

Trehalose-Induced Regulations in Physio-biochemical Attributes of *Helianthus annuus* L. under Lead Stress

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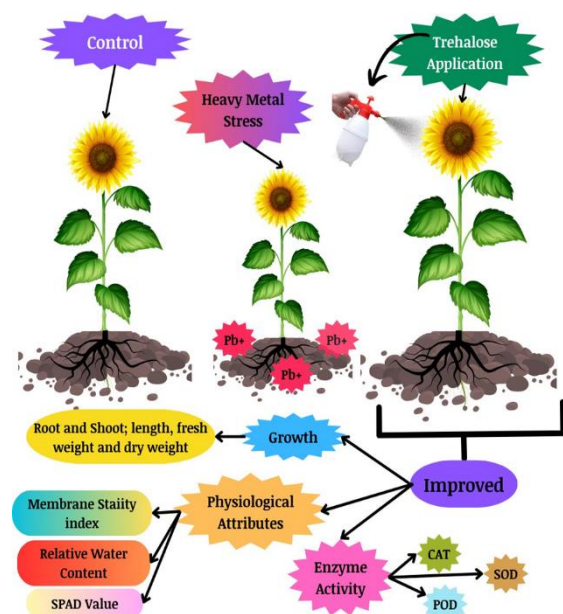
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Highlights

- Heavy metal stress significantly impairs various aspects of plant growth and physiology, including root and shoot length, biomass production, plant water relations, membrane stability, chlorophyll content, and antioxidant enzyme activities.
- The application of trehalose effectively mitigates the detrimental effects of heavy metal lead acetate stress on plants. Trehalose treatment leads to improvements in plant physiology, enhanced water relations, and strengthened antioxidant defense mechanisms.
- Trehalose treatment results in reduced levels of stress markers such as malondialdehyde (MDA) and hydrogen peroxide (H₂O₂), indicating a decrease in oxidative stress in plants subjected to heavy metal toxicity.

Graphical Abstract



Article History

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ABSTRACT

Heavy metal is a substantially severe abiotic environmental stress that is rising in many regions of the world. Current climate change circumstances and rapid industrialization increased heavy metal accumulation in soil and water. Overall, worldwide agricultural production decreased. Trehalose, a well-known plant growth regulator, is crucial in plants' responses to a variety of abiotic conditions. The Morpho-physiological and biochemical parameters were analyzed in hydroponic research to identify the possible role of trehalose in alleviating heavy metal lead acetate stress in sunflower (*Helianthus annuus* L.) crops. In the sole and mixed form, two Pb(C₂H₃O₂)₂ (Control and 30 micron) and

one trehalose (50 M L^{-1}) levels were used, and a half-strength Hoagland solution was used as a nutritional medium. The current study found that heavy metal stress had a significant negative impact on plant growth (root and shoot length), fresh and dry biomass, and plant water relations, as well as poor plant water relations, membrane stability, chlorophyll content, and antioxidant enzyme activities such as superoxide dismutase (SOD), peroxide (POD), ascorbate peroxide (APX), and catalase (CAT). MDA and H_2O_2 are the stress markers reduced by the application of trehalose. Exogenous treatment of trehalose, on the other hand, significantly mitigates the detrimental effects of metal toxicity by altering plant physiology, enhancing water relations, and providing robust antioxidant defense against heavy metal lead acetate stress. The findings of this study provide a new dimension to scientists' understanding of the beneficial usage of trehalose in plant science and offer up new research opportunities.

Keywords: Metal toxic environment, Trehalose, Sunflower, Antioxidant Defense, RWC, Lead.

