and 3.2% grade AEs.	Response kinetics differ from chemotherapy, requiring specific response criteria. No pre-treatment marker exists to predict response.	(O'Da y et al. 2010)
Ribociclib + Binimetinib	Ribociclib (selective CDK4/6 inhibitor) + Binimetinib (selective MEK inhibitor)	NRAS-mutant melanoma. Enhanced efficacy noted with concurrent alterations in CDKN2A, CDK4, or CCND1.	Phase I-b/II study. ORR: 19.5%. ORR with cell-cycle alterations: 32.5%. Median PFS: 3.7 months. Median OS: 11.3 months.	Common treatment-related toxicities included creatine phosphokinase elevation, rash, edema, anemia, nausea, diarrhea, and fatigue. Increased CPK, AST, and ALT elevations were the most common Grade 3/4 AEs.	Genomic analyses were descriptive and exploratory. The recommended Phase II dose; Ribociclib exposure was much lower than the label monotherapy dose.	(Schuler et al. 2022)
Tebentafusp	First-in-class T-cell receptor-bispecific molecule. Targets glycoprotein 100 (gp100) and CD3.	HLA-A*02:01-positive. Patients must have previously untreated unresectable or metastatic uveal melanoma	Phase 3 trial. Median OS: 21.6 months (vs. 16.9 months for control). 3-year survival Rate: 27% (vs. 18% for control). ORR: 11% (vs. 5% for control). Median PFS: 3.4 months (vs. 2.9 months for control).	Overall incidence of any grade AEs: 99%. Grade 3-4 AEs incidence: 50%. High incidence of Cytokine Release Syndrome (CRS) (80% any grade). Low discontinuation rate due to AEs (1%).	Most common AEs (any grade): rash (83%), pyrexia (76%), pruritus (70%), and hypotension (38%). Cytokine Release Syndrome (CRS) occurred in 89% (mostly Grade 1/2). Grade 3/4 AEs occurred in 47% of patients, commonly rash (19%) and elevated AST (6%). Low discontinuation rate due to AEs (2%).	(Hasse l et al. 2023)
Pembrolizumab Monotherapy	Anti-PD-1 agent (Immune Checkpoint Inhibitor).	Non-uveal, unresectable stage III/IV melanoma.	Cohort study. Median OS (1st line): 32.6 months. ORR (1st line): 49%. Median real-world PFS (1st line): 4.7 months.	Toxicities were not examined in this specific real-world study.	Effectiveness appears lower in later lines (median OS 14.4 months for 2nd line, 9.3 months for 3rd line).	(Casar otto et al. 2021)
Nivolumab Monotherapy	Anti-PD-1 immune checkpoint inhibitor.	High Neutrophil-to-Lymphocyte Ratio and presence of liver metastases predicted poor OS outcome in this study.	Real-World Retrospective study in first line metastatic melanoma. Median PFS: 26 months. Median OS: 31 months. 2-year OS: 62.5%.	Most common AEs were dermatological (23.5%) and endocrinological (15.6%). Grade 3 AEs occurred in 6.8% of patients.	Retrospective design, small patient cohort (N=51), and selection bias. Potential for incomplete data or underreporting of adverse events.	(Afräs ânie et al. 2023)
Dabrafenib + Trametinib	Dabrafenib (BRAF inhibitor) + Trametinib (MEK inhibitor).	BRAF V600-mutant metastatic malignant melanoma (specifically BRAF V600E).	Case reported achieved long-term Complete Metabolic Remission (CR) after 7 months. Contextual trial data: 5-year OS: 28%; ORR: 76%.	Combination therapy is generally well tolerated. Common AEs reported include diarrhea, anorexia, nausea, and vomiting. Case reported modest Grade 1-2 cutaneous	Acquired resistance is common, evidenced by a strong decline in mean survival curves after about 6 months of	(Brugn ara et al. 2018)

					and mucosal side effects.	treatment. Lack of clear guidelines on safely discontinuing treatment after CR.
Dabrafenib Monotherapy	BRAF inhibitor targeting the MAPK pathway.	BRAF mutation.	V600E	Case achieved maintenance of radiological response (stable disease/partial reduction of lesions) for 7.1 months. Patients survived for 15 months.	Minimal toxicity, including beard alopecia and milium cysts.	Patients with the BRAF V600 mutation have been systematically excluded from most pivotal trials. Clinical evidence is primarily limited to case reports and small series. (Casad evall et al. 2016)

Abbreviations: OS (Overall Survival), ORR (Objective Response Rate), PFS (Progression-free Survival).

that enhance TERT transcriptional activity (Heidenreich et al. 2014). Invasion into the deeper layers is primarily restricted due to the regulatory mechanisms involving cell-to-cell adhesion molecules and integrity of the basement membrane (Ciarletta, Foret, and Ben Amar 2011).

2.3.4. Vertical-Growth Phase

Moving to the vertical-growth phase, these lesions proliferate into the papillary dermis, forming an expanded nodule. Tumor cells may extend lower into the reticular dermis and fat region. Further tumor progression requires additional genetic or epigenetic hits that suppress apoptosis, allowing the cells to survive independently of keratinocytes. The primary genetic events seen at this stage include the loss of *PTEN* (which inhibits apoptosis via AKT activation) and RAS activation (Parkman et al. 2025). Phenotypically, progression is marked by the loss of E-cadherin and the aberrant expression of α V β 3 integrin and N-cadherin (Arias-Mejias et al. 2020). The diminished expression of microphthalmia-associated transcription factor (*MITF*) causes the cells to switch from the differentiated, melanocytic phenotype to a less differentiated, motile state (Huang et al. 2021). Increased activity of matrix metalloproteinases (MMPs) leads to the degradation of the extracellular matrix, allowing deeper penetration into the dermal tissue (Napoli et al. 2020). Tumor cells at this stage can be used for tumor studies in nude mouse models. The reduction of TRPM1, increased expression of MMP-2, α V β 3 integrin, and N-cadherin, and loss

of E-cadherin can be seen during both the vertical-growth phase and metastatic melanoma phase (Wagstaff et al. 2022).

2.3.5. Metastatic Melanoma

Melanoma spreads to regional in-transit lesions, lymph nodes, and distant organs. Chemotaxis and lymph flow guide the movement of melanoma cells away from their initial location. After settling, melanoma cells develop a variety of methods to defend themselves against destruction by the immune defense through strategies such as by retaining MHC class 1 polypeptide-related sequence A (*MICA*) intracellularly to prevent attack by natural killer (NK) cells, and by modifying oncostatin M receptor (*OSMR*) to resist the antiproliferative effects of cytokines (Zbytek et al. 2008; Miller and Mihm Jr 2006). Molecular markers typically observed in metastases, but not in primary tumors, are loss-of-function *CDKN2A* mutations, increased TERT promoter mutations, and genetic or epigenetic alterations targeting the *PTEN* tumor suppressor or copy number gains/overexpression of *AKT3* that activate the PI3K/AKT pathway in a constitutive manner (Guo et al. 2022; Wagstaff et al. 2022). *PTEN* silencing is recognized as a molecular marker of metastatic melanoma (Vidotto et al. 2020). *KISS1* (a metastasis suppressor gene) knockout results in neoplastic proliferation after the tumor cells seed in the metastatic niche. Increased *MITF* amplification was observed in metastases as compared to primary melanomas. Increased expression of anti-apoptotic proteins (*MCL1*, *BCL-*

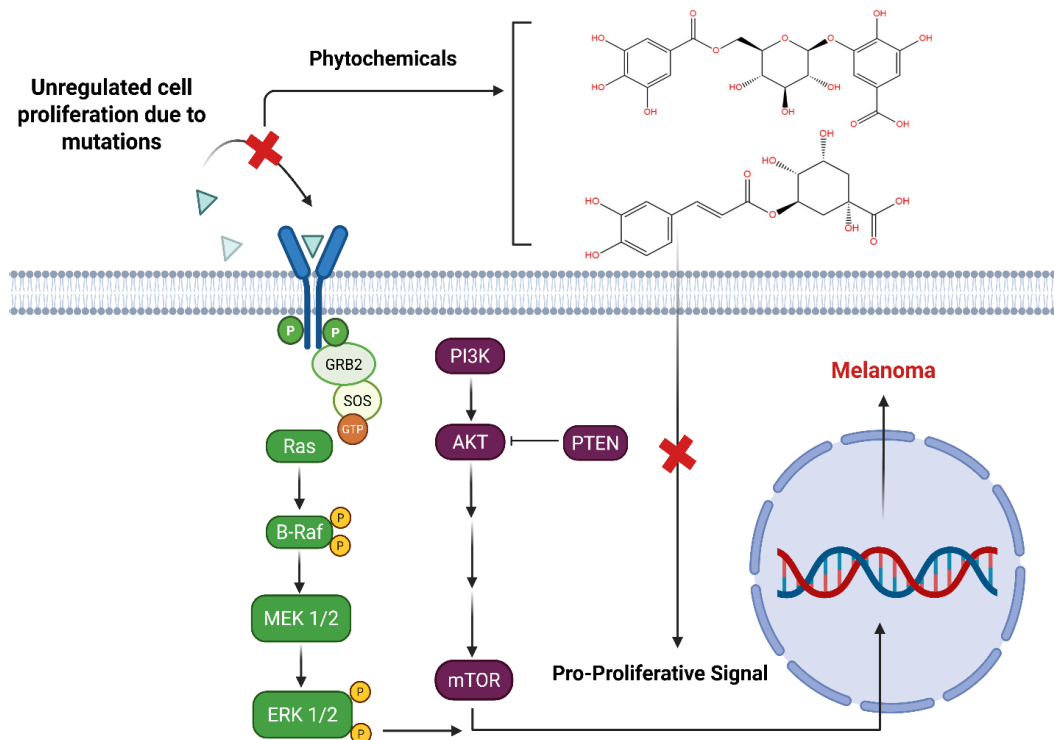


Figure 1: Schematic representation of the attenuation of the molecular pathways of melanoma through phytochemicals derived from various plant sources.

