



Check for  
updates



## Review Article

# Intelligent Reprogramming of Crops for Resistance Against Emerging Plant Pathogens: From Conventional Breeding to Genome Editing

Jabir Asghar<sup>1</sup>, Ameer Jan<sup>2</sup>, Muhammad Hamayun<sup>3</sup>, Khatir Ali<sup>4</sup>, Atiq U Rehman<sup>3</sup>, Mushtaq Ahmed<sup>3</sup>, Izaz Ullah<sup>5</sup>, Abdul Basit<sup>6</sup>, Shaikh Saddam<sup>7</sup>

<sup>1</sup>Southwest University of Science and Technology, Mianyang, Sichuan, China.

<sup>2</sup>Department of Botany, University of Makran, Panjgur, Pakistan.

<sup>3</sup>Muhammad Nawaz Shareef University of Agriculture, Multan, Pakistan.

<sup>4</sup>Department of Plant Breeding and Genetics, University of Agriculture Faisalabad, Pakistan.

<sup>5</sup>Government Post Graduate College, Dargai, Malakand, Pakistan.

<sup>6</sup>Abdul Wali Khan University, Mardan, Pakistan.

<sup>7</sup>Department of Plant Pathology, Faculty of Agriculture, Lasbela University of Agriculture Water and Marine Sciences, Uthal, Pakistan.

## ABSTRACT

Emerging plant pathogens are threat to global food security. Plant pathogens are rapidly spreading due to increasing globalization, agricultural intensification and rapidly changing climatic conditions. These emerging pathogens are rapidly adapting to overcome host resistance. Therefore, necessitating the development of sustainable disease management strategies. Breeding for resistance is an important approach for sustainable disease management. The advancement in molecular genetics, genomics, and biotechnology have revolutionized the field of breeding for resistance. Precise incorporation of resistant genes into elite cultivars has been possible due to advanced breeding techniques such as Marker-assisted selection, genomic selection, and genome editing tools. However, there are still several challenges in plant breeding. The development of resistant varieties is further complicated by public perception and regulatory hurdles surrounding genetically modified crops. However, using integrated pest management such as resistant varieties along with cultural, biological and chemical control methods can enhance the durability of resistance. The development of resilient crop varieties has been possible with Advances in omics technologies, high-throughput phenotyping, and pathogen detection methods. Besides the development of resilient variety, it is essential to deliver resistant varieties to farmers. International collaboration, open-access breeding networks, and public-private partnerships play important role in delivering the resistant varieties to farmers. Resistance breeding needs to be proactive to face the challenges of emerging plant pathogens and global food security. This review highlights the factors causing spread of emerging plant pathogens, discuss modern approaches, key challenges and future direction of resistant breeding.

**Keywords:** Emerging plant pathogens, Disease resistance breeding, Marker-assisted selection, Genomic selection, Genome editing

## INTRODUCTION

Emerging plant pathogens such as nematodes, fungi, bacteria, viruses, and oomycetes are becoming a major threat to agricultural systems. According to Kantor *et al.* (2024), these pathogens possess challenges for plant health and food production systems. Pathogens host range have increased causing increased pathogen incidences geographically. Further globalization, climate change, and intensification of agricultural practices are generally increasing the introduction



### \*Correspondence

Ameer Jan

ameerJan@uomp.edu.pk

ameer.jeehand143@gmail.com

### Article History

Received: August 24, 2025

Accepted: September 21, 2025

Published online: September 25, 2025

### Cite this article

Asghar, J., Jan, A., Hamayun, M., Ali, K., Rehman, A. U., Ahmed, M., Ullah, I., Basit, A., & Saddam, S. (2025). Intelligent reprogramming of crops for resistance against emerging plant pathogens: From conventional breeding to genome editing. *Integrative Plant Biotechnology*, 03, 253-268.



**Copyright:** © 2025 by the authors.

**Licensee:** Roots Press, Rawalpindi, Pakistan.

This article is an open-access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license: <https://creativecommons.org/licenses/by/4.0>

and establishment of these emerging pathogens in new ecological systems (Hossain *et al.*, 2024). Emerging plant pathogens have significantly reduced crop yield and quality worldwide. Moreover, rising global population and demand for food products are further affecting food security. Staple food supplies are threatened by outbreaks of emerging pathogens such as virulent races of rust fungi in wheat and bacterial wilt of banana in banana plantations (Gai and Wang, 2024; Virk *et al.*, 2024). The management of plant pathogens has become increasingly challenging in recent years due to rapid evolution of pathogen populations. Environmental pressure on plants such as changing temperatures, elevated atmospheric carbon dioxide is weakening the plant resistance and pathogens are adapting to these conditions within few cropping cycles. Therefore, development of environment and insect resistant varieties is crucial for food security (Dracatos *et al.*, 2023).

Breeding for resistance is environmentally and economically sustainable method of plant disease management. Breeding plants for resistance has been revolutionized by advances in molecular genetics, genomics and biotechnology. Modern breeding techniques such as marker assisted selection (MAS), genomic selection (GS), and genome editing tools (CRISPR/Cas9) are efficient to develop elite cultivars with precise incorporation of resistant genes (Yıldırım *et al.*, 2023). Resistance breeding has progressed from traditional methods such as phenotypic selection to precise molecular breeding, detecting cloned resistance (R) genes and pathogen effectors that facilitate resistance breeding, as conceptualized in the gene-for-gene model (Figure 1). Laying the groundwork for a new era of plant disease resistance breeding. The advancement in resistance breeding is the solution to present and future emerging pathogen threats (Pedrozo *et al.*, 2025; Švara *et al.*, 2024; Dracatos *et al.*, 2023). This review paper discusses the factors of emerging plant pathogens, explores the advances in breeding for resistance, identify the key challenges and future direction for sustainable crop protection.

## EMERGING PLANT PATHOGENS: CHARACTERISTICS AND CHALLENGES

### Factors Driving Emergence of Pathogens

The emergence and spread of plant pathogens are driven by a combination of environmental, economic, and agricultural factors. Climate change is playing central role as it modifies the temperature regimes, precipitation patterns, and atmospheric composition, including elevated CO<sub>2</sub> levels. The changes speed up life cycles of pathogens, make them more fertile, and expand their geographic boundaries (Kumar and Mukhopadhyay, 2024; Nitta *et al.*, 2024). Warm winters allow pests and pathogens to overwinter in the areas where they were previously unable to overwinter and changed rainfall patterns enable environmental factors to contribute to the occurrence of epidemics (Kumar and Mukhopadhyay, 2024). In addition, plant health can be negatively affected by the reactive nitrogen gases and other pollution materials in the atmosphere, which increase susceptibility to emerging diseases (Alfizar and Nasution, 2024). Trade and globalization have significantly contributed in movement of plant material (seeds, fruits, nurseries) across continents. It facilitates the process of agricultural diversification, although it also enables the interruption of ecological barriers to plant pathogens and their vectors (Brooker *et al.*, 2023). Pathogens establish in new introduced regions rapidly due to absence of their natural predators or resistant local strains (Oraon *et al.*, 2024). The agricultural intensification also increases the risk of pathogens emergence. Economically efficient large-scale monocultures lead to the creation of genetically identical host environments that allow the development of pathogens and their evolution (Fan *et al.*, 2025). Crop rotation and shifts in cropping patterns whereby crops are cultivated into different ecological zones due to market demand and population pressure subject crops to new pathogens outbreaks (Mihrete and Mihretu, 2025).

Table 1 summarizes the most threatening emerging plant pathogens and pests affecting staple crops worldwide, underscoring the diversity of causal agents and their devastating consequences. The listed pathogens, ranging from fungi such as *Puccinia graminis* (Singh *et al.*, 2015; Soko *et al.*, 2018) and *Magnaporthe oryzae* (Nalley *et al.*, 2016) to viruses like ACMV (Combala *et al.*, 2024) and BBTV (Watanabe *et al.*, 2013), bacteria such as *Ralstonia solanacearum* (Kurabachew and Ayana, 2017) and *Xanthomonas vasicola* (Uwimana *et al.*, 2024), and even parasitic weeds like *Striga hermonthica* (Midega *et al.*, 2017) illustrate the broad spectrum of biotic stresses confronting global agriculture. Bacterial blight of cotton (*Xanthomonas citri* pv. *malvacearum*) (Shah *et al.*, 2025; Anwer *et al.*, 2024) and brown spot of rice (*Bipolaris oryzae*) (Shamshad *et al.*, 2024; Qi *et al.*, 2025) are also recognized as serious emerging threats, causing significant yield and quality losses in major cotton- and rice-growing regions worldwide. Their modes of spread, including windborne spores, insect vectors, contaminated tools, and seedbanks, facilitate rapid dissemination across regions, often leading to severe yield reductions and economic losses. The data highlights not only the vulnerability of staple crops such as wheat, rice, banana, cassava, and maize but also the urgent necessity of breeding for durable resistance as a central strategy to safeguard global food security.