INTRODUCTION

The sunflower, (*Helianthus annuus* L.), ranks as the 4th largest oilseed crop worldwide, following soybean, palm oil, and canola. In Pakistan, it ranks as the third most important oilseed crop, following rapeseed. Heavy metal pollution is a ubiquitous problem caused by anthropogenic activities [1]. These significantly impact Agri-production losses [2]. The first decade of this century perceived a substantial improvement in the crop yield of sunflowers; however, the average crop yield has gradually decreased since 2010 [3]. Heavy metals impose strong and detrimental effects on plants' morpho-physical, biochemical, and osmotic attributes. Some essential heavy metals i.e., Cu, Co, Fe, Mg, Mo, Ni, and Zn play their role in plant development but, lead (Pb) makes stable complexes such as lead phosphate $\text{Pb}_3(\text{PO}_4)_2$, lead carbonate PbCO_3 , lead sulfate PbSO_4 , and hydroxides $\text{Pb}(\text{OH})_2$, which degrade fertility of soil and crop yields in the agricultural sector [4]. Under current climate conditions globally 70% of agricultural yield is affected by heavy metals while 3.5 % [5]. of the world's geographical area is unaffected by it [6]. United States, China, India, Japan, and Pakistan, almost around the world have reported that Pb concentration in soils has gradually increased and become a key factor in reducing crop yields [7] [8]. This study focused on lead acetate $\text{Pb}(\text{C}_2\text{H}_3\text{O}_2)_2$ as stress for the hydroponic experiment on sunflowers. Lead acetate $\text{Pb}(\text{C}_2\text{H}_3\text{O}_2)_2$ is not biocompatible and its exposure to plants for a long time has a deleterious impact on seed growth, development of seedlings, and transpiration. It affects root growth, synthesis of chlorophyll, and cell division in plants [9, 10]. In exposure to heavy metals,

plants must respond to trigger changes in a wide range of physiological and biochemical attributes to develop or adopt a series of strategies that allow them to manage the toxic consequences of heavy metals i.e., $\text{Pb}(\text{C}_2\text{H}_3\text{O}_2)_2$.

Synthetic organic osmolytes like salicylic acid [11], Polyamine [12], abscisic acid [13], glycine betaine [14] melatonin [15, 16] and trehalose [17] are known as plant growth regulators; exogenously applied to crops help them to become more stress-tolerant [18]. Trehalose plays a significant role in plant functioning and it is considered to be a potential Osmo protectant. It stabilizes the cellular membranes, enzymes, proteins and protects the biological system from the damaging impacts of stress on plants [19, 20]. Trehalose protects the biological structures from toxicity by lead $\text{Pb}(\text{C}_2\text{H}_3\text{O}_2)_2$ stress through preventing damage during desiccation, through serving as a signaling molecule as antioxidant molecules and enzymes [21].

Numerous species, produce trehalose (Tre), a non-reducing disaccharide of glucose composed of dual units of glucose [22]. Trehalose is assumed to function by removing reactive oxygen species (ROS) and safeguarding the machinery which has to take charge of protein synthesis [23]. By subjecting sunflowers, to increasing levels of heavy metals aim is to determine the relative tolerance capacity of sunflowers in the presence of trehalose and measure their stress tolerance utilizing morpho-physiological, biochemical, antioxidative, and photosynthetic attributes.

Sunflower is an oilseed crop grown for edible oil production in the world [24]. About 87 % of vegetable oil is produced by sunflower [25]. This crop is well-liked among farmers because of its

short lifespan, adaptability to abiotic stresses, and high cash return. The 3rd most important oilseed crop in Pakistan after Cotton, Rapeseed, and Mustard [26]. Due to the lower latitude of Pakistan, the disastrous effects of climate change on agricultural productivity result in Pakistan's persistently getting worse production of edible oil. The sunflower crop was grown on 0.09 M hectares, producing 0.11 and 0.04 M tons of seed and oil, respectively [27]. Recent studies have provided evidence indicating that plants possess the ability to uptake trehalose from exogenous sources, leading to its subsequent accumulation in various plant tissues. The observed phenomenon has been found to yield advantageous results for agricultural production. This study investigates the impact of heavy metal toxicity on several sunflower cultivars. Investigating the morpho-physiological and biochemical reactions of these cultivars about the changing Agricultural sector and prevailing environmental circumstances. The research hypothesis, trehalose application may have the capacity to mitigate the negative effects caused by exposure to metals. The work is a comprehensive protective mechanism that incorporates a range of tactics. These strategies are designed to safeguard biological membranes, enhance plant biomass, fortify antioxidative capacity, optimize nutrient levels, and preserve chlorophyll pigments in sunflower crops. The primary objective of this research was as follows: to assess the efficacy of trehalose-induced tolerance in sunflower plants exposed to stress generated by lead acetate. Moreover, the primary aim of this research is to evaluate the influence of trehalose on many aspects such as morphology, physiology, biochemistry, and antioxidant enzyme activities. Overall, this study examines the degree of resistance demonstrated by different sunflower cultivars in response to the toxicity induced by lead acetate.