XL, survivin (*BIRC5*), XIAP) and decreased expression of BCL-2 are seen when moving from the radial to vertical growth phase. Overall, the process requires tumor cells first to dissociate from the primary tumor mass, survive the trip, and begin to proliferate within a distant niche, therefore requiring the coordinated expression of many genes. Tumor cells of this stage can grow in soft agar and are used for murine model studies (Frischhut et al. 2022).

3. Chemotherapeutic Drugs Used in Melanoma Treatment and Their Limitations

Treatment options for melanoma are provided in ASCO guidelines. HLA-A*02:01-positive uveal melanoma can be treated using tebentafusp. For cutaneous melanoma that can be surgically resected, pembrolizumab as neoadjuvant and adjuvant is recommended for stage IIB or IIC cutaneous melanoma. Adjuvant nivolumab, pembrolizumab, or a combination of dabrafenib and trametinib is used for resectable melanoma

stage IIIA to IIID (the combination of dabrafenib and trametinib is used in BRAF mutant (V600E/K*) patients and not in BRAF wild type ones). Metastatic or non-resectable cutaneous melanoma can be dealt with options including pembrolizumab, nivolumab, a combination of relatlimab and nivolumab, and nivolumab given after combination of ipilimumab and nivolumab for BRAF wild-type, while those having BRAF-mutant (V600), nivolumab alone or combinations of cobimetinib plus vemurafenib, binimetinib plus encorafenib, trametinib plus dabrafenib, pembrolizumab plus nivolumab, relatlimab plus nivolumab, and ipilimumab after nivolumab are recommended for use (Seth et al. 2023). An overview of the safety, toxicity, and efficacy status of chemotherapeutic drugs used in melanoma treatment, as provided in guidelines, is summarized in **Table 1**.

In the management of melanoma, limitations are seen, such as unwanted AEs which can result in GI and dermatologic toxicities, and reduced drug

effectiveness as a result of drug resistance (Domingues et al. 2018). This resistance, developed either during administration of chemotherapy (acquired resistance) or present at onset (primary or intrinsic resistance) is a major issue limiting the efficacy of such drugs (Kalal, Upadhy, and Pai 2017). Molecular mechanisms such as defective drug transport through ABC efflux pumps, altered apoptotic pathway regulation (like Bcl-2 overexpression), enhanced enzymatic detoxification, and increased DNA repair activity are observed in chemoresistance (Nigam et al. 2025). This resistance and the invasive and metastatic potential of melanoma are a major reason behind treatment failure and extremely poor prognosis (Krasowska, Kurzepa, and Błaszczak 2024).

4. Phytochemicals with Anti-proliferative Effect on Melanoma Cells

The WHO reports that a significant number of the world's population uses traditional medicine to treat various ailments. These medicinal plants contain phytochemicals in their stem, leaves, roots, flowers, and grains. that exhibit significant therapeutic potential in a multitude of diseases (Sood, Choi, and Lee 2024). Furthermore, due to their minimal cost and toxicity, and use as supplements, they are worth considering for disease treatment. Multiple studies have identified phytochemicals that can be used in the treatment and prevention of metastatic melanoma, such as silymarin, capsaicin, genistein, curcumin, resveratrol, proanthocyanidins, luteolin, apigenin, fisetin, indole-3-carbinol, and epigallocatechin-3-gallate (Chandra Pal et al. 2016).

4.1. Insights from Published *in vitro* and *in vivo* Studies

Compounds that are derived from plants have demonstrated significant anti-melanoma effects in *in vitro* and *in vivo* studies. A summary of 24 studies demonstrating the anti-neoplastic effects of plant-derived phytochemicals is given in Table 2. Many phytochemicals, especially polyphenols (such as chlorogenic acid, rosmarinic acid,

quercetin, catechin derivatives, and rutin), broadly impact oxidative stress, inflammation, and the activity of multiple melanoma-related proteins (such as TYRP1, MITF, and certain kinases) to consistently reduce melanoma cell viability (Nargatti and Wadkar 2024; Sarikurku, Erdoğan, and Yazar 2024; Ko et al. 2020). This is illustrated in **Figures 1** and **2**. The limited pre-clinical and clinical data regarding the safety and efficacy of these extracts highlight the need for a broader understanding of the mechanisms of multi-target effects of these phytochemicals. Extracts rich in alkaloids and terpenoids from *Berberis* species, *Cedrus atlantica*, *Corallocarpus epigaeus*, and plants that produce isoquinoline alkaloids disrupt mitochondrial function, inhibit protein synthesis, and interfere with cell-cycle regulation (Aiswarya et al. 2022; Tuzimski et al. 2024; Moyo, Mhlongo, and Sitole 2024). Multiple studies also report that these compounds amplify the effects of standard chemotherapeutic agents. For instance, extract from *Cedrus atlantica* increases the sensitivity of melanoma cells to 5-fluorouracil (5-FU) and diminishes markers of drug resistance. This is significant as the development of resistance to 5-fluorouracil, dacarbazine, and even BRAF and MEK inhibitors is a major hurdle in the treatment of melanoma. There are many phytochemicals that directly target the signaling pathways that are involved in cell survival and apoptosis, and these may also help reverse or delay resistance to treatment (Moyo, Mhlongo, and Sitole 2024).

Animal studies further support this. Extracts from plants that include *Melilotus indicus*, *Aloysia citrodora*, *Allium subhirsutum*, and *Ramalina terebrata* have all been shown to reduce tumor burden, inflammation, angiogenesis, or metastasis in mouse models (Choi et al. 2023; Badraoui et al. 2020; Rashid et al. 2022; Bashir et al. 2022). Such studies are relevant as most approved therapies, whether immune checkpoint inhibitors or targeted therapies, have efficacy but are often associated with AEs. Many reports on these phytochemicals cite far lower toxicity, although pharmacokinetic

Table 2: Summary of phytochemical constituents in plant extracts in in vivo, in vitro, and in silico studies demonstrating anti-melanoma activity.