Table 1. Major emerging plant pathogens, their hosts, geographic distribution, modes of spread, and associated economic impacts

Pathogen name	Pathogen type	Primary host crop(s)	Current geographic distribution	Mode of spread	Estimated yield loss / economic impact	Key references
<i>Puccinia graminis</i> f. sp. <i>tritici</i>	Fungus (basidiomycete)	Wheat	East Africa, Middle East, spreading to Asia	Windborne spores	Up to 80% yield loss in susceptible varieties	(Singh <i>et al.</i> , 2015; Soko <i>et al.</i> , 2018)
<i>African cassava mosaic virus</i> (ACMV)	Virus (Geminiviridae)	Cassava	Sub-Saharan Africa, expanding in Asia	Whitefly vector	24–75% yield reduction; up to US\$2.7 billion losses regionally	(Combala <i>et al.</i> , 2024)
<i>Xanthomonas vasicola</i> pv. <i>musacearum</i> (BXW)	Bacterium	Banana	East and Central Africa	Insect vectors, tools, planting material	Up to 100% yield loss; ~US\$25 billion potential losses in SSA over 10 years	(Uwimana <i>et al.</i> , 2024)
<i>Phytophthora infestans</i>	Oomycete	Potato, tomato	Global; notable in Latin America, Asia, Africa	Wind- and rain-borne sporangia	Up to 100% yield loss in unmanaged fields	(Fry <i>et al.</i> , 2015; Shattock, 2002)
<i>Magnaporthe oryzae</i>	Fungus (Ascomycete)	Rice	Asia, emerging globally	Windborne spores, irrigation	Major losses; often 10–30% in outbreaks	(Nalley <i>et al.</i> , 2016)
<i>Ralstonia solanacearum</i>	Bacterium	Potato, tomato, Solanaceae	Tropical and subtropical areas	Soil, water, infected tools	Significant losses; up to 60–80% under conducive conditions	(Kurabachew and Ayana, 2017)
<i>Puccinia graminis</i> f. sp. <i>hordei</i>	Fungus	Barley	Global, particularly Europe, North America	Windborne spores	Frequent yield losses, often 20–50% in susceptible areas	(Hatta <i>et al.</i> , 2020)
Banana bunchy top virus (BBTV)	Virus	Banana, plantain	Asia, Pacific, Africa	Aphid/vector, planting material	Severe yield reductions; plant stunting leads to crop failure	(Watanabe <i>et al.</i> , 2013)
<i>Fusarium graminearum</i> (FHB)	Fungus	Wheat, barley	Global, especially North America	Airborne spores	Up to 30% yield loss; high quality penalties due to mycotoxins	(Boutigny <i>et al.</i> , 2013)
<i>Striga hermonthica</i>	Parasitic weed	Cereals (maize, sorghum)	Sub-Saharan Africa	Seedbank, host proximity	Infestation on 5 million ha, losses >70%	(Midega <i>et al.</i> , 2017)

<i>Xanthomonas citri</i> pv. <i>malvacearum</i>	Bacterium	Cotton	Africa, Asia, some parts of USA	Seedborne, rain splash, contaminated tools	Up to 40% yield loss in severe epidemics	(Shah et al., 2025; Anwer et al., 2024)
<i>Bipolaris oryzae</i>	Fungus (Ascomycete)	Rice	Asia, Africa, Latin America	Seed and air borne conidia	16-43% yield loss; grain quality reduction	(Shamshad et al., 2024; Qi et al., 2025)

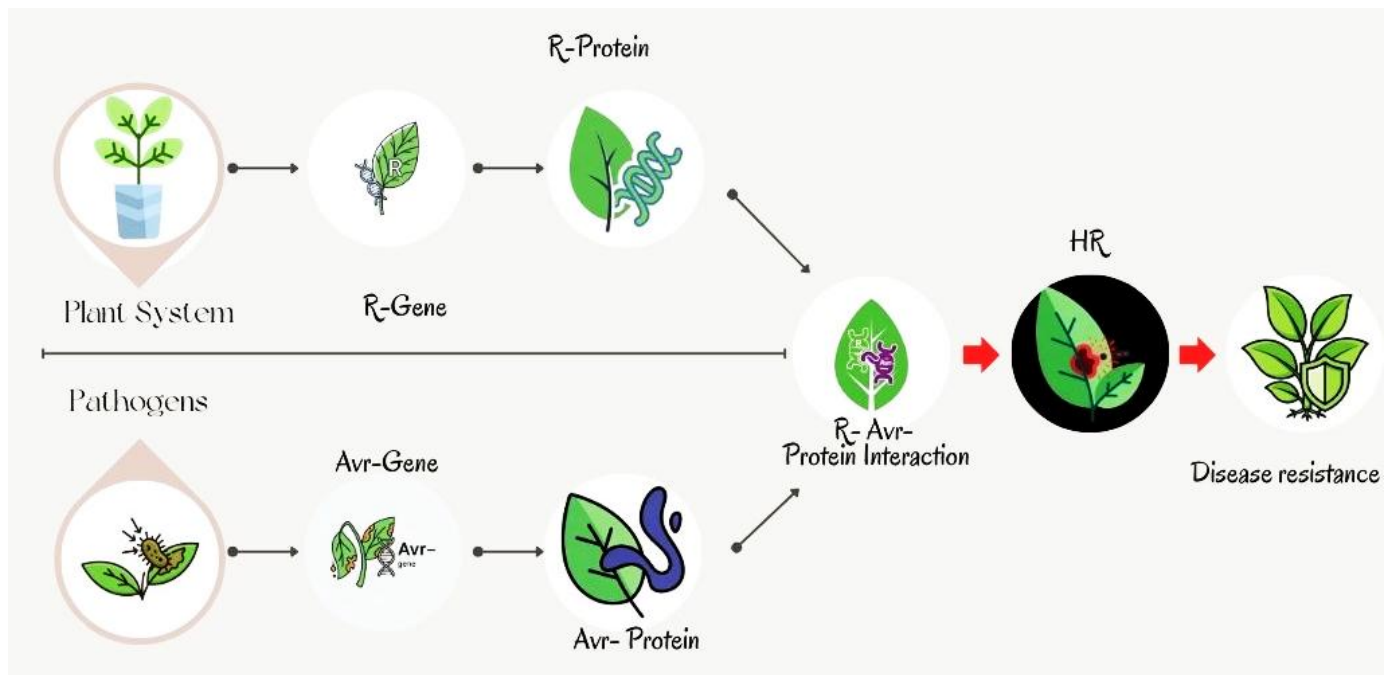


Figure 1: The gene-for-gene model of plant disease resistance, illustrating the interaction between plant resistance (R) genes and pathogen avirulence (Avr) genes, leading to hypersensitive response (HR) and resistance.

### Pathogen Adaptation and Resistance Breakdown

Even when resistant cultivars are achieved, pathogens can overcome defenses, by several mechanisms. A typical approach is mutation within a virulence gene, which modifies the molecular targets recognized by the plant immune receptors and this inhibits discovery as illustrated in the gene-for-gene model (Figure 1). There is also another mechanism based on effector diversification whereby the pathogens alter the effector proteins which are important to manipulate the host cellular activity. The mechanisms behind pathogen evasion of host recognition allows the pathogen to neutralize previous resistance genes (Zdrzałek et al., 2024). The durability of host resistance depends on both host genetics and pathogen adaptability. Polygene quantitative resistance is more durable than monogenic resistance (Çelik Oğuz and Karakaya, 2021) This durability is largely explained by the activation of complex host defense signaling cascades (Figure 2). Nevertheless, the prevention of quantitative resistance can also be eliminated over time as pathogens obtain low-cost escape mutations that do not reduce pathogen fitness (Chen et al., 2024).

## BREEDING FOR RESISTANCE: APPROACHES AND TECHNOLOGICAL ADVANCES

### Conventional Breeding Methods

Traditional breeding still constitutes a major direction in breeding disease-resistant crop varieties. Backcross breeding, mass selection, pure-line selection, and recurrent selection are methods that have widely been used in enhancing the adaptation and resistance characteristics in plants (Anand et al., 2023). The method of backcrossing is preferable to self-pollinated crops, whereas recurrent selection and hybridization are common among cross-pollinating species (Ajayi et al., 2024). In the past, the methods have recorded remarkable successes. In common beans, traditional breeding has been successful in increasing environmental adaptation to diverse conditions coupled with increased resistance to major pathogens (Cha et al., 2024).

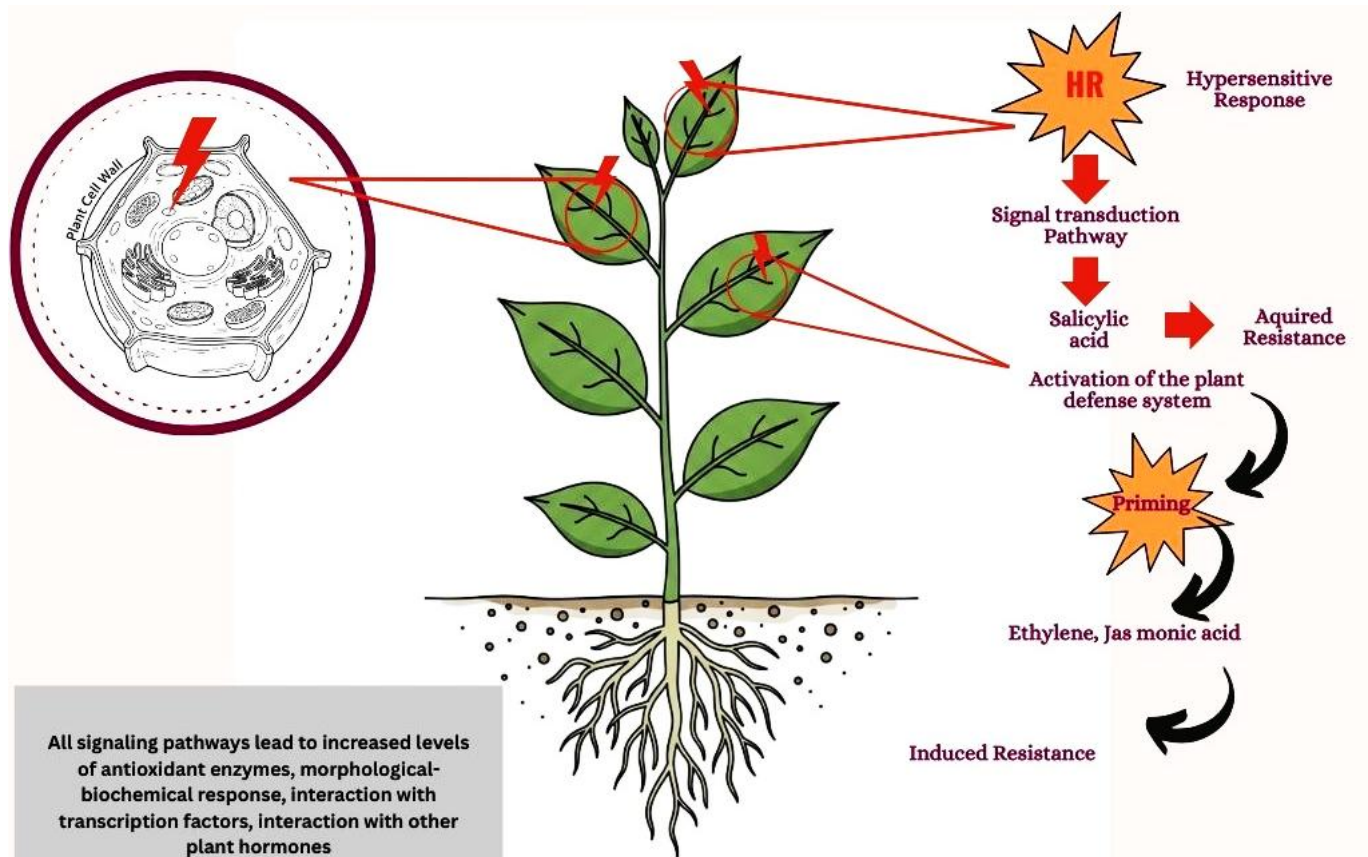


Figure 2. Stages of plant defense signaling pathways, including hypersensitive response (HR), salicylic acid-mediated acquired resistance, and priming-induced resistance through jasmonic acid and ethylene pathways.