Our results emphasize the vital role of trehalose in enhancing sunflower plant development and resistance under heavy metal stress and use this knowledge and information to extract the potential yield of sunflower in soils that have been impacted by heavy metals. This study also revealed that sunflowers can be used potentially as a phytoextractor.

MATERIAL AND METHODS

Experimental Description

The hydroponic system was successfully implemented at wire house. The above-mentioned institution is situated at an elevation of 116 meters above sea level at The Islamia University of Bahawalpur, Pakistan. The sunflower cultivars were planted in a substrate consisting of sand, characterized by elevated levels of humidity. These cultivars were subjected to an average daily duration of sunlight ranging from 10 to 12 hours, accompanied by a temperature of 30 °C. Before the commencement of the planting process, the seeds underwent sterilization by a thorough washing procedure including a solution comprising 2.5 % sodium hypochlorite (NaOCl). The seedlings that exhibited robust health and possessed comparable genetic traits were carefully retrieved, with meticulous consideration paid to the preservation of their root systems, and subsequently rinsed using distilled water. Once the seedlings have reached the stage of developing two leaves, they are carefully transplanted onto Polystyrene sheets. These sheets are then placed within plastic tubs, each with a capacity of 20 L. Each tub possesses the capability to accommodate a maximum of nine plants. The research utilized a metal acute toxicity level of 30 μM , achieved using the precise combination of lead acetate $\text{Pb}(\text{C}_2\text{H}_3\text{O}_2)_2$. After the transplantation of the seedlings, the application of trehalose was subsequently administered at a concentration of 50 $\mu\text{mol L}^{-1}$, both in its individual and combined forms, following a two-week gap. The study was conducted using a fully randomized design, comprising three replications. The research employed three distinct treatment groups: the Control group, which did not receive $\text{Pb}(\text{C}_2\text{H}_3\text{O}_2)_2$ or trehalose supplementation; the $\text{Pb}(\text{C}_2\text{H}_3\text{O}_2)_2$ 30 μM lead stress group; and the Pb^+ Tre group, which experienced 30 μM lead stress along with 50 $\mu\text{mol L}^{-1}$ trehalose supplementation. The delivery of oxygen to the plants was facilitated through the utilization of aeration pumps. At the same time, a nutritional medium consisting of a Hoagland solution at a concentration of 50 % was utilized. The nutritional media utilized in this investigation consisted of various constituents L^{-1} . The components used in the experiment consisted of

KH_2PO_4 at a concentration of 68 grams, KNO_3 at a concentration of 101 grams, $\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ at a concentration of 118 grams, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ at a concentration of 61.5 grams, H_3BO_3 at a concentration of 2.86 grams, MnCl_2 at a concentration of 1.81 grams, ZnSO_4 at a concentration of 0.22 grams, $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ at a concentration of 0.08 grams, $\text{H}_2\text{MoO}_4 \cdot \text{H}_2\text{O}$ at a concentration of 0.02 grams and Fe-EDTA at a concentration of 18.6 grams. The replenishment of the nutrition solution occurred on a weekly frequency. The pH of the growth medium was consistently maintained at a value of 6.0 ± 0.5 throughout the whole duration of the experiment.