Plant Source	Phytochemicals Identified	Cell Line / Model Used	Findings	References
<i>Marrubium lutescens</i>	Chlorogenic acid, Rosmarinic acid	G361 cells	Significant anti-tumor activity, inhibited TYRP1 and MARK4 in docking results	(Sarikurku, Erdoğan, and Yazar 2024)
<i>Berberis pruinosa</i>	Palmitine, Berberine	Danio rerio larvae, A375 melanoma cells	Strong cytotoxic activity (IC ₅₀ = 32.54 µg/mL)	(Tuzimski et al. 2024)
<i>Lansea coromandelica</i>	Quercetin, Myricadiol, Catechin	B16F10 melanoma cells	Potential anti-cancer activity (IC ₅₀ = 9.69 µg/mL), highest binding affinity for TYRP1	(Nargatti and Wadkar 2024)
<i>Orbea variegata</i>	Flavonoids, Tannins, Phenolic compounds	Swiss mouse model	Reduced oxidative stress, inflammation, and skin hyperplasia	(Chgari et al. 2024)
<i>Galium verum</i>	Isoquercitrin, Rutin, Chlorogenic acid	A375 melanoma cells	Reduced cell viability even at low concentrations (55 µg/mL)	(Semenescu et al. 2024)
<i>Alchemilla vulgaris</i>	Not mentioned	Fem-X, 518A2, B16F10, B16F1 melanoma cells	Suppressed tumor growth and reduced viability	(Jelača et al. 2024)
<i>Campomanesia xanthocarpa</i>	Epigallocatechin gallate, Ellagic acid, Rutin, Chlorogenic acid, Isoquercitrin, Kaempferol-3-O-rutinoside	SK-MEL-28 melanoma cells	Fruit infusion had stronger antineoplastic activity than leaf infusion	(da Silva et al. 2024)
<i>Luisia tenuifolia Blume</i>	Tannins, Flavonoids, Phenolic compounds	A431 human skin squamous carcinoma cells	Significant anti-melanoma activity (IC ₅₀ = 24.29 µg/mL)	(Sethuraman and Ramachandran 2024)
<i>Kalanchoe fedtschenkoi</i>	Gallic acid 3-O-(6'-O-galloyl)-glucopyranoside, Gallic acid 4-O-(6'-O-galloyl)-glucopyranoside, Gallic acid	MV3, B16F10 melanoma cells	Cytotoxic activity: IC ₅₀ = 56.6 µg/mL, 43.0 µg/mL (MV3), 66.1 µg/mL, 93.6 µg/mL (B16F10)	(Casanova et al. 2024)
<i>Cedrus atlantica</i>	α-cedrene, Cedrol, Cuparene, γ-muuroloene, Thujopsene	B16F10 murine cells	Synergistic suppression of tumor growth and reduction in 5-FU resistance	(Gao et al. 2024)
<i>Ocimum tenuiflorum</i>	Methyl palmitate, Linolenic acid, Stearic acid, Palmitic acid, Myristic acid	A375 melanoma cells	IC ₅₀ values < 50 µg/mL, ethyl acetate extract had greater cytotoxicity	(Moyo, Mhlongo, and Sitole 2024)
<i>Betula pendula</i>	Lupeol, Botulin, Betulinic acid, Erythrodiol	Walker 256 carcinoma cells, B16F10 murine cells	Nanolipid complexes and Pegylated liposomes showed in vitro antitumor activity	(Rugină et al. 2024)
<i>Melilotus indicus</i>	Quercetin, Coumarin	DMBA-induced cancer mouse model	Significant anti-tumor activity plus multiple tumor reduction parameters	(Bashir et al. 2022)
<i>Pseudofumaria lutea, Fumaria officinalis, Thalictrum foetidum, Macleaya cordata, Lamprocapnos spectabilis, Meconopsis cambrica</i>	Isoquinoline alkaloids	SK-MEL-3, G-361, A375 melanoma cells	Cytotoxic activity dependent on isoquinoline alkaloid levels	(Misiurek et al. 2023)
<i>Ramalina terebrata</i>	1,3,7,9-tetrahydroxy-2,8-dimethyl-4,6-di[ethanoyl]dibenzofuran	B16F10 melanoma cells	Dose and time-dependent reduction of cell viability, reduced tumor growth	(Choi et al. 2023)
<i>Aloysia citrodora</i>	Eupatilin, 5,6,4'-Trihydroxy-7,3'-Dimethoxyflavone, 5,7-Dihydroxy-2'-Methoxyflavone, Baicalein, Hispidulin, Naringenin	MDA-MB231, Vero, EMT-6, T47D, MCF7 cell lines	Significant reduction in tumor size and undetected tumors in Balb/C mice	(Rashid et al. 2022)
<i>Corallocarpus epigaeus</i>	Cucurbitacin B	A375 melanoma cells, Swiss albino mice, NOD-SCID murine model	Tumor reduction and safety assessment confirmed antineoplastic activity	(Aiswarya et al. 2022)
<i>Piper capense</i>	Saponins, Polyphenols, Sterols, Tannins, Alkaloids	B16F10 cells, C57BL/6J mice	Cytotoxic effects in vitro and tumor size mitigation in vivo	(Wamba et al. 2021)
<i>Allium subhirsutum</i>	p-coumaric acid	Murine model and Walker 256/B cells	Reduced osteolytic metastases and tumor angiogenesis. IC ₅₀ value is roughly 150 µg/mL.	(Badraoui et al. 2020)
<i>Dracocephalum kotschyi Boiss</i>	Xanthomicrol trimethoxylated hydroxyflavone)	(a) C57BL/6 mice + B16F10 melanoma cells	Reduced cell viability. IC ₅₀ = 3.433 µg/mL. Significant potential for	(Ghazizadeh et al. 2020)

				melanoma treatment is shown by the MTT assay.	
<i>Annona squamosa</i>	β -sitosterol, 18-oxokauran-17-yl acetate, ent-kaur-16-en-19-ol	B16F10 melanoma cell line	cell	Anti-proliferative activity via activation of p38 and inhibition of MITF	(Ko et al. 2020)
<i>Oenothera biennis</i>	Rosmarinic acid, ferulic acid, epicatechin, gallic acid, coumaric acid, rutin, caffeic acid	A375 melanoma cell line		Cytotoxic effects observed at 60 μ g/m;	(Fecker et al. 2020)
<i>Moringa oleifera</i>	Flavonoids	B16-F10 melanoma cell lines	– Mouse	Significant cytotoxicity (IC ₅₀ of 127.12 μ g/ml); Showed high apoptosis potential (67.89%);	(Umair et al. 2025)
<i>Sphagneticola trilobata</i>	Terpenoids, Trilobolide-6-O-isobutyrate, and a mixture of kaurenoic acid/grandiflorenic acid.	A-431 cell line (human epidermoid carcinoma).	(human skin	Kaurenoic acid/grandiflorenic acid exhibited stronger cytotoxicity.	(Ali et al. 2025)

studies to date are still sparse (Güven et al. 2022). The pharmacokinetic profile is one of the limitations of current melanoma therapies. For instance, dacarbazine is poorly bioavailable, and targeted therapies require carefully planned dosing due to narrow safety margins. Immunotherapies can produce long-lasting toxicities that affect quality of life. While phytochemicals offer lower toxicity, they can suffer from issues such as poor circulation time, compromised stability, modest bioavailability, and rapid clearance (Mohapatra et al. 2022). New formulations, such as nanolipid carriers, can help address these issues and improve delivery to tumor tissue through *Betula pendula* extracts, providing therapeutic strategies. In summary, published literature indicates multiple strengths of phytochemicals, such as their ability to act on multiple pathways at once. They have demonstrated evidence of providing a synergistic response with chemotherapeutic agents. In general, pre-clinical and clinical evidence suggest phytochemicals have lower toxicity than systemic therapy. Phytochemicals have also been shown to alter pathways against which chemotherapies are not consistently effective, such as metastasis, inflammation, and angiogenesis. This provides a strong scientific basis for the investigation of melanoma treatment and the exploration of phytochemicals as part of combination approaches, to decrease the potential for resistance with toxic agents, decrease toxicity, and broaden therapeutic options.

4.2. Limitation of Phytochemicals

Despite being considered safe to consume, herbal products can also produce AEs, urging their careful administration. The emergence of AEs can be related to factors like contamination by microbes, pollutants, or adulterants, and the use of certain plant metabolites or parts that have a toxic potential. Unlike allopathic medicine, the limited availability of clinical and toxicity data and loose regulations on manufacture reduce the safety of herbal medicine. When they are taken concomitantly with certain medicines, they may manipulate their plasma levels by inhibiting or inducing certain enzymes that play a role in the metabolism of that drug, thus decreasing their efficacy or causing AEs, especially for drugs with a low therapeutic index. The absorption of phytochemicals is compromised because of their diverse hydrophilicity and lipophilicity. They also have reduced pharmacologic activity, a high risk of degradation, and low stability. As a result, the bioavailability of phytochemicals is relatively low, thus limiting the therapeutic potential of these compounds. In addition, it is difficult to convince patients of their usage due to the limited understanding of the biological pathways of phytochemicals, which restricts translation of phytochemical models from preclinical models to practice (Kim et al. 2025). Furthermore, for approval of their use as anticancer agents in patients, phytochemicals require substantial data of efficacy, safety, and quality derived from appropriate clinical trials (Garcia-Oliveira et al. 2021).

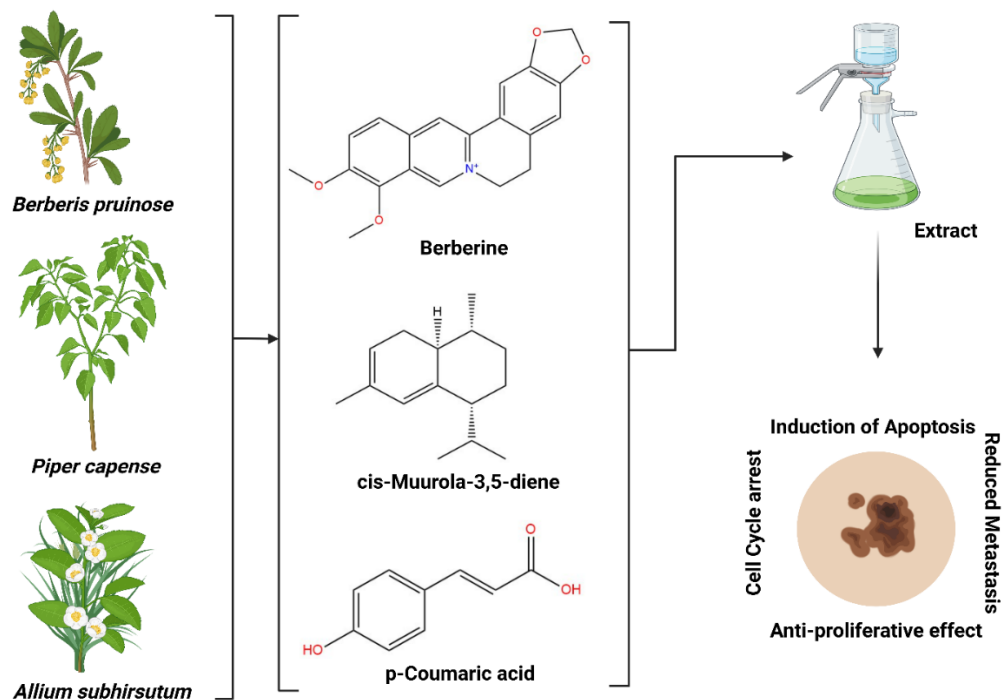


Figure 2: Schematic representation of the anti-proliferative activity of the major phytochemicals obtained from various plant extracts.