However, it is time-consuming, especially against traits such as drought-tolerance or resistance to complex diseases. Limitations are dependent upon the existing genetic variability, long generation time and predisposed to genotype  $\times$  environment interactions. Although various disadvantages of conventional breeding are present, this breeding method costs less, and the infrastructure does not require much specialized equipment as modern molecular techniques and is still required when combined with newer technology (Isiyel *et al.*, 2024). Conventional approaches such as backcrossing and recurrent selection remain widely used for incorporating resistance traits. For example, recent work has demonstrated the successful introgression of multiple *Sr* genes (*Sr22*, *Sr33*, *Sr35*, *Sr45*) into wheat, significantly improving resistance to stem rust under diverse environments (Hatta *et al.*, 2020). Similarly, recurrent selection continues to be applied in maize breeding, yielding enhanced resistance to foliar diseases across different environments (Ćeran *et al.*, 2024) as elaborated in Table 2.

### Modern Breeding Tools

Limitations of conventional procedures have led to development of molecular and genomic tools that shorten breeding cycles, enhance the selection accuracy. Speed breeding time decreases between generations through manipulated environmental conditions such as ideal light, temperature, and day length so that up to five or six crop generations may be harvested in a year (Kumar and Walia, 2024). This accelerates genetic gains and can be used in conjunction with genomic for faster introduction of traits (Potts *et al.*, 2023). In marker-assisted selection (MAS), readily available molecular markers associated with desirable traits can be used to select individuals at an early stage of growth, preferably as seedlings, and minimize the requirement of extensive phenotypic screening (Song *et al.*, 2023). MAS is especially helpful for our understanding of single-major gene-based traits and finds a broader application on wheat crops, barley and rice (Mapari and Mehandi, 2024).

Genomic selection (GS) is an advancement of this concept by using genome-wide data on markers to predict the breeding value of all individuals in a population (Michel *et al.*, 2021). GS is the most effective on polygenic characteristics, including quantitative disease resistance, and in some scenarios, GS has been found to be more predictive than MAS, which was demonstrated in *Fusarium* head blight resistance in wheat (Mapari and Mehandi, 2024).

Table 2. Examples of resistance breeding approaches applied in major crops.

Approach	Crop and Disease	Key Achievement	Key References
Conventional backcrossing	Wheat - stem rust ( <i>Puccinia graminis</i> )	Resistant cultivars developed with Sr22, Sr33, Sr35, Sr45 genes introgressed from wild relatives	Hatta et al., 2020
Conventional recurrent selection	Maize - multiple foliar diseases	Improved resistance and stability under different environments	Ćeran et al., 2024
Marker-Assisted Selection (MAS)	Rice -bacterial blight ( <i>Xanthomonas oryzae</i> pv. <i>oryzae</i> )	Introgression of Xa4, Xa21, Xa33 genes for durable resistance	Derbyshire et al., 2024
MAS (QTL-based)	Wheat - stripe rust	Deployment of QTLs for adult plant resistance in elite germplasm	Wang et al., 2023
Genomic Selection (GS)	Wheat - Fusarium head blight	Enhanced prediction accuracy and faster selection compared to MAS	Poland and Rutkoski, 2016; Ćeran et al., 2024
High-Throughput Phenotyping (HTP)	Rice - blast	UAV-based hyperspectral imaging for early resistance screening	Khan et al., 2022
Gene pyramiding	Wheat - stripe rust ( <i>Puccinia striiformis</i> )	Additive resistance from pyramiding multiple adult plant resistance genes	Wang et al., 2023
QTL Mapping	Maize - northern leaf blight	Identification and deployment of QTLs linked with quantitative resistance	Samantara et al., 2022
Genome editing (CRISPR/Cas9)	Rice - bacterial blight	Editing of SWEET susceptibility genes improved durable resistance	Erdoğan et al., 2023
Genome editing (CRISPR/Cas9)	Wheat - powdery mildew	CRISPR knockout of TaMLO genes for durable resistance	Yıldırım et al., 2023
Utilization of wild relatives	Wheat - stripe rust	Resistance introgressed from <i>Aegilops tauschii</i> and wild einkorn	Derbyshire et al., 2024
Utilization of landraces	Rice - blast	Mining resistance alleles from Asian landraces for durable Pi genes	Umakanth et al., 2017; Kaur et al., 2022

High throughput phenotyping (HTP) is the integration of imaging and sensor technology and automation to measure quickly and non-destructively a high volume of plants. The identification of genotype phenotype relationships in disease resistance can be made by integrating genomic information and data obtained from RGB imaging, hyperspectral cameras, LiDAR and thermal sensors. HTP reduces phenotyping cost and accelerates selection decisions (Sabag et al., 2024; Thapa et al., 2024). The relative strengths and limitations of these strategies are captured in Figure 3, which highlights differences in cost, speed, and durability.

Marker-assisted selection has been particularly effective for rice bacterial blight, where genes such as *Xa4*, *Xa21*, and *Xa33* have been pyramided into elite cultivars to deliver durable resistance (Derbyshire et al., 2024). More recently, genomic selection has gained traction for predicting polygenic resistance traits; studies in wheat have shown superior performance in managing Fusarium head blight compared to traditional MAS (Ćeran et al., 2024). High-throughput phenotyping is also revolutionizing selection, with UAV-based hyperspectral imaging used successfully to detect early blast infections in rice (Khan et al., 2022) as given in Table 2.

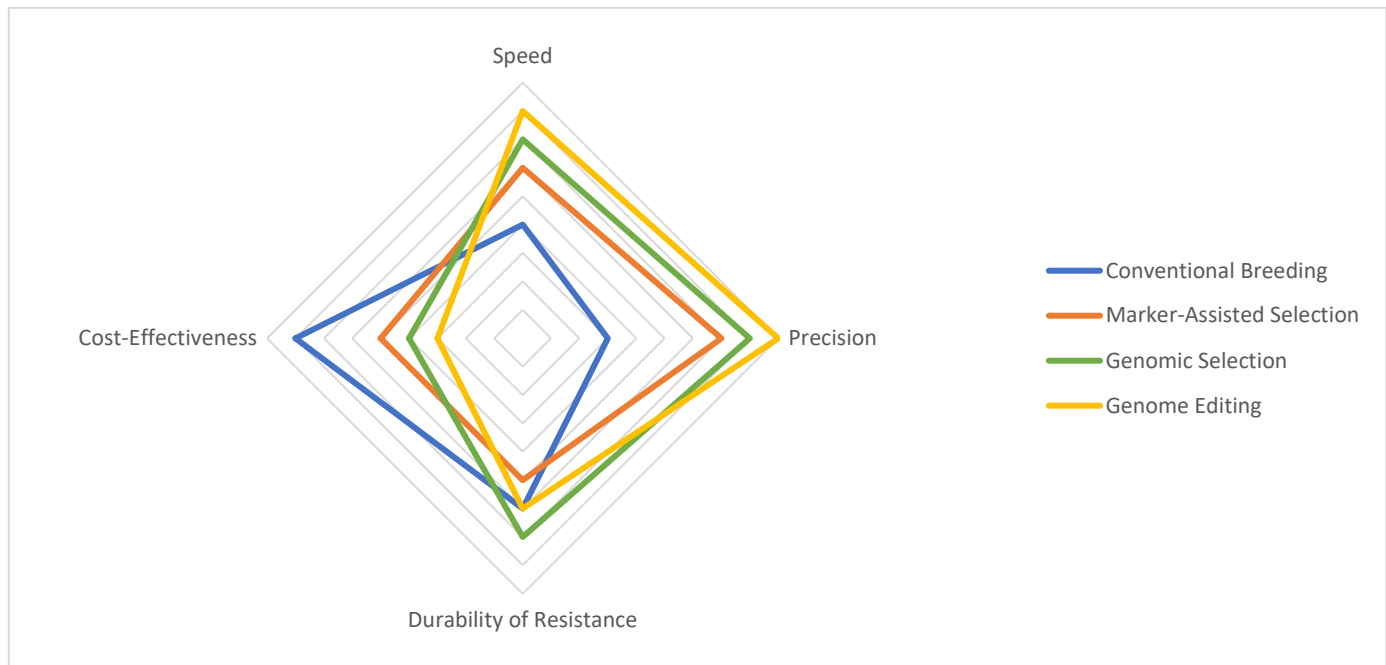


Figure 3. Comparison of breeding strategies for disease resistance based on speed, precision, durability, and cost-effectiveness.

### Gene Pyramiding and Multiline Varieties

Gene pyramiding involves combining multiple resistance genes in a single cultivar for broad-spectrum and durable resistance. This method may offer the advantage that it is less likely that resistance will break down since several defence mechanisms have to be countered simultaneously by pathogens. Sequential combination of QTLs conferring rice blast resistance has yielded long lasting, non-race-based resistance (Liu *et al.*, 2024). The combination of several genes conferring adult-plant resistance has delivered additive effects of stripe rust resistor in wheat (Younas *et al.*, 2024; Wang *et al.*, 2023). Multiline varieties are a combination of a variety of lines (each with different resistant genes) planted together as a single variety. Such genetic heterogeneity of a crop stand can delay epidemic development and enhance long-term stability of resistance and minimize the use of chemical pesticides (Tong *et al.*, 2024). Pyramiding multiple resistance genes has proven effective in delivering additive and durable resistance. Recent studies in wheat demonstrated that stacking adult-plant resistance genes substantially reduced stripe rust severity across diverse environments (Wang *et al.*, 2023). Such gene combinations provide broad spectrum protection, delaying pathogen adaptation as reported in Table 2.