Plant Growth

The plant growth is significantly increased by height and weight. There are various distinct standards used in scientific literature to determine plant growth rates. Plant harvesting was done after 4 weeks of the establishment of treatments with stress. Plants are washed with tap water and quickly transferred to the laboratory.

Chlorophyll Content

Leaf chlorophyll concentration (SPAD) is measured by using a portable chlorophyll SPAD meter (SPAD-502-Minolta Japan).

Relative Water Contents (RWC)

Fresh leaf tissues were selected and after obtaining their fresh weight (FW), leaves were soaked for 24 hours to determine their turgid weight (TW). Subsequent samples were dried at 80°C to get their dry weight (DW) (Lazcano-Ferrat and Lovatt, 1999).

$$\text{RWC \%} = (\text{FW} - \text{DW}) / (\text{TW} - \text{DW}) \times 100$$

Membrane Stability Index (MSI)

MSI was determined using the method described by [30]. Leaf disc (100 mg) was heated in a water bath for 30 minutes at 40°C in distilled water (10 ml), and the EC was determined (C_1). The same samples were heated at 100°C for ten minutes, and EC was calculated as C_2 . The membrane stability index was computed as $\text{MSI \%} = [1 - (C_1/C_2) \times 100]$.

Stress Marker MDA and H_2O_2

The methodology developed by Velikova was utilized to evaluate the amounts of hydrogen peroxide (H_2O_2) [31]. The determination of H_2O_2 concentration was conducted by assessing the optical density at a wavelength of 390 nm, employing a standard curve.

The examined-on lipid degradation in leaves encompassed the evaluation of malondialdehyde (MDA) concentrations, employing a leaf sample size of 200 mg. The foliage was submerged in a solution including trichloroacetic acid at a concentration of 0.1%. Following this, the samples underwent centrifugation at a magnitude of 10,000 times the acceleration due to gravity for 10 minutes. A volume of 1 ml of the supernatant was mixed with a solution of TBA with a concentration of 0.5 %.

The combination obtained was subjected to heat treatment at a temperature of 95°C for 30 minutes. Subsequently, the mixture recorded centrifugation. The measurement of absorbance in the supernatant was conducted at a wavelength of 532 nm, and any non-specific absorbance detected at 600 nm was subtracted to correct the results. The quantification of MDA content was conducted by employing the extinction coefficient of $155 \text{ mM}^{-1} \text{ cm}^{-1}$ and the results were reported in terms of nmol g^{-1} fresh weight [32].

Antioxidant Enzymes

The collection of samples and the subsequent assessment of absorbance are conducted under controlled temperature conditions ranging from 0 to 4°C . The leaf fragments were gathered and homogenized using an extraction buffer solution consisting of 10 mL, containing 50 mM phosphate with a pH of 7.8, 1 g of polyvinyl pyrrolidone (PVP), 1 mM of EDTA, and 0.5 % Triton X-100, following the procedure described by Kamran [33]. The solution underwent centrifuging at a rotating speed of 12,000 rpm for 20 minutes. Subsequently, the liquid component, with a volume of approximately 2-3 mL, supernatant, was collected and utilized for enzyme analyses. The supernatant that was collected was used to investigate the activity of superoxide dismutase (SOD) using the NBT approach, with absorbance measurements taken at 560 nm [34]. The assessment of peroxidase (POD) activity was conducted by employing guaiacol, hydrogen peroxide (H_2O_2), and a KH_2PO_4 buffer solution supplemented with EDTA at a pH level of 7. The measurements were conducted at a specific wavelength of 470 nm utilizing a spectrophotometer. The assessment of catalase (CAT) activity [35]. In the study conducted by [34], the evaluation of ascorbate peroxidase (APX) activity was carried out using

an enzyme extract consisting of potassium phosphate buffer and EDTA. The quantification and documentation of changes in light absorption levels at a specific wavelength of 290 nm were performed.