5. Conclusion

The undesirable therapeutic outcome, low survival rate, high mortality, and increasing incidence of melanoma exist despite the presence of clinical choices like radiation therapy, surgery, and chemotherapy. This is linked to the metastatic potential and heterogeneity of melanoma and the many AEs caused by chemotherapy drugs. This is the reason why phytochemicals have received focus by researchers, as evident from the increase in the quantity of scholarly articles in the last decade. This review compiled such *in vivo*, *in vitro*, and *in silico* studies to showcase the anti-proliferative potential of such plant-derived secondary metabolites against melanoma while having the benefit of being significantly safer to use in comparison to chemotherapy. However, herbal medicine should be taken following careful consideration, as it can also cause certain AEs. If we are considering the use of phytochemicals in melanoma, they must be prepared to the level of quality and regulation as allopathic medicine and must undergo clinical trials to assess their safety

and clinical efficacy, find optimal dosage, investigate possible interactions between herbs and medications, and see if the preclinical benefits can be translated to patients with melanoma.

Conflict of Interest

The authors declare no competing interests.

Funding

No funding to disclose.

Availability of data and materials

All data generated or analyzed during this study are included in this published article. Data can be available on demand.

Ethics approval and consent to participate.

Not applicable

Authors' contributions

AI designed the study and carried out most of the data-acquiring parts. MMJ and MHH performed data acquisition and search. All authors read and approved the final manuscript.

Acknowledgments

Not applicable

References

- Ahad, A., F. Shakeel, M. Raish, A. Ahmad, Y. A. Bin Jordan, F. I. Al-Jenoobi, and A. M. Al-Mohizea. 2022. "Thermodynamic Solubility Profile of Temozolomide in Different Commonly Used Pharmaceutical Solvents." *Molecules* no. 27 (4):1437. doi:10.3390/molecules27041437.
- Aiswarya, S. U. D., G. Vikas, N. H. Haritha, V. B. Liju, A. Shabna, M. Swetha, T. P. Rayginia, C. K. Keerthana, L. R. Nath, M. V. R. Reshma, S. Sundaram, N. P. Anto, R. S. Lankalapalli, R. J. Anto, and S. V. Bava. 2022. "Cucurbitacin B, purified and characterized from the rhizome of *Corallocarpus epigaeus* exhibits anti-melanoma potential." *Frontiers in Oncology* no. 12:903832. doi:10.3389/fonc.2022.903832.
- Akin-Bali, D. F. 2021. "Bioinformatics analysis of GNAQ, GNA11, BAP1, SF3B1, SRSF2, EIF1AX, PLCB4, and CYSLTR2 genes and their role in the pathogenesis of uveal melanoma." *Ophthalmic Genetics* no. 42 (6):732–743. doi:10.1080/13816810.2021.1961280.
- Ali Khani J., P. A. Harsini, G. Imani, and S. Hamzehie. 2023. "Melanoma epidemiology: symptoms, causes, and preventions." *In Melanoma – Standard of Care, Challenges, and Updates in Clinical Research*, IntechOpen. doi:10.5772/intechopen.107096.
- Ali, M. T., D. A. Al-Mahdy, A. M. El Fishawy, A. Salama, A. A. Al-Karmalawy, and A. M. Otify. 2025. "Phytochemical investigation, role in wound healing process and cytotoxicity of *Sphagneticola trilobata*: in vitro, in vivo and in silico approach." *Journal of Ethnopharmacology* no. 342:119394. doi:10.1016/j.jep.2025.119394.
- Almoussalam, M., and H. Zhu. 2016. "Encapsulation of cancer therapeutic agent dacarbazine using nanostructured lipid carrier." *Journal of Visualized Experiments* (110):53760. doi:10.3791/53760.
- Altun, I., and A. Sonkaya. 2018. "The most common side effects experienced by patients receiving first cycle of chemotherapy." *Iranian Journal of Public Health* no. 47 (8):1218–1219.
- Arias-Mejias, S. M., K. Y. Warda, E. Quattrocchi, H. Alonso-Quinones, S. Somnidi-Damodaran, and A. Meves. 2020. "The role of integrins in melanoma: a review." *International Journal of Dermatology* no. 59 (5):525–534. doi:10.1111/ijd.14850.
- Badraoui, R., T. Rebai, S. Elkahoui, M. Alreshidi, V. N. Veettil, E. Noumi, K. A. Al-Motair, K. Aouadi, A. Kadri, V. De Feo, and M. Snoussi. 2020. "Allium subhirsutum L. as a potential source of antioxidant and anticancer bioactive molecules: HR-LCMS phytochemical profiling, in vitro and in vivo pharmacological study." *Antioxidants* 9 (10):1003. doi:10.3390/antiox9101003.
- Bashir, A., M. Asif, M. Saadullah, M. Saleem, S. H. Khalid, L. Hussain, I. U. Khan, H. S. Yaseen, H. M. Zubair, M. U. Shamas, R. Al Zarzour, and T. A. Chohan. 2022. "Therapeutic potential of standardized extract of *Melilotus indicus* (L.) All. and its phytochemicals against skin cancer in animal model: in vitro, in vivo, and in silico studies." *ACS Omega* 7 (29):25772–25782. doi:10.1021/acsomega.2c03053.
- Blank, C. U., M. W. Lucas, R. A. Scolyer, B. A. van de Wiel, A. M. Menzies, M. Lopez-Yurda, L. L. Hoeijmakers, R. P. M. Saw, J. M. Lijnsvelt, N. G. Maher, S. M. Pulleman, M. Gonzalez, A. Torres Acosta, W. J. van Houdt, S. N. Lo, A. M. J. Kuijpers, A. Spillane, W. M. C. Klop, T. E. Pennington, and others. 2024. "Neoadjuvant Nivolumab and Ipilimumab in Resectable Stage III Melanoma." *New England Journal*