### Quantitative Trait Loci (QTL) Mapping for Durable Resistance

The quantitative disease resistance (QDR) which is often regulated by several genes is more lasting than the qualitative resistance. The mapping of QTLs related to QDR makes it possible to apply MAS to enable selection of complex resistance characteristics by breeders. Crops that have benefited greatly due to QTL-based approaches in the improvement of resistance include wheat, barley, common bean, tomato, and pepper (Womack *et al.*, 2020). Another advantage linked to the incorporation of QTLs is that there is decreased risk of adaptation in pathogens due to the distribution of defence response to several genetic locations (Mungalu *et al.*, 2020). Breeders are mainly focusing on QTL mapping. For example, mapping QTLs for northern leaf blight (NLB) resistance in maize have been recently identified and deployed in breeding programs for incorporation into elite germplasm (Samantara *et al.* 2022). The importance of QTL mapping for complex and polygenic traits is evident (Table 2).

### Integration of Phenotypic and Genotypic Data

With the combination of high-quality phenotypic data and genotypic information, breeding outcomes could be predicted better. Genotyping-by-sequencing (GBS) and genome-wide association studies (GWAS) are some of the techniques that can aid in the identification of molecular markers associated with the traits of the resistance (Qureshi *et al.*, 2025). With the help of bioinformatics, these markers are also associated with the phenotypes observed and allow the superior selection of traits and a reduction in the length of breeding cycles (Kumar *et al.*, 2024). The integration of high-quality phenotypic data with genotyping-by-sequencing (GBS) and genome-wide association studies (GWAS) enabling superior trait prediction. Recent advances emphasize the utility of bioinformatics pipelines in linking markers with

resistance phenotypes, reducing breeding cycles and increasing efficiency (Derbyshire *et al.*, 2024) as elaborated in Table 2.

### Genome Editing

Genome editing technologies like CRISPR/Cas9, transcription activator-like effector nucleases (TALENs) and zinc-finger nucleases (ZFNs) have revolutionized agriculture with the targeted editing of the genes being targeted. These tools can insert, delete or edit disease resistance genes without excessive linkage drag and preserve useful agronomic characteristics (Sahito *et al.*, 2024; Erdoğan *et al.*, 2023).

Nevertheless, there are obstacles. Deployment can be prevented by off-target mutations, regulatory uncertainty and hazard to perception (Oguejiofor *et al.*, 2023). Countries have different regulatory regimes to genome-edited crops, which limit the pace and magnitude of uptake. Genome editing tools have delivered major breakthroughs in plant disease resistance. In wheat, CRISPR/Cas9-mediated knockout of *TaMLO* genes has conferred durable resistance against powdery mildew, while in rice, editing of *SWEET* susceptibility genes has significantly enhanced resistance to bacterial blight (Erdoğan *et al.*, 2023; Yıldırım *et al.*, 2023). These examples highlight the transformative role of precise genome engineering in modern resistance breeding Table 2.

### Utilization of Wild Relatives and Landraces

Wild crop relatives and traditional landraces can serve as very useful reservoirs of resistance breeding. They usually have resistance genes that have been lost in domestication. Allele mining, genomic selection, and chromosome segment substitution line (CSSL) development methods assist in the isolation and transfer of the resistance's traits of such sources (Huang *et al.*, 2023; Li *et al.*, 2023). However, introgression can be complex due to crossing barriers, chromosome incompatibility, and linkage drag, in which these undesirable traits come with useful alleles (Kashyap *et al.*, 2022). Careful breeding strategies and molecular tools are required to reduce these problems while maintaining yield and quality. Crop wild relatives and landraces remain invaluable reservoirs of resistance genes. For example, resistance alleles against stripe rust have been introgressed from *Aegilops tauschii* and other wild wheats into modern cultivars (Derbyshire *et al.*, 2024). Likewise, allele mining in Asian rice landraces continues to provide durable *Pi* genes against blast (Kaur *et al.*, 2022) as presented in Table 2. Figure 3 shows that genome editing is more efficient than other strategies in terms of speed, precision, and durability of resistance, though it is less cost-effective than conventional breeding (Mariam *et al.*, 2025). Genomic selection ranks high in durability and precision, while marker-assisted selection offers moderate performance across traits (Sharma *et al.*, 2024). Conventional breeding remains the most cost-effective but lags in speed and precision. Modern molecular approaches (genome editing, genomic selection) provide more precise and durable resistance compared to traditional methods (Pandey *et al.*, 2022).

## KEY CHALLENGES IN RESISTANCE BREEDING

### Balancing Resistance with Yield and Quality

Achieving disease resistance without loss of yield, quality, other essential agronomic traits remain one of the most consistent limitations in resistance breeding. Traits associated with resistance may in some cases be associated with reductions in yields, or poor-quality traits when they are introduced between wild relatives or landraces (Derbyshire *et al.*, 2024). A way to keep this trade-off in check is by using multi-trait selection mixed designs like index selection, tandem selection and independent culling. Genomic selection model with genetically correlated traits usage can provide a more advanced dual methodology of enhancing resistance levels and hence preserving the level or even the qualities of yield (Rehman *et al.*, 2024). One such case is a successful balancing gene, rice *Xa4* gene, leads to resistance to bacterial blight and enhances cell wall thickening due to which lodging resistance is also increased without a reduction in produce (Yuan *et al.*, 2024).

### Accelerating Breeding Cycles

The emerging pathogens evolve rapidly, and this requires a breeding response to be able to adjust to its evolution. Combined with the genomic selection, and marker-assisted breeding, speed breeding (SB) can even shorten breeding cycles to a fraction of the time (Çeran *et al.*, 2024). We can also use metabolomics-aided breeding to also deploy a biosensor to determine metabolite biomarkers that are associated with variability in resilience to stress (Mashabela *et al.*, 2022). However, these will require control-environment space, precision of environmental manipulations as well as intense technical know-how (Potts *et al.*, 2023).

### Infrastructure, Funding, and Resource Limitations

As we have seen so far, resistance breeding requires long term investment in infrastructure as well. Such as controlled growth chamber, well advanced phenotyping systems, cutting edge bioinformatics systems. Much of the breeding programs, especially those in developing nations, cannot afford to have multi-year selection pipelines (Dong and Fan,

2024). In the case of promising technologies, such as CRISPR/Cas9, the bottleneck commonly resides in the fact that the lab capacity is not sufficient, and the projects need funding to expand into significant breeding programs. Environmental regulation, climatic patterns of change and shifts in land use are matters that also limit the operation of global breeding networks disrupting coordinated deployment strategies of genes (Lallawmkimi *et al.*, 2024; Xing and Wang, 2024).

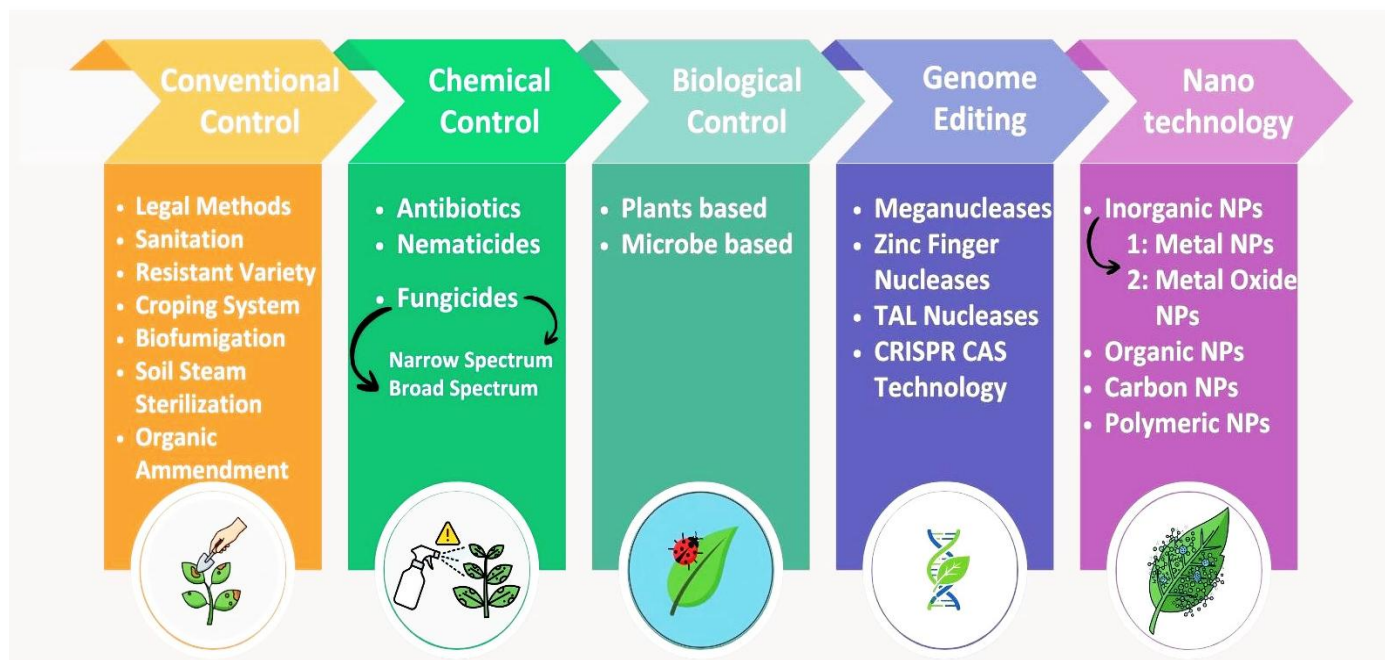


Figure 4. Overview of plant disease control methods, ranging from conventional practices and chemical applications to biological control, genome editing, and nanotechnology-based approaches

### Regulatory Hurdles and Public Perception

The regulatory approval system is complex, lengthy and even costly in case of the varieties developed by biotechnology (Hallerman *et al.*, 2024). In other jurisdictions, non-harmonized procedures and divergent biosafety guidelines slow the passage to release of resistant cultivars. The use of misinformation and widespread scepticism on the part of the population towards genetically modified (GM) crops also restricts adoption. To find solutions to these problems, direct communication, informational campaigns, and the presentation of the positive effects of decreasing the use of pesticides on the environment are needed (Kotur *et al.*, 2024).