Statistical Analysis

The data acquired from the present experiment underwent analysis utilizing the statistical program Statistics 8.1 (Statistics USA) and is displayed graphically. The bars depicted in the graph represent the mean value derived from three separate replicates, while the error bars indicate the standard deviation.

RESULTS

Plant Growth and Biomass

This study aimed to evaluate the effects of a substantial lead concentration (30 μM) on the morphological traits of many sunflower cultivars.

The experimental procedure involved the inclusion and exclusion of trehalose (50 $\mu\text{mol L}^{-1}$) as an exogenous supplement. The results obtained from the experiment revealed a decrease of 27 % in shoot fresh weight, 34% in shoot dry weight, 26 % in root fresh weight, 36 % in root dry weight, 11 % in shoot length, and 30 % in root length. However, the use of trehalose led to a significant improvement in various growth metrics. The shoot fresh weight, shoot dry weight, root fresh weight, root dry weight, shoot length, and root length demonstrated statistically significant enhancements of 23 %, 21 %, 22 %, 31 %, 5 %, and 21 % respectively, in comparison to plants grown in a metal-contaminated substrate without the application of trehalose (Pb) mentioned in figure 1 and 2.

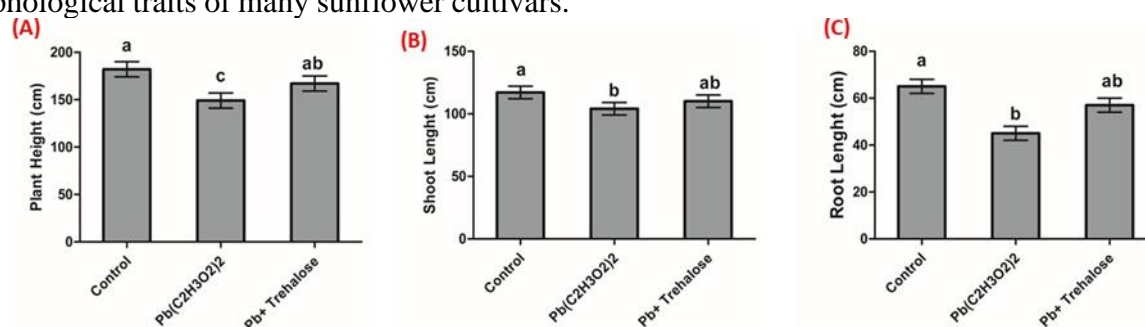


Figure. 1 The effect of exogenous trehalose (Tre) supply on (A) Plant height (cm) (B) Shoot length (cm) (C) Root length (cm) of Sunflower (*Helianthus Annuus* L.) cultivars subjected to heavy metal stress.