- of *Medicine* no. 391 (18):1696–1708. doi: 10.1056/NEJMoa2402604.
- Boutros, A., E. Croce, M. Ferrari, R. Gili, G. Massaro, R. Marconcini, L. Arecco, E. T. Tanda, and F. Spagnolo. 2024. "The treatment of advanced melanoma: Current approaches and new challenges." *Critical Reviews in Oncology/Hematology* no. 196: 104276. doi: 10.1016/j.critrevonc.2024.104276.
- Broit, N., P. A. Johansson, C. B. Rodgers, S. T. Walpole, N. K. Hayward, and A. L. Pritchard. 2022. "Systematic review and meta-analysis of genomic alterations in acral melanoma." *Pigment Cell & Melanoma Research* no. 35 (3):369–386. doi:10.1111/pcmr.13034.
- Brugnara, S., M. Sicher, E. M. Bonandini, D. Donner, F. Chierichetti, M. Barbareschi, C. R. Girardelli, and O. Caffo. 2018. "Treatment with combined dabrafenib and trametinib in BRAF(V600E)-mutated metastatic malignant melanoma: a case of long-term complete response after treatment cessation." *Drugs in Context* no. 7: 212515. doi: 10.7573/dic.212515.
- Caraban, B. M., M. Aschie, M. Deacu, G. C. Cozaru, M. B. Pundiche, C. I. Orasanu, and R. I. Voda. 2024. "A narrative review of current knowledge on cutaneous melanoma." *Clinics and Practice* no. 14 (1):214–241. doi:10.3390/clinpract14010018.
- Casadevall, D., J. Vidal, F. Gallardo, F. Zuccarino, M. Arumí-Uría, A. Dalmases, B. Bellosillo, and C. Montagut. 2016. "Dabrafenib in an elderly patient with metastatic melanoma and BRAF V600R mutation: a case report." *Journal of Medical Case Reports* no. 10 (1):158. doi:10.1186/s13256-016-0953-0.
- Casanova, L. M., J. X. do Nascimento Júnior, J. E. de Souza, R. do Couto Rodrigues, A. B. Araújo, E. Arrais, L. C. de Oliveira Silva, Y. M. B. Menezes, R. M. K. Yien, L. W. Tinoco, S. S. Costa, M. Sola-Penna, and P. Zancan. 2024. "Biochemometrics-based identification of gallic acid and gallic acid galloylglucosides from *Kalanchoe fedtschenkoi* with cytotoxic effects on cultured melanoma cells." *Phytochemical Analysis* no. 36 (4):957–972. doi:10.1002/pca.3480.
- Casarotto, E., S. Chandwani, L. Mortier, O. Dereure, C. Dutriaux, S. Dalac, E. Scherrer, L. Lévy-Bachelot, L. Verdoni, G. Farge, C. Allayous, B. Oriano, S. Dalle, and C. Lebbé. 2021. "Real-World Effectiveness of Pembrolizumab in Advanced Melanoma: Analysis of a French National Clinicobiological Database." *Immunotherapy* no. 13(11):905–916. doi: 10.2217/imt-2021-0077.
- Chgari, O., H. Wahnou, M. Ndayambaje, F. Moukhfi, O. Benkhniq, F. Marnissi, Y. Limami, and M. Oudghiri. 2024. "Orbea variegata (L.) Haw in Skin Carcinogenesis: Insights from an In Vivo Male Swiss Mouse Model Study." *Journal of Toxicology and Environmental Health, Part A* no. 87(15):630–645. doi: 10.1080/15287394.2024.2354790.
- Choi, S., H. Kim, S.-A. Shin, M. Kim, S. Y. Moon, M. Kim, S. Lee, J. H. Lee, H. H. Park, U. J. Youn, and C. S. Lee. 2023. "DB3 from Antarctic Lichen Inhibits the Growth of B16F10 Melanoma Cells In Vitro and In Vivo." *Applied Biological Chemistry* no. 66(1):77. doi: 10.1186/s13765-023-00835-w.
- Ciarletta, P., L. Foret, and M. Ben Amar. 2011. "The Radial Growth Phase of Malignant Melanoma: Multi-Phase Modelling, Numerical Simulations and Linear Stability Analysis." *Journal of the Royal Society Interface* no. 8(56):345–368. doi: 10.1098/rsif.2010.0285.
- da Silva, V. R. F., G. B. da Silva, D. Manica, C. T. P. Deolindo, M. D. Bagatini, and A. P. Kempka. 2024. "Phytotherapeutic Potential of *Campomanesia xanthocarpa* (Mart.) O. Berg: Antitumor Effects in Vitro and In Silico, With Emphasis on SK-MEL-28

- Melanoma Cells-A Study on Leaf and Fruit Infusions." *In Silico Pharmacology* no. 12(2):105. doi: 10.1007/s40203-024-00286-1.
- Damsky, W. E., and M. Bosenberg. 2017. "Melanocytic Nevi and Melanoma: Unraveling a Complex Relationship." *Oncogene* no. 36(42):5771–5792. doi: 10.1038/onc.2017.189.
- Delgado-Bellido, D., A. Chacón-Barrado, J. Olmedo-Pelayo, C. Jordán Pérez, P. Gilabert-Prieto, J. Díaz-Martín, A. García-Díaz, F. J. Oliver, and E. de Álava. 2025. "Chromosomal 3p Loss and 8q Gain Drive Vasculogenic Mimicry via HIF-2 α and VE-Cadherin Activation in Uveal Melanoma." *Cell Death & Differentiation* no. 32:1473–1483. doi: 10.1038/s41418-025-01469-9.
- Dessinioti, C., A. C. Geller, and A. J. Stratigos. 2022. "A Review of Nevus-Associated Melanoma: What Is the Evidence?" *Journal of the European Academy of Dermatology and Venereology* no. 36(11):1927–1936. doi: 10.1111/jdv.18453.
- Domingues, B., J. M. Lopes, P. Soares, and H. Pópulo. 2018. "Melanoma Treatment in Review." *ImmunoTargets and Therapy* no. 7:35-49. doi: 10.2147/ITT.S134842.
- Eggermont, A. M. M., M. Kicinski, C. U. Blank, M. Mandala, G. V. Long, V. Atkinson, S. Dalle, A. Haydon, A. Meshcheryakov, A. Khattak, M. S. Carlino, S. Sandhu, J. Larkin, S. Puig, P. A. Ascierto, P. Rutkowski, D. Schadendorf, M. Boers-Sonderen, A. M. Di Giacomo, A. J. M. van den Eertwegh, J.-J. Grob, R. Gutzmer, R. Jamal, A. C. J. van Akkooi, P. Lorigan, D. Grebennik, C. Krepler, S. Marreaud, S. Suci, and C. Robert. 2022. "Five-Year Analysis of Adjuvant Pembrolizumab or Placebo in Stage III Melanoma." *NEJM Evidence* no. 1(11):EVIDoA2200214. doi: 10.1056/EVIDoA2200214.
- Faghfuri, E., M. A. Faramarzi, S. Nikfar, and M. Abdollahi. 2015. "Nivolumab and Pembrolizumab as Immune-Modulating Monoclonal Antibodies Targeting the PD-1 Receptor to Treat Melanoma." *Expert Review of Anticancer Therapy* no. 15(9):981–993. doi: 10.1586/14737140.2015.1074862.
- Fecker, R., V. Buda, E. Alexa, S. Avram, I. Z. Pavel, D. Muntean, I. Cocan, C. Watz, D. Minda, C. A. Dehelean, C. Soica, and C. Danciu. 2020. "Phytochemical and Biological Screening of *Oenothera biennis* L. Hydroalcoholic Extract." *Biomolecules* no. 10(6):818. doi: 10.3390/biom10060818.
- Ferrara, G., and G. Argenziano. 2021. "The WHO 2018 Classification of Cutaneous Melanocytic Neoplasms: Suggestions From Routine Practice." *Frontiers in Oncology* no. 11:675296. doi: 10.3389/fonc.2021.675296.
- Flanagan, N., E. Healy, A. Ray, S. Philips, C. Todd, I. J. Jackson, M. A. Birch-Machin, and J. L. Rees. 2000. "Pleiotropic Effects of the Melanocortin 1 Receptor (MC1R) Gene on Human Pigmentation." *Human Molecular Genetics* no. 9(17):2531–2537. doi: 10.1093/hmg/9.17.2531.
- Frischhut, N., B. Zelger, F. André, and B. G. Zelger. 2022. "The Spectrum of Melanocytic Nevi and Their Clinical Implications." *Journal der Deutschen Dermatologischen Gesellschaft* no. 20(4):483–504. doi: 10.1111/ddg.14776.
- Galus, Ł., D. T. Małecka, M. Michalak, S. Kopeć, K. Kozak, K. Piejko, M. Ziętek, Z. Chowaniec, B. C. Stopa, R. Dziura, J. Żubrowska, G. K. Winciorek, W. Bal, P. Rutkowski, and J. Mackiewicz. 2025. "Chemotherapy Efficacy in Advanced Melanoma Patients After Failure of Immune Checkpoint and BRAF/MEK Inhibitors." *Contemporary Oncology (Poznań)* no. 29(2):165–170. doi: 10.5114/wo.2025.150451.
- Gao, H.-W., K.-F. Chang, X.-F. Huang, M.-C. Lee, N.-M. Tsai, and T.-H. Chen. 2024. "Cedrus atlantica Extract Inhibits Melanoma Progression by Suppressing Epithelial-Mesenchymal Transition and Inducing