Non-transgenic approaches such as cis-genesis and precise genome editing may help overcome public resistance by avoiding the transfer of genes from unrelated species (Lironcurti *et al.*, 2023). However, genome editing faces uncertain regulatory status in many countries, necessitating adaptive policies that distinguish it from traditional GM technology (Wray-Cahen *et al.*, 2024).

## INTEGRATED MANAGEMENT FOR DURABLE RESISTANCE

### Integrated Pest Management (IPM)

Although genetic resistance is an effective means of combatting plant pathogens, it works best when included in larger management systems that integrate multiple approaches (Figure 4). IPM involves use of resistant varieties integrated with complementary controlling practices including cultural, biological and chemical control methods in a bid to minimize the influence of the pathogen and postponement of resistance degradation. The rise of biotechnology fueled by genetically modified insect-resistant crops including Bt cotton and Bt maize has substantially curbed the use of pesticides while also ensuring that production remains high (Jan *et al.*, 2025). Crop rotation, field sanitation and destruction of the alternative hosts can break the life cycle of the pathogen, and reduce the pathogen load (Gyawali *et al.*, 2023). In cases when chemical interventions are required, they are used in a way that will not have much negative effect on the environment, and their use will reduce the pace at which pesticide resistance occurs (Galli *et al.*, 2024).

### Disease Forecasting and Early Warning Systems

The ability to forecast diseases and early warning systems are essential to safeguard against emerging pathogens - especially those that spread quickly. Forecasting is more effective when it is associated with the full range of control

techniques available (Figure 4). You can predict outbreaks with these systems. Also, you can use them to guide early interventions using models and reports (Sumithra et al., 2025). When new diseases are discovered more quickly, it helps to prevent epidemics on a larger scale and further limit the economic costs and damage to the environment in response to any emergency. By putting together weather-based forecasting models along with inspecting the pathogen population has gotten better rust disease management in wheat-growing regions (Vashisht et al., 2023).

Recent case studies highlight these advances. The identification of more than 40 different races of a pathogenic fungus strain, *Puccinia striiformis*, in Egypt is indicative of the scale of the threat and the grave risk that pathogen diversity poses to agricultural crops around the world. Esmail et al. (2025) note the discovery of these races in over 40 states in Egypt. This is a good example of the need for continuous monitoring of pathogens that threaten agricultural crops around the world. Comparable research on *Puccinia triticina* (wheat leaf rust) has found the emergence of new virulent races after 2020 that quickly overcome the effectiveness of previously effective resistance genes (Omara et al. 2021). The examples highlight the need for integrated forecasting platforms connecting pathogen surveillance with resistance deployment strategies.

### Agroecological Approaches

This supports the notion that when breeding for crop resistance takes place within agroecological farming systems, the result will be sustainable and resilient crop production. Biodiversity, ecosystem services, and resource efficiency are key goals of agroecology which can be enhanced particularly using genetic varieties and systems of rotations (Akanmu et al., 2023). These systems can be supported by developing low-input varieties with multi-stress resistance. It is also important to ensure compatibility with organic and conservation agriculture. Intercropping or agroforestry combined with resistance breeding can limit the spread of pathogens besides improving soil health and moisture retention. These agroecological strategies complement other disease control methods highlighted in Figure 4, ensuring sustainability and resilience. Agroecological strategies can be effective when combined with resistance breeding. For instance, deploying varietal mixtures has been shown to slow down the breakdown of resistance in cereals by reducing uniform selection pressure on pathogen populations (Yazdani et al., 2025). Such approaches ensure that resistance remains durable while supporting ecosystem resilience.

## FUTURE PERSPECTIVES

### Role of Omics and Advanced Phenotyping

Breeding and resistance are being transformed by advances in genomics, transcriptomics proteomics and metabolomics. Such omics technologies can be used to identify host resistance genes and regulatory networks in response to both biotic and abiotic stresses, and how host-pathogen interactions occur (Naqvi et al., 2024), these regulatory networks often map onto signaling pathways such as those depicted in Figure 2. The combination of the multi-omics data can help in more specific selecting and decrease the time of resilient cultivar development (Haidar et al., 2024). Artificial intelligence and high-throughput imaging High-throughput phenotyping (HTP) increases breeding efficiency. LiDAR, hyperspectral, and thermal-sensing platforms may be used to assess disease symptoms, canopy structure, and stress response within both real-world and experimental settings (Cembrowska-Lech et al., 2023). These tools, when combined with genomic information, have the potential to fill the genotype-phenotype gap and increase accuracy of predicting complex traits related to resistance (Nguyen et al., 2023).

Recent advancements emphasize the increasing utilization of AI and ML in resistance breeding. Several simulation tools have been developed in recent years, which were not available at the beginning of genomic selection. For instance, using deep learning models trained on hyperspectral and phenotypic datasets for wheat and rice has significantly improved the prediction of disease resistance over traditional methods. Combining omics, phenotyping, and AI is reducing breeding cycle time and increasing prediction efficiency (Ma et al., 2025; Farooq et al., 2024).

### Advances in Pathogen Detection Technologies

It is important to identify the pathogens early and correctly to manage the pathogens. By using metagenomics, it became possible to recognize known as well as unknown pathogens in one sample at a time and without the prior hypothesis regarding a sample by using next-generation sequencing (Jabran et al., 2024). The introduction of nano-based biosensors and Raman spectroscopy into agricultural supply chain is being developed to provide real-time monitoring of diseases (Oon et al., 2023; Bhowmik et al., 2025). Using lightweight diagnostics, like loop-mediated isothermal amplification (LAMP) microchips, which are quick, sensitive, and can be performed on-site, can aid in the deployment of these kinds of diagnostics in low resource settings (Yadav and Yadav, 2025). New approaches will also be used wastewater surveillance to monitor community pathogens and bacteriophage-sensor

solutions to detect plant pathogens in a short period (Panhwar *et al.*, 2023). Early detection and intervention can be done with the help of these technologies, minimizing economic losses and inhibited pathogen spread.

Continuous genomic surveillance networks are increasingly important. By combining real-time metagenomics with global data-sharing platforms, outbreaks of rust fungi and viral pathogens can now be predicted with higher accuracy (Lappan *et al.*, 2024). This integration allows resistance breeding programs to anticipate pathogen shifts rather than react after resistance breakdown.

### **International Collaboration and Germplasm Exchange**

International collaboration is crucial for management of emerging plant pathogens. Plant disease diagnostic networks, genomic surveillance networks, and germplasm exchange networks allow the rapid distribution of resistant varieties across countries (Engels *et al.*, 2024; Islam, 2024). Collaboration networks in breeding-networks with open access, resistance breeding can be accelerated more speedily than in other breeding networks, as seen in cases like those facilitated by CGIAR. Nonetheless, some issues persist, which include phytosanitary limitations, intellectual property rights, and inconsistent diagnostic practices (Jha *et al.*, 2023).

Recent collaborative initiatives emphasize not only germplasm exchange but also open-access breeding databases that integrate multi-omics and phenotypic data globally (Lappan *et al.*, 2024). Such platforms allow breeders in resource-poor regions to access advanced tools and materials, increasing equity in resistance breeding.

### **Policy Support and Public–Private Partnerships**

The regulatory frameworks must be in tandem with technological innovations. Genome edited crops, specifically through cisgenesis or precision editing technologies, should be assessed at science-based, proportionate regulations to prevent the occurrence of unjust delays (Dowd-Urbe *et al.*, 2023). A clear policy definition between conventional GM crops and targeted genome edits will lead to increased levels of trust and adoption. Partnerships between the public and private sectors can help deploy research innovations. According to Van Asseldonk *et al.* (2023) Public-private partnerships can align the incentive of both the public research organizations and the seed companies, thus enabling faster access to resistant varieties by growers especially smallholders. Partnerships between varieties that address the environmental and market needs of the specific locality of the technology are needed (Zuberi *et al.*, 2023). Many recent PPP initiatives have enhanced the delivery of resistant cultivars in Africa and South Asia by linking CGIAR centers with regional seed companies (Yazdani *et al.*, 2025). Such partnerships illustrate how innovative pipelines can align with local agroecological and market opportunities.

## **CONCLUSION**

Emerging plant pathogens threaten global food security due to changing climatic conditions, rapid globalization and intensification of agriculture. The rapid evolution of plant pathogens is overcoming host resistance in a few cropping seasons. Therefore, it is essential to have resilient breeding methods. Development of resistant breeding techniques is critical in sustainable disease management. Modern molecular breeding techniques such as genomic selection, genome editing and use of wild relatives in breeding have provided more tools for sustainable breeding. Combining these modern tools with standard breeding techniques enables development of durable resistance providing high yield and quality traits. Resistant breeding with integrated pest management techniques enforced with early warning systems and agroecology provides even better durability in breeding. However, challenges persist in such breeding approaches are balancing desired resistant traits with yield and quality traits, regulatory hurdles of genetically modified crops and consumer perception of genetically modified crops. To overcome these challenges, it is necessary to develop specific policies, develop public private partnerships and open communications for building trust and acceptance. Besides that, rapid pathogen detection, use of omics technologies and novel phenotyping techniques should be used to control emerging plant pathogens. Following these management approaches, plant breeders, pathologists and policymakers can keep pace with emerging plant pathogens and protect global food security.

## **AUTHOR CONTRIBUTIONS**

All authors contributed to the conception and design of the study. Data collection, and literature review were mainly performed by Jabir Asghar, Ameer Jan, Muhammad Hamayun, Khatir Ali, Atiq u Rehman, Mushtaq Ahmed, Izaz Ullah, Abdul Basit, and Shaikh Saddam. The first draft of the manuscript was prepared by Jabir Asghar, and all authors contributed to manuscript revision. Jabir Asghar and Ameer Jan were responsible for formal analysis and visualization. Critical revisions, validation, and editing were provided by Muhammad Hamayun, Khatir Ali, Atiq u Rehman, Mushtaq Ahmed, Izaz Ullah, Abdul Basit, and Shaikh Saddam. All authors read and approved the final manuscript.

## CONFLICT OF INTEREST

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## ACKNOWLEDGEMENTS

We are grateful to Vice Chancellor Prof. Dr. Mumtaz Baloch University of Makran, Panjgur for Encouraging and Supporting in this Scientific write up.