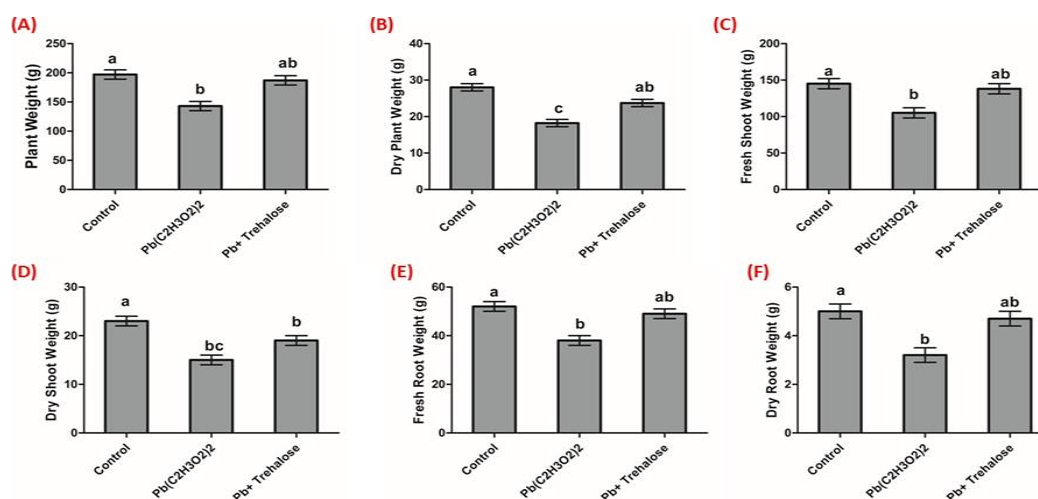


Figure. 2 The effect of exogenous trehalose (Tre) supply on (A) Fresh plant weight (g) (B) Dry plant weight (g) (C) Fresh shoot weight (D) Dry shoot weight (g) (E) Fresh root weight (g) (F) Dry root weight (g) of Sunflower (*Helianthus Annuus* L.) cultivars subjected to heavy metal stress.

Physiological Attributes

The results about the comparative water levels, photosynthetic pigments, and membrane stability index of a hybrid sunflower variety when exposed

to metal-induced stress conditions, specifically lead. In comparison to the control treatment, the relative water contents, membrane stability, and chlorophyll index exhibited a reduction of 38.16

%, 36.71 %, and 42.23 % respectively. The incorporation of trehalose in conjunction with metal stress (Pb + Tre) led to a notable augmentation in the levels of relative water contents (RWC), membrane stability index (MSI), and chlorophyll in sunflower seedlings.

The introduction of trehalose in a metal-hazardous media resulted in significant increases in the RWC, MSI, and chlorophyll levels by 21.67 %, 19.35 %, and 29.73 % respectively, as compared to plants grown in the absence of trehalose mentioned in Figure 3.

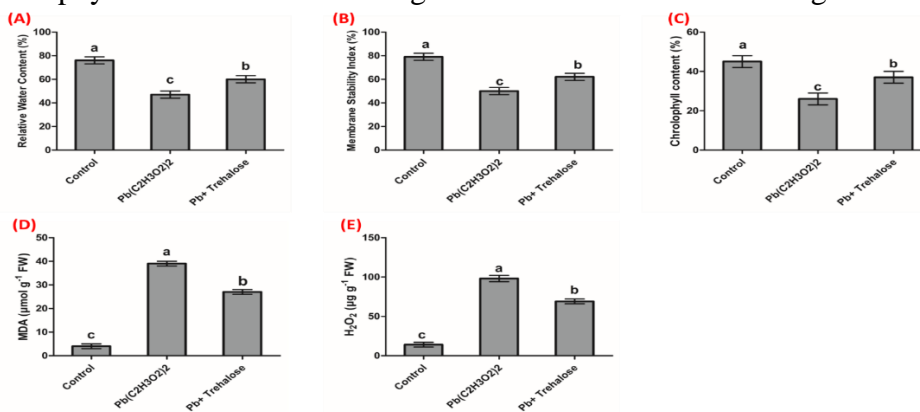


Figure 3 The effect of exogenous trehalose (Tre) supply on (A) Relative water content (%) (B) Membrane stability index (%) (C) Chlorophyll content (%) (D) MDA ($\mu\text{mol g}^{-1}$ FW) and H₂O₂ ($\mu\text{g g}^{-1}$ FW) of Sunflower (*Helianthus annuus* L.) cultivars subjected to heavy metal stress.

Antioxidant Enzyme Activities

The present study aimed to examine the impact of heavy metal toxicity, specifically at a concentration of 30 μM , on the enzymatic activity of several antioxidative enzymes in hybrid sunflower seedlings. The evaluation also encompassed the examination of the effects of trehalose supplementation at a concentration of 50 $\mu\text{mol L}^{-1}$. The findings indicate that the presence of heavy metals resulted in a reduction in the activity of antioxidant enzymes in sunflower plants. The investigation unveiled a significant decrease in the enzymatic functions of superoxide dismutase (SOD), peroxidase (POD), catalase (CAT), and ascorbate peroxidase (APX). The aforementioned decreases

corresponded to percentages of 58.64 %, 26.09 %, 38.78 %, and 47.44%, respectively, about plants that were grown under standard conditions, which served as the control treatment. Nevertheless, previous studies have provided evidence indicating that the use of trehalose resulted in a significant augmentation in the enzymatic functions of superoxide dismutase (SOD), peroxidase (POD), catalase (CAT), and ascorbate peroxidase (APX) by 18.75%, 15 %, 23.08 %, and 18 %, correspondingly. The reported augmentation was noted in plants cultivated in a substrate containing elevated concentrations of heavy metals, although lacking the inclusion of trehalose mentioned in Figure 4.