- Mitochondria-Mediated Apoptosis." *Medical Oncology* no. 42(1):22. doi: 10.1007/s12032-024-02573-5.
- Garbe, C., T. Amaral, K. Peris, A. Hauschild, P. Arenberger, N. Basset-Seguín, L. Bastholt, V. Bataille, V. Del Marmol, B. Dréno, M. C. Fagnoli, A.-M. Forsea, J.-J. Grob, C. Höller, R. Kaufmann, N. Kelleners-Smeets, A. Lallas, C. Lebbé, B. Lytvynenko, J. Malvey, D. Moreno-Ramirez, P. Nathan, G. Pellacani, P. Saiag, A. J. Stratigos, A. C. J. van Akkooi, R. Vieira, I. Zalaudek, and P. Lorigan. 2022. "European Consensus-Based Interdisciplinary Guideline for Melanoma. Part 1: Diagnostics: Update 2022." *European Journal of Cancer* no. 170:236–255. doi: 10.1016/j.ejca.2022.03.008.
- Garcia-Oliveira, P., P. Otero, A. G. Pereira, F. Chamorro, M. Carpena, J. Echave, M. Fraga-Corral, J. Simal-Gandara, and M. A. Prieto. 2021. "Status and Challenges of Plant-Anticancer Compounds in Cancer Treatment." *Pharmaceuticals* no. 14 (2):157. doi: 10.3390/ph14020157.
- Gaughan, E. M., and B. J. Horton. 2022. "Outcomes From Cytotoxic Chemotherapy Following Progression on Immunotherapy in Metastatic Melanoma: An Institutional Case-Series." *Frontiers in Oncology* no. 12. doi: 10.3389/fonc.2022.855782.
- Ghazizadeh, F., M. Shafiei, R. Falak, M. Panahi, N. Rakhshani, S. A. Ebrahimi, and P. Rahimi-Moghaddam. 2020. "Xanthomicrol Exerts Antiangiogenic and Antitumor Effects in a Mouse Melanoma (B16F10) Allograft Model." *Evidence-Based Complementary and Alternative Medicine* no. 2020:8543872. doi: 10.1155/2020/8543872.
- Goldinger, S. M., K. Buder-Bakhaya, S. N. Lo, A. Forschner, M. McKean, L. Zimmer, C. Khoo, R. Dummer, Z. Eroglu, E. I. Buchbinder, P. A. Ascierto, R. Gutzmer, E. A. Rozeman, C. Hoeller, D. B. Johnson, A. Gesierich, P. Kölblinger, N. Bennannoune, J. V. Cohen, K. C. Kähler, M. A. Wilson, J. Cebon, V. Atkinson, J. L. Smith, O. Michielin, G. V. Long, J. C. Hassel, B. Weide, L. E. Haydu, D. Schadendorf, G. McArthur, P. A. Ott, C. Blank, C. Robert, R. Sullivan, A. Hauschild, M. S. Carlino, C. Garbe, M. A. Davies, and A. M. Menzies. 2022. "Chemotherapy after immune checkpoint inhibitor failure in metastatic melanoma: a retrospective multicentre analysis." *European Journal of Cancer* no. 162:22–33. doi: 10.1016/j.ejca.2021.11.022.
- Goldstein, A. M., and M. A. Tucker. 2013. "Dysplastic nevi and melanoma." *Cancer Epidemiology, Biomarkers & Prevention* no. 22 (4):528–532. doi: 10.1158/1055-9965.EPI-12-1346.
- Gray-Schopfer, V. C., S. C. Cheong, H. Chong, J. Chow, T. Moss, Z. A. Abdel-Malek, R. Marais, D. Wynford-Thomas, and D. C. Bennett. 2006. "Cellular senescence in naevi and immortalisation in melanoma: a role for p16?" *British Journal of Cancer* no. 95 (4):496–505. doi: 10.1038/sj.bjc.6603283.
- Guo, Y., Y. Chen, L. Zhang, L. Ma, K. Jiang, G. Yao, and L. Zhu. 2022. "TERT Promoter Mutations and Telomerase in Melanoma." *Journal of Oncology* no. 2022:6300329. doi: 10.1155/2022/6300329.
- Güven, D. C., T. K. Sahin, A. Rizzo, A. D. Ricci, S. Aksoy, and K. Sahin. 2022. "The Use of Phytochemicals to Improve the Efficacy of Immune Checkpoint Inhibitors: Opportunities and Challenges." *Applied Sciences* no. 12 (20):10548. doi: 10.3390/app122010548.
- Haist, M., H. Stege, R. Ebner, M. I. Fleischer, C. Loquai, and S. Grabbe. 2022. "The Role of Treatment Sequencing with Immune-Checkpoint Inhibitors and BRAF/MEK Inhibitors for Response and Survival of Patients with BRAFV600-Mutant Metastatic Melanoma-A Retrospective, Real-World Cohort Study." *Cancers*

- (*Basel*) no. 14 (9):2082. doi: 10.3390/cancers14092082.
- Hassel, J. C., S. Piperno-Neumann, P. Rutkowski, J.-F. Baurain, M. Schlaak, M. O. Butler, R. J. Sullivan, R. Dummer, J. M. Kirkwood, M. Orloff, J. J. Sacco, S. Ochsenreither, A. M. Joshua, L. Gastaud, B. Curti, J. M. Piulats, A. K. S. Salama, A. N. Shoushtari, L. Demidov, M. Milhem, B. Chmielowski, K. B. Kim, R. D. Carvajal, O. Hamid, L. Collins, K. Ranade, C. Holland, C. Pfeiffer, and P. Nathan. 2023. "Three-Year Overall Survival with Tebentafusp in Metastatic Uveal Melanoma." *New England Journal of Medicine* no. 389 (24):2256–2266. doi: 10.1056/NEJMoa2304753.
- Heidenreich, B., E. Nagore, P. S. Rachakonda, Z. Garcia-Casado, C. Requena, V. Traves, J. Becker, N. Soufir, K. Hemminki, and R. Kumar. 2014. "Telomerase Reverse Transcriptase Promoter Mutations in Primary Cutaneous Melanoma." *Nature Communications* no. 5:3401. doi: 10.1038/ncomms4401.
- Hoffner, B., and K. Benchich. 2018. "Trametinib: A Targeted Therapy in Metastatic Melanoma." *Journal of the Advanced Practitioner in Oncology* no. 9 (7):741–745.
- Huang, F., F. Santinon, R. E. Flores González, and S. V. Del Rincón. 2021. "Melanoma Plasticity: Promoter of Metastasis and Resistance to Therapy." *Frontiers in Oncology* no. 11:756001. doi: 10.3389/fonc.2021.756001.
- Jelača, S., I. Jovanovic, D. Bovan, S. Pavlovic, N. Gajovic, D. Dunderović, Z. Dajić-Stevanović, A. Acović, S. Mijatović, and D. Maksimović-Ivanić. 2024. "Antimelanoma Effects of *Alchemilla vulgaris*: A Comprehensive In Vitro and In Vivo Study." *Diseases* no. 12 (6):125. doi: 10.3390/diseases12060125.
- Kalal, B. S., D. Upadhyaya, and V. R. Pai. 2017. "Chemotherapy Resistance Mechanisms in Advanced Skin Cancer." *Oncology Reviews* no. 11 (1):326. doi: 10.4081/oncol.2017.326.
- Kibbi, N., H. Kluger, and J. N. Choi. 2016. "Melanoma: Clinical Presentations." *Cancer Treatment and Research* no. 167:107–129. doi: 10.1007/978-3-319-22539-5_4.
- Kim, H. K., S. J. Kim, W. J. Gil, and C.-S. Yang. 2025. "Exploring the Therapeutic Potential of Phytochemicals: Challenges and Strategies for Clinical Translation." *Phytomedicine* no. 145:157090. doi: 10.1016/j.phymed.2025.157090.
- Ko, G.-A., H. R. Kang, J. Y. Moon, M. K. Ediriweera, S. Eum, T. T. Bach, and S. K. Cho. 2020. "Annona squamosa L. Leaves Inhibit Alpha-Melanocyte-Stimulating Hormone (α -MSH) Stimulated Melanogenesis via p38 Signaling Pathway in B16F10 Melanoma Cells." *Journal of Cosmetic Dermatology* no. 19 (7):1785–1792. doi: 10.1111/jocd.13223.
- Krasowska, D., J. Kurzepa, and E. Błaszczak. 2024. "Current State of Melanoma Treatment – From Conventional Therapies to Nanotechnology and Beyond." *Journal of Pre-Clinical and Clinical Research* no. 18 (4):333–340. doi: 10.26444/jpcrr/193990.
- Kreuger, I. Z. M., R. C. Sliker, T. van Groningen, and R. van Doorn. 2023. "Therapeutic Strategies for Targeting CDKN2A Loss in Melanoma." *Journal of Investigative Dermatology* no. 143 (1):18–25.e1. doi: 10.1016/j.jid.2022.07.016.
- Lopes, J., C. M. P. Rodrigues, M. M. Gaspar, and C. P. Reis. 2022. "Melanoma Management: From Epidemiology to Treatment and Latest Advances." *Cancers (Basel)* no. 14 (19):4652. doi: 10.3390/cancers14194652.
- Ma, Y., R. Xia, X. Ma, R. L. Judson-Torres, and H. Zeng. 2021. "Mucosal Melanoma: Pathological Evolution, Pathway Dependency and Targeted Therapy." *Frontiers in Oncology* no. 11:702287. doi: 10.3389/fonc.2021.702287.