## REFERENCES

- Ajayi, T., Lacombe, J., Ince, G., & Yeats, T. (2024). Analytical prediction of genetic contribution across multiple recurrent backcrossing generations. *Theoretical and Applied Genetics*, 137(12), 412.
- Akanmu, A. O., Kutu, F. R., Babalola, O. O., Akol, A. M., & Ndolo, D. O. (2023). Agroecological techniques: adoption of safe and sustainable agricultural practices among the smallholder farmers in Africa. *Frontiers in Sustainable Food Systems*, 7, 1143061.
- Alfizar, A., & Nasution, S. S. (2024). The explosion of pests and diseases due to climate change. *IOP Conference Series: Earth and Environmental Science*, 1297(1), 012072.
- Anand, A., Subramanian, M., & Kar, D. (2023). Breeding techniques to dispense higher genetic gains. *Frontiers in Plant Science*, 13, 1491.
- Anwer, M., Ikram, M. T., Hanif, M., Afzal, T., Javed, A., Murtaza, M. A., Yasir, M. A., Tarafder, E., Adnan, M., & Ahmed, R. (2024). Evaluation of biochemical methods for identification of *Xanthomonas citri* pv. malvacearum causing bacterial blight of cotton. *Plant and Environment*, 5(1), 25–34.
- Bhowmik, D., Rickard, J. J. S., Jelinek, R., & Goldberg Oppenheimer, P. (2025). Resilient sustainable current and emerging technologies for foodborne pathogen detection. *Sustainable Food Technology*, 3(1), 10–31.
- Boutigny, A.-L., Ballois, N., Iancu, G., Ward, T. J., & Ios, R. (2013). Diversity of the *Fusarium graminearum* species complex on French cereals. *European Journal of Plant Pathology*, 138(1), 133–148.
- Brooker, R. W., Iannetta, P. P. M., Karley, A. J., Renard, D., & Hawes, C. (2023). Plant diversity and ecological intensification in crop production systems. *Journal of Plant Ecology*, 16(6), ryad072.
- Çelik Oğuz, A., & Karakaya, A. (2021). Genetic Diversity of Barley Foliar Fungal Pathogens. *Agronomy*, 11(3), 434.
- Cembrowska-Lech, D., Mikiciuk, G., Mikiciuk, M., Miller, T., Adamski, C., Radaczyńska, M., Krzemińska, A., & Nowakowska, A. (2023). An Integrated Multi-Omics and Artificial Intelligence Framework for Advance Plant Phenotyping in Horticulture. *Biology*, 12(10), 1298.
- Ćeran, M., Glogovac, S., Trkulja, D., Đorđević, V., Miladinović, D., Kondić-Špika, A., & Radanović, A. (2024). Genomics-assisted speed breeding for crop improvement: present and future. *Frontiers in Sustainable Food Systems*, 8, 1329596.
- Cha, J.-K., Park, H., Lee, S.-M., Lee, J.-H., Kwon, S.-W., Kwon, Y., & Jang, S.-G. (2024). Synergizing breeding strategies via combining speed breeding, phenotypic selection, and marker-assisted backcrossing for the introgression of Glu-B1i in wheat. *Frontiers in Plant Science*, 15, 1394348.
- Chen, J.-R., Armijos-Jaramillo, V., Aguirre-Carvajal, K., Xue, D.-Y., Lin, Y.-P., Arone-Maxwell, L., Chang, H.-C., & Oliva, R. (2024). Exploring the genetic makeup of *Xanthomonas* species causing bacterial spot in Taiwan: evidence of population shift and local adaptation. *Frontiers in Microbiology*, 15, 1439074.
- Combala, M., Pita, J. S., Gbonamou, M., Samura, A. E., Amoakon, W. J.-L., Kouakou, B. S. M., Onile-Ere, O., Sawadogo, S., Eboulem, G. R., Otron, D. H., Seka, J. S. S., Eni, A., Ndougou, C., & Tiendrébéogo, F. (2024). An Alarming Eastward Front of Cassava Mosaic Disease in Development in West Africa. *Viruses*, 16(11), 1691.
- Derbyshire, M. C., Newman, T. E., Thomas, W. J. W., Batley, J., & Edwards, D. (2024). The complex relationship between disease resistance and yield in crops. *Plant Biotechnology Journal*, 22(9), 2612–2623.
- Dong, G., & Fan, Z. (2024). CRISPR/Cas-mediated germplasm improvement and new strategies for crop protection. *Crop Health*, 2(1), 100021.
- Dowd-Urbe, B., Uminsky, D., Rock, J. S., Spreadbury, T., & Chiril, P. (2023). Bridging the gap? Public-private partnerships and genetically modified crop development for smallholder farmers in Africa. *PLANTS, PEOPLE, PLANET*, 6(2), 437–451.
- Dracatos, P. M., Lu, J., Sánchez-Martín, J., & Wulff, B. B. H. (2023). Resistance that stacks up: engineering rust and mildew disease control in the cereal crops wheat and barley. *Plant Biotechnology Journal*, 21(10), 1938–1951.
- Engels, J. M. M., Ebert, A. W., & Van Hintum, T. (2024). Collaboration between Private and Public Genebanks in Conserving and Using Plant Genetic Resources. *Plants*, 13(2), 247.
- Erdoğan, İ., Cevher-Keskin, B., Bilir, Ö., Hong, Y., & Tör, M. (2023). Recent Developments in CRISPR/Cas9 Genome-Editing Technology Related to Plant Disease Resistance and Abiotic Stress Tolerance. *Biology*, 12(7), 1037.

- Esmail, S. M., Draz, I. S., Sehsah, M. D., Saad-El-Din, H. I., Youssef, W. A., & Komeil, D. A. (2025). Evolutionary and dispersal dynamics in invasive races of *Puccinia striiformis* f. sp. *tritici* attributed to virulence and SCAR markers. *Physiological and Molecular Plant Pathology*, 136, 102524.
- Fan, H., Bao, X., Miao, R., He, W., Zhao, C., Sun, Y., & Guo, C. (2025). Research on the Effect of Diversified Cropping on Crop Quality: A Review. *Agriculture*, 15(5), 456.
- Farooq, M. A., Gao, S., Hassan, M. A., Huang, Z., Rasheed, A., Hearne, S., Prasanna, B., Li, X., & Li, H. (2024). Artificial intelligence in plant breeding. *Trends in Genetics: TIG*, 40(10), 891–908.
- Fry, W. E., Birch, P. R. J., Judelson, H. S., Grünwald, N. J., Danies, G., Everts, K. L., & Smart, C. D. (2015). Five reasons to consider *Phytophthora infestans* a reemerging pathogen. *Phytopathology*, 105(7), 966–981.
- Gai, Y., & Wang, H. (2024). Plant Disease: A Growing Threat to Global Food Security. *Agronomy*, 14(8), 1615.
- Galli, M., Feldmann, F., Vogler, U. K., & Kogel, K.-H. (2024). Can biocontrol be the game-changer in integrated pest management? A review of definitions, methods and strategies. *Journal of Plant Diseases and Protection*, 131(2), 265–291.
- Gyawali, P., Bohara, K., Rijal, S., Karki, N., & Shahi, J. (2023). A comprehensive review on integrated pest management of melon fruit fly (*Bactrocera cucurbitae*). *International Journal of Pest Management*, ahead-of-print(ahead-of-print), 1–9.
- Haidar, S., Hooker, J., Lackey, S., Elian, M., Puchacz, N., Szczyglowski, K., Marsolais, F., Golshani, A., Cober, E. R., & Samanfar, B. (2024). Harnessing Multi-Omics Strategies and Bioinformatics Innovations for Advancing Soybean Improvement: A Comprehensive Review. *Plants (Basel, Switzerland)*, 13(19), 2714.
- Hallerman, E., Karembu, M., Walton, M., Tizard, M., Kovich, D., Rocha-Salavarieta, P. J., Wray-Cahen, D., Camargo, L. S. A., Bredlau, J., Romero-Aldemita, R., Muia, A. N., Murrone, M. L., & Dagli, M. L. Z. (2024). Enabling regulatory policy globally will promote realization of the potential of animal biotechnology. *CABI Agriculture and Bioscience*, 5(1), 28.
- Hatta, M. A. M., Arora, S., Ghosh, S., Matny, O., Smedley, M. A., Yu, G., Chakraborty, S., Bhatt, D., Xia, X., Steuernagel, B., Richardson, T., Mago, R., Lagudah, E. S., Patron, N. J., Ayliffe, M., Rouse, M. N., Harwood, W. A., Periyannan, S., Steffenson, B. J., & Wulff, B. B. H. (2020). The wheat Sr22, Sr33, Sr35 and Sr45 genes confer resistance against stem rust in barley. *Plant Biotechnology Journal*, 19(2), 273–284.
- Hossain, M. M., Sultana, F., Mostafa, M., Ferdus, H., Rahman, M., Rana, J. A., Islam, S. S., Adhikary, S., Sannal, A., Al Emran Hosen, M., Nayeema, J., Emu, N. J., Kundu, M., Biswas, S. K., Farzana, L., & Al Sabbir, M. A. (2024). Plant disease dynamics in a changing climate: impacts, molecular mechanisms, and climate-informed strategies for sustainable management. *Discover Agriculture*, 2(1), 32.
- Huang, K., Boniface, M.-C., Duhnen, A., Mangin, B., Muñoz, S., Gouzy, J., Owens, G. L., Donnadiou, C., Carrere, S., Cauet, S., Legendre, A., Staton, S. E., Lázaro-Guevara, J. M., Jahani, M., Poncet, C., Bellec, A., Gautier, V., Mandel, T., Langlade, N., & Hübner, S. (2023). The genomics of linkage drag in inbred lines of sunflower. *Proceedings of the National Academy of Sciences of the United States of America*, 120(14), e2213710120.
- Isiyel, M., İlhan, E., Kasapoğlu, A. G., Muslu, S., Öner, B. M., Aygören, A. S., Yiğider, E., Aydın, M., & Yıldırım, E. (2024). Identification and characterization of *Phaseolus vulgaris* CHS genes in response to salt and drought stress. *Genetic Resources and Crop Evolution*, 72(1), 271–293.
- Islam, T. (2024). Genomic surveillance for tackling emerging plant diseases, with special reference to wheat blast. *CABI Reviews*, 2024, 038.
- Jabran, M., Ijaz, M., Ali, M. A., Ali, F., Hussain, S., Gao, L., Muhae-Ud-Din, G., Zahoor, A., & Muzammil, S. (2024). Exploring the potential of nanomaterials (NMs) as diagnostic tools and disease resistance for crop pathogens. *Chemical and Biological Technologies in Agriculture*, 11(1), 22.
- Jan, A., Hussain, Z., Ullah, A., Ahmed, Z., Bakhsh, B. P., Latif, A., & Ahmed, M. (2025). Sugarcane Whip Smut: A Comprehensive Review of Pathogen Biology, Epidemiology, and Control Measures. *Annual Methodological Archive Research Review*, 3(5), 211–232.
- Jha, U. C., Nayyar, H., Chattopadhyay, A., Beena, R., Lone, A. A., Naik, Y. D., Thudi, M., Prasad, P. V. V., Gupta, S., Dixit, G. P., & Siddique, K. H. M. (2023). Major viral diseases in grain legumes: designing disease resistant legumes from plant breeding and OMICS integration. *Frontiers in Plant Science*, 14(1), 1214088.
- Kantor, C., Eisenback, J. D., & Kantor, M. (2024). Biosecurity risks to human food supply associated with plant-parasitic nematodes. *Frontiers in Plant Science*, 15, 1407357.
- Kashyap, A., Ha, P. T. T., Rao, M., Sharma, J., Gupta, N. C., Mason, A. S., Bhattacharya, R. C., Garg, P., & Tanwar, K. (2022). Strategies for utilization of crop wild relatives in plant breeding programs. *Theoretical and Applied Genetics*, 135(12), 4151–4167.
- Khan, N. (2024). Unlocking Innovation in Crop Resilience and Productivity: Breakthroughs in Biotechnology and Sustainable Farming. *Innovation Discovery*, 1(4), 28.
- Kolur, S. M., Sampath, V., Burud, A., Satapathy, S. N., Chellem, S. R., Ramasamy, M., & Rout, A. (2024). A Review on Biotech Innovations in Seed Technology for Robust Crop Production. *Journal of Advances in Biology & Biotechnology*, 27(5), 535–550.
- Kumar, D., & Mukhopadhyay, R. (2024). Climate change and plant pathogens: Understanding dynamics, risks and mitigation strategies. *Plant Pathology*, 74(1), 59–68.