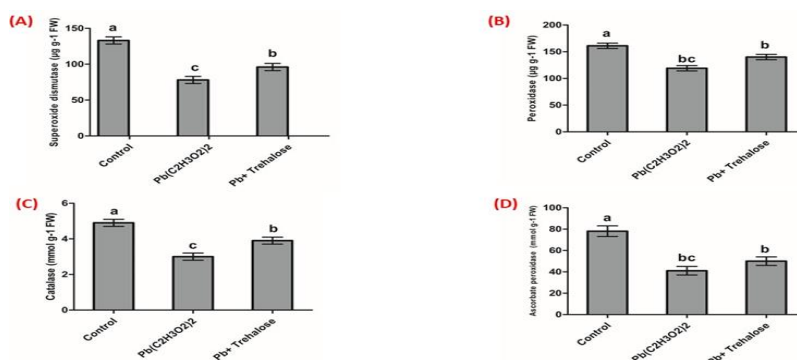


Figure 4 The effect of exogenous trehalose (Tre) supply on (A) SOD (B) POD (C) CAT (D) APX of Sunflower (*Helianthus Annuus* L.) cultivars subjected to heavy metal stress.

DISCUSSION

The sunflower, (*Helianthus annuus* L.), ranks as the 4th largest oilseed crop worldwide, following

soybean, palm oil, and canola. In Pakistan, it ranks as the third most important oilseed crop,

following rapeseed. The practice of growing crops in regions impacted by industrial activities is linked to efforts aimed at mitigating the issues related to the absorption and buildup of heavy metals in plants. The intricate interplay between metallurgical facilities and agricultural regions exerts a multifaceted impact on crops, mostly attributed to the uptake of heavy metals by plant roots and the infiltration of aerosol pollutants through the leaf epidermis. As per the findings of Aslam, the occurrence of heightened levels of metal toxicity in the growth medium has been associated with disruptions in morphological traits and physical attributes, alongside the suppression of antioxidant defense enzyme activity [39]. As a result, scientists have determined that this phenomenon harms the productivity of agricultural commodities [40]. A study that revealed the effectiveness of trehalose in mitigating the adverse effects of stress induced by heavy metals [41]. There is a direct association between increased concentrations of lead in the growth medium and a decline in certain plant growth attributes. The decline in performance can be ascribed to the instability of the membrane and the disturbance of photosynthetic processes. As a result, an imbalance arises in the course of nutritional intake, the discontinuation of cellular division, and the generation of reactive oxygen species (ROS). The combined effects of these factors have a substantial impact on the biomass of plants [40]. The exogenous administration of trehalose has been seen to elicit diverse impacts on plant physiology, both in the presence of stress and under non-stressful conditions. The molecule

known as trehalose has been found to have multiple benefits in the mitigation of lead risks. The aforementioned benefits encompass the ability to acclimate to oxidative stress, augment the antioxidant defense mechanism, and elevate photosynthetic efficacy. According to the findings of Ashraf their analysis demonstrated that the existence of metal toxicity negatively impacts plant water relations and undermines the structural integrity of cellular membranes[38]. The observed decrease in performance was seen in the cultivated sunflower hybrids. The escalating occurrence of heavy metals in soil has a discernible influence on the relative water content (RWC) and membrane stability index (MSI), resulting in a decline in plant water absorption, leaf succulence, compromised membrane stability, and impairment of cellular membranes [39, 40, 43].