- Mackiewicz-Wysocka, M., P. Czerwińska, V. Filas, E. Bogajewska, A. Kubicka, A. Przybyła, E. Dondajewska, T. Kolenda, A. Marszałek, and A. Mackiewicz. 2017. "Oncogenic BRAF Mutations and p16 Expression in Melanocytic Nevi and Melanoma in the Polish Population." *Advances in Dermatology and Allergology* no. 34 (5):490–498. doi: 10.5114/ada.2017.71119.
- Márquez-Rodas, I., E. Muñoz Couselo, J. F. Rodríguez Moreno, A. M. Arance Fernández, M. Á. Berciano Guerrero, B. Campos Balea, L. de la Cruz Merino, E. Espinosa Arranz, A. García Castaño, and A. Berrocal Jaime. 2024. "SEOM-GEM Clinical Guidelines for Cutaneous Melanoma (2023)." *Clinical and Translational Oncology* no. 26 (11):2841–2855. doi: 10.1007/s12094-024-03497-2.
- Miller, A. J., and M. C. Mihm Jr. 2006. "Melanoma." *New England Journal of Medicine* no. 355 (1):51–65. doi: 10.1056/NEJMra052166.
- Misiurek, J., T. Plech, B. Kaproń, A. Makuch-Kocka, M. Szultka-Młyńska, B. Buszewski, and A. Petruczynik. 2023. "Determination of Some Isoquinoline Alkaloids in Extracts Obtained from Selected Plants of the Ranunculaceae, Papaveraceae and Fumarioideae Families by Liquid Chromatography and In Vitro and In Vivo Investigations of Their Cytotoxic Activity." *Molecules* no. 28 (8):3503. doi: 10.3390/molecules28083503.
- Mohapatra, P., P. Singh, D. Singh, S. Sahoo, and S. K. Sahoo. 2022. "Phytochemical Based Nanomedicine: A Panacea for Cancer Treatment, Present Status and Future Prospective." *OpenNano* no. 7:100055. doi: 10.1016/j.onano.2022.100055.
- Morgan, A. M., J. Lo, and D. E. Fisher. 2013. "How Does Pheomelanin Synthesis Contribute to Melanomagenesis?: Two Distinct Mechanisms Could Explain the Carcinogenicity of Pheomelanin Synthesis." *BioEssays* no. 35 (8):672–676. doi: 10.1002/bies.201300020.
- Moyo, B., M. Mhlongo, and L. Sitole. 2024. "Cytotoxic Activity of *Ocimum tenuiflorum* Crude Extracts Against an A375 Malignant Melanoma Cell Line." *South African Journal of Botany* no. 175:514–522. doi: 10.1016/j.sajb.2024.10.049.
- Napoli, S., C. Scuderi, G. Gattuso, V. Di Bella, S. Candido, M. S. Basile, M. Libra, and L. Falzone. 2020. "Functional Roles of Matrix Metalloproteinases and Their Inhibitors in Melanoma." *Cells* no. 9 (5):1151. doi: 10.3390/cells9051151.
- Nargatti, P. I., and K. A. Wadkar. 2024. "Anticancer Activity of *Lannea coromandelica* on B16F10 Melanoma Cell Line: An In Vitro and Molecular Docking Approach." *Journal of Herbal Medicine* no. 48:100958. doi: 10.1016/j.hermed.2024.100958.
- Nelson, A. A., and H. Tsao. 2009. "Melanoma and Genetics." *Clinics in Dermatology* no. 27 (1):46–52. doi: 10.1016/j.clindermatol.2008.09.005.
- Ng, M. F., J. L. Simmons, and G. M. Boyle. 2022. "Heterogeneity in Melanoma." *Cancers (Basel)* no. 14 (12):3030. doi: 10.3390/cancers14123030.
- Nigam, S., E. Enshaie, J. Smith, and V. Rai. 2025. "Chemoresistance in Cutaneous Melanoma: Contemporary and Future Aspects." *Chinese Clinical Oncology* no. 14 (3):34. doi: 10.21037/cco-25-9.
- Nurla, L. A., and A.-M. Forsea. 2024. "Melanoma Epidemiology in Europe: What Is New?" *Italian Journal of Dermatology and Venereology* no. 159 (2):128–134. doi: 10.23736/s2784-8671.24.07811-3.
- O'Day, S. J., M. Maio, V. Chiarion-Sileni, T. F. Gajewski, H. Pehamberger, I. N. Bondarenko, P. Queirolo, L. Lundgren, S. Mikhailov, L. Roman, C. Verschraegen, R. Humphrey, R. Ibrahim, V. de Pril, A. Hoos, and J. D. Wolchok. 2010. "Efficacy and

- Safety of Ipilimumab Monotherapy in Patients with Pretreated Advanced Melanoma: A Multicenter Single-Arm Phase II Study." *Annals of Oncology* no. 21 (8):1712–1717. doi: 10.1093/annonc/mdq013.
- Olmedo, M. E., R. Cervera, L. Cabezon-Gutierrez, Y. Lage, E. Corral de la Fuente, A. Gómez Rueda, X. Mielgo-Rubio, J. C. Trujillo, and F. Couñago. 2022. "New Horizons for Uncommon Mutations in Non-Small Cell Lung Cancer: BRAF, KRAS, RET, MET, NTRK, HER2." *World Journal of Clinical Oncology* no. 13 (4):276–286. doi: 10.5306/wjco.v13.i4.276.
- Pal, H. C., K. M. Hunt, A. Diamond, C. A. Elmets, and F. Afaq. 2016. "Phytochemicals for the Management of Melanoma." *Mini Reviews in Medicinal Chemistry* no. 16 (12):953–979. doi: 10.2174/1389557516666160211120157.
- Paluncic, J., Z. Kovacevic, P. J. Jansson, D. Kalinowski, A. M. Merlot, M. L. H. Huang, H. C. Lok, S. Sahni, D. J. R. Lane, and D. R. Richardson. 2016. "Roads to Melanoma: Key Pathways and Emerging Players in Melanoma Progression and Oncogenic Signaling." *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research* no. 1863 (4):770–784. doi: 10.1016/j.bbamcr.2016.01.025.
- Parkman, G. L., X. Xu, S. L. Holmen, and F. A. Karreth. 2025. "The Roles of PTEN in Melanoma Suppression." *Pigment Cell & Melanoma Research* no. 38 (5):e70054. doi: 10.1111/pcmr.70054.
- Penta, D., B. S. Somashekar, and S. M. Meeran. 2018. "Epigenetics of Skin Cancer: Interventions by Selected Bioactive Phytochemicals." *Photodermatology, Photoimmunology & Photomedicine* no. 34 (1):42–49. doi: 10.1111/phpp.12353.
- Rashid, H. M., A. I. Mahmood, F. U. Afifi, and W. H. Talib. 2022. "Antioxidant and Antiproliferation Activities of Lemon Verbena (*Aloysia citrodora*): An In Vitro and In Vivo Study." *Plants* no. 11 (6):785. doi: 10.3390/plants11060785.
- Rashid, S., M. Shaughnessy, and H. Tsao. 2023. "Melanoma Classification and Management in the Era of Molecular Medicine." *Dermatologic Clinics* no. 41 (1):49–63. doi: 10.1016/j.det.2022.07.017.
- Rugină, D., M. A. Socaciu, M. Nistor, Z. Diaconeasa, M. Cenariu, F. A. Țăbăran, and C. Socaciu. 2024. "Liposomal and Nanostructured Lipid Nanoformulations of a Pentacyclic Triterpenoid Birch Bark Extract: Structural Characterization and In Vitro Effects on Melanoma B16-F10 and Walker 256 Tumor Cells Apoptosis." *Pharmaceuticals* no. 17 (12):1630. doi: 10.3390/ph17121630.
- Saginala, K., A. Barsouk, J. S. Aluru, P. Rawla, and A. Barsouk. 2021. "Epidemiology of Melanoma." *Medical Sciences* no. 9 (4):63. doi: 10.3390/medsci9040063.
- Samłowski, W., G. V. Long, and J. F. Thompson. 2024. "The Effect of Non-Overlapping Somatic Mutations in BRAF, NRAS, NF1, or CKIT on the Incidence and Outcome of Brain Metastases During Immune Checkpoint Inhibitor Therapy of Metastatic Melanoma." *Cancers (Basel)* no. 16 (3):594.
- Sample, A., and Y.-Y. He. 2018. "Mechanisms and Prevention of UV-Induced Melanoma." *Photodermatology, Photoimmunology & Photomedicine* no. 34 (1):13–24.
- Sarikurkcü, C., S. F. Erdoğan, and T. Yazar. 2024. "Phytochemical Analysis and In Vitro Anti-Inflammatory, Anticancer Activities of *Marrubium lutescens* on Melanoma Cancer Cell Line and Molecular Docking Studies." *Journal of Herbal Medicine* no. 46:100907.
- Schuler, M., L. Zimmer, K. B. Kim, J. A. Sosman, P. A. Ascierto, M. A. Postow, F. De Vos, C. M. L. van Herpen, M. S. Carlino, D. B. Johnson, C. Berking, M. B. Reddy, A. S. Harney, J. D. Berlin, and R. N. Amaria.