- Kumar, K. P., & Walia, P. (2024). Speed Breeding in Cereal Crops: Accelerating Genetic Improvement for Rapid Agricultural Advancement. *Journal of Experimental Agriculture International*, 46(6), 465–477.
- Kumar, R., Das, R., Verma, R., Mishra, V. K., Das, S. P., Devi, H. L., Das, B., Das, B., Bhattacharjee, B., Kumar, A., Chakraborti, M., Choudhury, B. U., Rawat, S., Prakash, N. R., & Devi, A. G. (2024). Advances in genomic tools for plant breeding: harnessing DNA molecular markers, genomic selection, and genome editing. *Biological Research*, 57(1), 28.
- Kurabachew, H., & Ayana, G. (2017). Bacterial Wilt caused by *Ralstonia solanacearum* in Ethiopia: Status and Management Approaches: A Review. *International Journal of Phytopathology*, 5(3), 107–119.
- Lallawmkimi, M. C., Kumar, M., Rout, A., P, A., Veda, D., Yadav, A., & B, D. (2024). Innovative Approaches in Crop Genetic Engineering for Sustainable Agriculture: A Review. *Journal of Advances in Biology & Biotechnology*, 27(8), 615–631.
- Lappan, R., Chown, S. L., French, M., Perlaza-Jiménez, L., Macesic, N., Davis, M., Brown, R., Cheng, A., Clasen, T., Conlan, L., Goddard, F., Henry, R., Knight, D. R., Li, F., Luby, S., Lyras, D., Ni, G., Rice, S. A., Short, F., Song, J., Whittaker, A., Leder, K., Lithgow, T., & Greening, C. (2024). Towards integrated cross-sectoral surveillance of pathogens and antimicrobial resistance: Needs, approaches, and considerations for linking surveillance to action. *Environment International*, 192, 109046.
- Li, H., Zhu, L., Fan, R., Li, Z., Liu, Y., Shaheen, A., Nie, F., Li, C., Liu, X., Li, Y., Liu, W., Yang, Y., Guo, T., Zhu, Y., Bu, M., Li, C., Liang, H., Bai, S., Ma, F., & Song, C.-P. (2023). A platform for whole-genome speed introgression from *Aegilops tauschii* to wheat for breeding future crops. *Nature Protocols*, 19(2), 281–312.
- Liu, B., Liu, C., Hu, H., Wang, J., Chen, J., Yang, J., Dong, J., Zhao, J., Zhou, L., Zhang, S., Nie, S., Yang, T., Yang, W., Jiang, X., Zhu, X., Ma, Y., & Ning, Y. (2024). Pangenome-Wide Association Study and Transcriptome Analysis Reveal a Novel QTL and Candidate Genes Controlling both Panicle and Leaf Blast Resistance in Rice. *Rice*, 17(1), 44.
- Ma, J., Cheng, Z., & Cao, Y. (2025). Artificial Intelligence-Assisted Breeding for Plant Disease Resistance. *International Journal of Molecular Sciences*, 26(11), 5324.
- Mapari, A. R., & Mehandi, S. (2024). Enhancing Crop Resilience: Advances and Challenges in Marker-Assisted Selection for Disease Resistance. *Journal of Advances in Biology & Biotechnology*, 27(7), 569–580.
- Marium, A., Arshad, H., Murtaza, B., Aziz, A., Gul, A., Tariq, S., Naheed, N., Aslam, H., Jabeen, A., & Khalid, M. (2025). Crispr/Cas genome editing to enhance resistance leading to enhanced productivity in crops. *Kashf Journal of Multidisciplinary Research*, 2(02), 53–89.
- Mashabela, M. D., Masamba, P., & Kappo, A. P. (2022). Metabolomics and Chemoinformatics in Agricultural Biotechnology Research: Complementary Probes in Unravelling New Metabolites for Crop Improvement. *Biology*, 11(8), 1156.
- Michel, S., Steiner, B., Buerstmayr, M., Samad-Zamini, M., Nosenko, T., Wagner, C., Mayer, K., & Buerstmayr, H. (2021). Merging Genomics and Transcriptomics for Predicting *Fusarium* Head Blight Resistance in Wheat. *Genes*, 12(1), 114.
- Midega, C. A. O., Wasonga, C. J., Hooper, A. M., Pickett, J. A., & Khan, Z. R. (2017). Drought-tolerant *Desmodium* species effectively suppress parasitic *striga* weed and improve cereal grain yields in western Kenya. *Crop Protection*, 98, 94–101.
- Mihrete, T. B., & Mihretu, F. B. (2025). Crop Diversification for Ensuring Sustainable Agriculture, Risk Management and Food Security. *Global Challenges (Hoboken, NJ)*, 9(2), e2400185.
- Mungalu, H., Hamabwe, S., Kamfwa, K., Mukuma, C., Kelly, J. D., Sansala, M., & Gepts, P. (2020). Identification of race-specific quantitative trait loci for resistance to *Colletotrichum lindemuthianum* in an Andean population of common bean. *Crop Science*, 60(6), 2843–2856.
- Nalley, L., Tsiboe, F., Durand-Morat, A., Shew, A., & Thoma, G. (2016). Economic and Environmental Impact of Rice Blast Pathogen (*Magnaporthe oryzae*) Alleviation in the United States. *PLoS ONE*, 11(12), e0167295.
- Naqvi, R. Z., Mahmood, M. A., Amin, I., Mansoor, S., & Asif, M. (2024). Omics-driven exploration and mining of key functional genes for the improvement of food and fiber crops. *Frontiers in Plant Science*, 14, 1305417.
- Nguyen, C., Bhadra, S., Moose, S., & Sagan, V. (2023). UAV Multisensory Data Fusion and Multi-Task Deep Learning for High-Throughput Maize Phenotyping. *Sensors*, 23(4), 1827.
- Nitta, A., Rakesh, T., Natarajan, V., & Reddy, A. J. (2024). Impact of Climate Change on Pest Biology, Behaviour and Their Distributions. *International Journal of Environment and Climate Change*, 14(4), 46–56.
- Oguejiofor, B., Abioye, K., Alabi, A., Omotosho, A., Oguntoyinbo, F., Daraojimba, A., & Daraojimba, C. (2023). A REVIEW ON DATA-DRIVEN REGULATORY COMPLIANCE IN NIGERIA. *International Journal of Applied Research in Social Sciences*, 5(8), 231–243.
- Omara, R. I., Nehela, Y., Mabrouk, O. I., & Elsharkawy, M. M. (2021). The Emergence of New Aggressive Leaf Rust Races with the Potential to Supplant the Resistance of Wheat Cultivars. *Biology*, 10(9), 925.
- Oon, Y.-L., Deng, M., Song, K., Li, L., Ayaz, M., & Oon, Y.-S. (2023). Waterborne pathogens detection technologies: advances, challenges, and future perspectives. *Frontiers in Microbiology*, 14, 1240366.
- Oraon, S., Sinha, D., G S, S., Karanwal, R., Padamini, R., Das, N., Shahni, Y. S., & Singh, O. B. (2024). Impact of Emerging Pathogens in Crop Production. *Microbiology Research Journal International*, 34(7), 80–92.