2022. "Phase Ib/II Trial of Ribociclib in Combination with Binimetinib in Patients with NRAS-Mutant Melanoma." *Clinical Cancer Research* no. 28 (14):3002–3010. doi: 10.1158/1078-0432.CCR-21-3872.
- Scolyer, R. A., G. V. Long, and J. F. Thompson. 2011. "Evolving Concepts in Melanoma Classification and Their Relevance to Multidisciplinary Melanoma Patient Care." *Molecular Oncology* no. 5 (2):124–136. doi: 10.1016/j.molonc.2011.03.002.
- Semenescu, A.-D., E.-A. Moacă, A. Iftode, C.-A. Dehelean, D.-S. Tchiakpe-Antal, L. Vlase, S. Rotunjanu, D. Muntean, S. D. Chiriac, and R. Chioibaş. 2024. "Recent Updates Regarding the Antiproliferative Activity of *Galium verum* Extracts on A375 Human Malignant Melanoma Cell Line." *Life* no. 14 (1):112.
- Seth, R., H. Messersmith, V. Kaur, J. M. Kirkwood, R. Kudchadkar, J. L. McQuade, A. Provenzano, U. Swami, J. Weber, K. C. Alluri, S. Agarwala, P. A. Ascierto, M. B. Atkins, N. Davis, M. S. Ernstoff, M. B. Faries, J. S. Gold, S. Guild, D. E. Gyorki, N. I. Khushalani, M. O. Meyers, C. Robert, M. Santinami, A. Sehdev, V. K. Sondak, G. Spurrier, K. K. Tsai, A. van Akkooi, and P. Funchain. 2020. "Systemic Therapy for Melanoma: ASCO Guideline." *Journal of Clinical Oncology* no. 38 (33):3947–3970. doi: 10.1200/JCO.20.00198.
- Seth, R., S. S. Agarwala, H. Messersmith, K. C. Alluri, P. A. Ascierto, M. B. Atkins, K. Bollin, M. Chacon, N. Davis, and M. B. Faries. 2023. "Systemic Therapy for Melanoma: ASCO Guideline Update." *Journal of Clinical Oncology* no. 41 (30):4794–4820.
- Sethuraman, S. P., and K. Ramachandran. 2024. "Phytochemical Profiling, In-Vitro Antioxidant and Cytotoxic Effects of *Luisia tenuifolia* Extracts Against Human Skin Squamous Carcinoma." *Applied Biochemistry and Biotechnology* no. 196 (1)
- Shain, A. H., I. Yeh, I. Kovalyshyn, A. Sriharan, E. Talevich, A. Gagnon, R. Dummer, J. North, L. Pincus, and B. Ruben. 2015. "The Genetic Evolution of Melanoma from Precursor Lesions." *New England Journal of Medicine* no. 373 (20):1926–1936.
- Sibira, R., A. Vu, A. Giubellino, and P. Murugan. 2024. "Spitz Melanoma with MAP3K8::ABLIM1 Rearrangement: A Case Report with Review of the Literature." *Diagnostic Pathology* no. 19 (1):133.
- Skrehot, H. C., A. F. Alsoudi, and A. C. Scheffler. 2025. "Co-Occurrence of EIF1AX, SF3B1, or BAP1 Variants in Uveal Melanomas: A Case Series and Review." *American Journal of Ophthalmology Case Reports* no.:102327.
- Sood, R., H.-K. Choi, and H.-J. Lee. 2024. "Potential Anti-Cancer Properties of Malvidin and Its Glycosides: Evidence from In Vitro and In Vivo Studies." *Journal of Functional Foods* no. 116:106191.
- Staeger, R., A. Tastanova, A. Ghosh, N. Winkelbeiner, P. Shukla, I. Kolm, P. Turko, A. Benlahrech, J. Harper, A. Broomfield, and others. 2025. "Tebentafusp Elicits On-Target Cutaneous Immune Responses Driven by Cytotoxic T Cells in Uveal Melanoma Patients." *Journal of Clinical Investigation* no. 135 (12):e181464.
- Swetter, S. M., J. A. Thompson, M. R. Albertini, C. A. Barker, J. Baumgartner, G. Boland, B. Chmielowski, D. DiMaio, A. Durham, and R. C. Fields. 2021. "NCCN Guidelines® Insights: Melanoma: Cutaneous, Version 2.2021: Featured Updates to the NCCN Guidelines." *Journal of the National Comprehensive Cancer Network* no. 19 (4):364–376.
- Tabolacci, C., D. De Vita, A. Facchiano, G. Bozzuto, S. Beninati, C. M. Failla, M. Di Martile, C. Lintas, C. Mischiati, A. Stringaro, D. Del Bufalo, and F. Facchiano. 2023. "Phytochemicals as

- Immunomodulatory Agents in Melanoma." *International Journal of Molecular Sciences* no. 24 (3):2657. doi: 10.3390/ijms24032657
- Takata, M., H. Murata, and T. Saida. 2010. "Molecular Pathogenesis of Malignant Melanoma: A Different Perspective from the Studies of Melanocytic Nevus and Acral Melanoma." *Pigment Cell & Melanoma Research* no. 23 (1):64–71. doi: 10.1111/j.1755-148X.2009.00645.x.
- Tang, X., T. Yang, D. Yu, H. Xiong, and S. Zhang. 2024. "Current Insights and Future Perspectives of Ultraviolet Radiation (UV) Exposure: Friends and Foes to the Skin and Beyond the Skin." *Environment International* no. 185:108535. doi: 10.1016/j.envint.2024.108535.
- Tuzimski, T., A. Petruczynik, B. Kaproń, T. Plech, A. Makuch-Kocka, D. Janiszewska, M. Sugajski, B. Buszewski, and M. Szultka-Młyńska. 2024. "In Vitro and In Silico of Cholinesterases Inhibition and In Vitro and In Vivo Anti-Melanoma Activity Investigations of Extracts Obtained from Selected *Berberis* Species." *Molecules* no. 29 (5):1048. doi: 10.3390/molecules29051048.
- Umaima, U. S., V. Veeraraghavan, and G. Kavitha Singh. 2025. "Anti-Cancer Potential of *Moringa oleifera* Leaves Methanol Extract: An In Vitro Study." *National Journal of Pharmacology* no. 54 (3). doi: 10.48047/n86m3z81.
- Vergara, I. A., J. S. Wilmott, G. V. Long, and R. A. Scolyer. 2021. "Genetic Drivers of Non-Cutaneous Melanomas: Challenges and Opportunities in a Heterogeneous Landscape." *Experimental Dermatology* no. 30 (3). doi: 10.1111/exd.14287.
- Vidotto, T., C. M. Melo, E. Castelli, M. Koti, R. B. dos Reis, and J. A. Squire. 2020. "Emerging Role of PTEN Loss in Evasion of the Immune Response to Tumours." *British Journal of Cancer* no. 122 (12):1732–1743. doi: 10.1038/s41416-020-0834-6.
- Wagstaff, W., R. N. Mwamba, K. Grullon, M. Armstrong, P. Zhao, B. Hendren-Santiago, K. H. Qin, A. J. Li, D. A. Hu, A. Youssef, R. R. Reid, H. H. Luu, L. Shen, T.-C. He, and R. C. Haydon. 2022. "Melanoma: Molecular Genetics, Metastasis, Targeted Therapies, Immunotherapies, and Therapeutic Resistance." *Genes & Diseases* no. 9 (6):1608–1623. doi: 10.1016/j.gendis.2022.04.004.
- Wamba, B. E. N., P. Ghosh, A. T. Mbaveng, S. Bhattacharya, M. Debarpan, D. Saha, S. M. Mustafi, V. Kuete, and N. Murmu. 2021. "Botanical from *Piper capense* Fruit Can Help to Combat Melanoma as Demonstrated by In Vitro and In Vivo Studies." *Evidence-Based Complementary and Alternative Medicine* no. 2021:8810368. doi: 10.1155/2021/8810368.
- Yu, H., R. McDaid, J. Lee, P. Possik, L. Li, S. M. Kumar, D. E. Elder, P. Van Belle, P. Gimotty, M. Guerra, R. Hammond, K. L. Nathanson, M. Dalla Palma, M. Herlyn, and X. Xu. 2009. "The Role of BRAF Mutation and p53 Inactivation During Transformation of a Subpopulation of Primary Human Melanocytes." *American Journal of Pathology* no. 174 (6):2367–2377. doi: 10.2353/ajpath.2009.081057.
- Zbytek, B., J. A. Carlson, J. Granese, J. Ross, M. C. Mihm Jr., and A. Slominski. 2008. "Current Concepts of Metastasis in Melanoma." *Expert Review of Dermatology* no. 3 (5):569–585. doi: 10.1586/17469872.3.5.569.
- Zob, D. L., I. Augustin, L. Caba, M.-C. Pânzaru, S. Popa, A. D. Popa, L. Florea, and E. V. Gorduza. 2022. "Genomics and Epigenomics in the Molecular Biology of Melanoma - A Prerequisite for Biomarker Studies." *International Journal of Molecular Sciences* no. 24 (1):716.