- Pandey, S., Parida, S. K., Singh, A., & Prasad, M. (2022). Combining speed breeding with traditional and genomics-assisted breeding for crop improvement. *Plant Breeding*, 141(3), 301–313.
- Panhwar, S., Keerio, H. A., Ilhan, H., Boyaci, I. H., & Tamer, U. (2023). Principles, Methods, and Real-Time Applications of Bacteriophage-Based Pathogen Detection. *Molecular Biotechnology*, 66(11), 3059–3076.
- Pedrozo, R., Osakina, A., Huang, Y., Nicolli, C. P., Wang, L., & Jia, Y. (2025). Status on Genetic Resistance to Rice Blast Disease in the Post-Genomic Era. *Plants (Basel, Switzerland)*, 14(5), 807.
- Potts, J., Michael, V. N., Jangra, S., & Wu, X. (2023). Speed Breeding for Crop Improvement and Food Security. *Crops*, 3(4), 276–291.
- Qi, Y., Zhao, H., Zhang, Z., Ouyang, Y., & Zhang, X. (2025). Biological Characteristics and Fungicide Screening of *Bipolaris oryzae* Causing Leaf Spot on Banana in China. *Microorganisms*, 13(6), 1285.
- Qureshi, N., Singh, R. P., & Bhavani, S. (2025). Genetic Dissection of Triple Rust Resistance (Leaf, Yellow, and Stem Rust) in Kenyan Wheat Cultivar, “Kasuku”. *Plants (Basel, Switzerland)*, 14(7), 1007.
- Rehman, M., Mubeen, S., Chen, P., Saeed, W., Cao, S., Luo, D., & Pan, J. (2024). Progress in Agronomic Crops Lodging Resistance and Prevention: A Review. *Journal of Agronomy and Crop Science*, 210(6), e13023.
- Romeo Lironcurti, S., Demaria, F., Sardone, R., & D’Annolfo, R. (2023). Consumer Evaluations of and Attitudes towards New Genome Editing Techniques: An Italian Case Study. *Agriculture*, 14(1), 51.
- Sabag, I., Bi, Y., Sahoo, M. M., Herrmann, I., Morota, G., & Peleg, Z. (2024). Leveraging genomics and temporal high-throughput phenotyping to enhance association mapping and yield prediction in sesame. *The Plant Genome*, 17(3), e20436.
- Sahito, J. H., Gishkori, Z. G. N., Ma, C., Wang, Z., Zhang, H., Tang, J., Ding, D., & Zhang, X. (2024). Advancements and Prospects of Genome-Wide Association Studies (GWAS) in Maize. *International Journal of Molecular Sciences*, 25(3), 1918.
- Shah, S. M. A., Haq, F., Huang, K., Wang, Q., Liu, L., Li, Y., Wang, Y., Khan, A., Yang, R., Khojasteh, M., Xu, X., Xu, Z., & Chen, G. (2025). Two TAL effectors of *Xanthomonas citri* pv. *malvacearum* induce water soaking by activating *GhSWEET14* genes in cotton. *Molecular Plant Pathology*, 26(1), e70053.
- Shamshad, A., Rashid, M., Hameed, A., & Arshad, H. M. I. (2024). Identification of biochemical indices for brown spot (*Bipolaris oryzae*) disease resistance in rice mutants and hybrids. *PLOS ONE*, 19(4), e0300760.
- Sharma, P., Doultani, S., Highland, H., George, L., & Hadiya, K. K. (2024). Overview of Marker-assisted Selection in Animal Breeding. *Journal of Advances in Biology & Biotechnology*, 27(5), 303–318.
- Shattock, R. C. (2002). *Phytophthora infestans*: populations, pathogenicity and phenylamides. *Pest Management Science*, 58(9), 944–950.
- Singh, R. P., Hodson, D. P., Jin, Y., Lagudah, E. S., Ayliffe, M. A., Bhavani, S., & Hovmøller, M. S. (2015). Emergence and spread of new races of wheat stem rust fungus: continued threat to food security and prospects of genetic control. *Phytopathology*, 105(7), 872–884.
- Singh, R., Ryu, J., Hyoung Lee, W., Kang, J.-H., Park, S., & Kim, K. (2024). Wastewater-borne viruses and bacteria, surveillance and biosensors at the interface of academia and field deployment. *Critical Reviews in Biotechnology*, 45(2), 413–433.
- Soko, T., Prins, R., Pretorius, Z. A., & Bender, C. M. (2018). Yield Loss Associated with Different Levels of Stem Rust Resistance in Bread Wheat. *Plant Disease*, 102(12), 2531–2538.
- Song, L., Zhang, A., Yang, X., Wang, R., & Liu, D. (2023). Molecular Markers and Their Applications in Marker-Assisted Selection (MAS) in Bread Wheat (*Triticum aestivum* L.). *Agriculture*, 13(3), 642.
- Sumithra, S., Jancee, B. V., Mohankumar, N., Murugan, S., Radhika, M., Venkatesh, G., & Lakshmi, B. S. (2025). Deep learning for infectious disease surveillance integrating internet of things for rapid response. *International Journal of Electrical and Computer Engineering (IJECE)*, 15(1), 1175–1187.
- Švara, A., De Storme, N., Carpentier, S., Keulemans, W., & De Coninck, B. (2024). Phenotyping, genetics, and “-omics” approaches to unravel and introgress enhanced resistance against apple scab (*Venturia inaequalis*) in apple cultivars (*Malus domestica*). *Horticulture Research*, 11(2), uhad208.
- Thapa, S., Maimaitijiang, M., Gill, H. S., Rana, A., Sehgal, S. K., Halder, J., Gill, U., Bernardo, A., Ali, S., St Amand, P., & Bai, G. (2024). Integrating genomics, phenomics, and deep learning improves the predictive ability for *Fusarium* head blight-related traits in winter wheat. *The Plant Genome*, 17(3), e20427.
- Tong, J., Zhao, C., Liu, D., Jambuthenne, D. T., Sun, M., Dinglasan, E., Periyannan, S. K., Hickey, L. T., & Hayes, B. J. (2024). Genome-wide atlas of rust resistance loci in wheat. *Theoretical and Applied Genetics*, 137(8), 177.
- Uwimana, B., Vuylsteke, M., Mwanje, G., Swennen, R., Kanaabi, R., Shah, T., Mahuku, G. S., Nakato, G. V., Akech, V., & Nasuuna, C. (2024). Identification of the Loci Associated with Resistance to Banana *Xanthomonas* Wilt (*Xanthomonas vasicola* pv. *musacearum*) Using DaRTSeq Markers and Continuous Mapping. *Horticulturae*, 10(1), 87.
- Van Asseldonk, M., Ruben, R., Pamuk, H., Girvetz, E., & Wattel, C. (2023). Policy incentives for smallholder adoption of climate-smart agricultural practices. *Frontiers in Political Science*, 5, 1187640.
- Vashisht, V., Alptekin, A., Vashisht, A., Srinivas, A., Farmaha, J., Singh, H., Ahluwalia, P., Mondal, A. K., & Kolhe, R. (2023). Genomics for Emerging Pathogen Identification and Monitoring: Prospects and Obstacles. *BioMedInformatics*, 3(4), 1145–1177.

- Virk, V., Taneja, K., Deepak, H., Srivastava, R., & Giri, S. (2024). Amelioration in nanobiosensors for the control of plant diseases: current status and future challenges. *Frontiers in Nanotechnology*, 6, 1375369.
- Wang, F., Gan, M., Jiang, B., Zhang, M., Wu, B., Ning, S., Liu, D., Zhang, L., Hu, Y., Hao, M., Chen, X., Huang, L., Chen, X., & Yuan, Z. (2023). Pyramiding of Adult-Plant Resistance Genes Enhances All-Stage Resistance to Wheat Stripe Rust. *Plant Disease*, 107(3), 879–885.
- Watanabe, S., Greenwell, A. M., & Bressan, A. (2013). Localization, concentration, and transmission efficiency of Banana bunchy top virus in four asexual lineages of *Pentalonia* aphids. *Viruses*, 5(2), 758–776.
- Womack, E. D., Williams, W. P., Smith, J. S., Warburton, M. L., & Bhattaramakki, D. (2020). Mapping Quantitative Trait Loci for Resistance to Fall Armyworm (Lepidoptera: Noctuidae) Leaf-Feeding Damage in Maize Inbred Mp705. *Journal of Economic Entomology*, 113(2), 956–963.
- Wray-Cahen, D., Hallerman, E., & Tizard, M. (2024). Global regulatory policies for animal biotechnology: overview, opportunities and challenges. *Frontiers in Genome Editing*, 6, 1438997.
- Xing, Y., & Wang, X. (2024). Precision Agriculture and Water Conservation Strategies for Sustainable Crop Production in Arid Regions. *Plants (Basel, Switzerland)*, 13(22), 3184.
- Yadav, A., & Yadav, K. (2025). Portable solutions for plant pathogen diagnostics: development, usage, and future potential. *Frontiers in Microbiology*, 16, 1583023.
- Yazdani, M., Ashraf, R., Johansson, E., Vallenback, P., Hovmøller, M. S., Patpour, M., & Rahmatov, M. (2025). Marker-assisted selection for combining stem rust and stripe rust resistance in wheat using rye derived genes. *Crop Science*, 65(3), e70080.
- Yıldırım, K., Miladinović, D., Sweet, J., Akin, M., Galović, V., Kavas, M., Zlatković, M., & De Andrade, E. (2023). Genome editing for healthy crops: traits, tools and impacts. *Frontiers in Plant Science*, 14(12), 1288597.
- Younas, M. U., Qasim, M., Ahmad, I., Feng, Z., Iqbal, R., Jiang, X., & Zuo, S. (2024). Exploring the molecular mechanisms of rice blast resistance and advances in breeding for disease tolerance. *Molecular Biology Reports*, 51(1), 220.
- Yuan, H., Cheng, M., Fan, F., Zheng, X., Wang, R., Si, F., Luo, X., Li, N., & Li, S. (2024). OsGRF6-OsYUCCA1/OsWRKY82 Signaling Cascade Upgrade Grain Yield and Bacterial Blight Resistance in Rice. *Advanced Science (Weinheim, Baden-Wurtemberg, Germany)*, 11(46), e2402099.
- Zdrzałek, R., Xi, Y., Langner, T., Bentham, A. R., Bentham, A. R., Petit-Houdenet, Y., De La Concepcion, J. C., De La Concepcion, J. C., Harant, A., Shimizu, M., Were, V., Talbot, N. J., Terauchi, R., Terauchi, R., Kamoun, S., & Banfield, M. J. (2024). Bioengineering a plant NLR immune receptor with a robust binding interface toward a conserved fungal pathogen effector. *Proceedings of the National Academy of Sciences*, 121(28), e2403606121.
- Zuberi, M., Spies, M., & Nielsen, J. O. (2023). Is there a future for smallholder farmers in bioeconomy The case of 'improved' seeds in South Punjab, Pakistan. *Forest Policy and Economics*, 158, 103100.