The measurement of malondialdehyde (MDA) is a dependable indicator used to assess the presence of lipid peroxidation mechanisms in the cellular membrane. Therefore, the observed elevation in malondialdehyde (MDA) levels resulting from heavy metal stress addiction indicates the presence of oxidative stress induced by excessive production of reactive oxygen species (ROS). In their investigation, observed a positive association between the presence of heavy metal-contaminated soil and the generation of hydrogen peroxide (H_2O_2), leading to an increase in lipid peroxidation. Their work observed a significant decrease in H_2O_2 levels and subsequent reduction in H_2O_2 -induced lipid peroxidation.

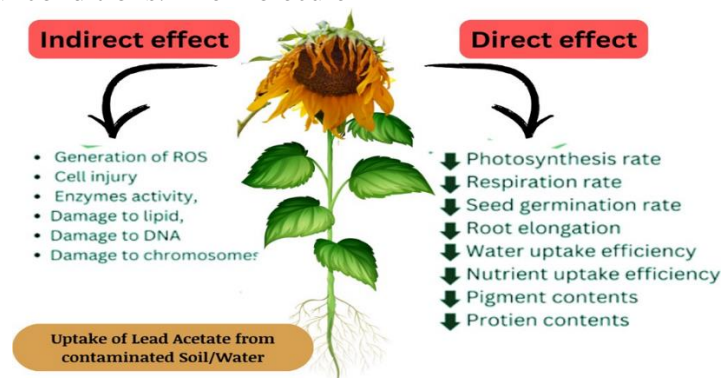


Figure. 5 Uptake of $PbCH_3COOH$ from contaminated soil, water, or air and its effect on sunflower

Figure 5 illustrates the impacts of elevated levels of Pb on plant species, spanning both direct and indirect pathways. The immediate consequences

encompass a reduction in the rates of photosynthesis and respiration, a diminished rate of seed germination, a decline in the elongation of

roots, a decreased efficiency in the absorption of water and nutrients, and a decrease in the concentrations of pigments and proteins. The idea of indirect impacts encompasses various pathways, including the production of reactive

oxygen species (ROS), cellular harm, modification of enzyme activity, and disturbance of lipid, DNA, and chromosomal integrity [40, 43].

References

1. Briffa, J., Sinagra, E., & Blundell, R. (2020). Heavy metal pollution in the environment and their toxicological effects on humans. *Heliyon*, 6(9).
2. Ashfaq, F., Inam, A., Sahay, S., & Iqbal, S. (2016). Influence of heavy metal toxicity on plant growth, metabolism and its alleviation by phytoremediation-a promising technology. *Journal of Agriculture and Ecology Research International*, 6(2), 1-19.
3. Butt, A., Umer, S., & Altaf, R. (2020). Under the Stress of Heavy Metals ie Zinc and Copper. *Journal Clean WAS (JCleanWAS)*, 4(1), 08-11.
4. Sarwar, N., Imran, M., Shaheen, M. R., Ishaque, W., Kamran, M. A., Matloob, A., ... & Hussain, S. (2017). Phytoremediation strategies for soils contaminated with heavy metals: modifications and future perspectives. *Chemosphere*, 171, 710-721.
5. Fatima, R., Mahmood, T., Moosa, A., Aslam, M. N., Shakeel, M. T., Maqsood, A., ... & Al-Shehri, M. (2023). *Bacillus thuringiensis* CHGP12 uses a multifaceted approach for the suppression of *Fusarium oxysporum* f. sp. *ciceris* and to enhance the biomass of chickpea plants. *Pest Management Science*, 79(1), 336-348.
6. Jewell, M. C., Campbell, B. C., & Godwin, I. D. (2010). Transgenic plants for abiotic stress resistance. *Transgenic crop plants*, 67-132.
7. Waseem, A., Arshad, J., Iqbal, F., Sajjad, A., Mehmood, Z., & Murtaza, G. (2014). Pollution status of Pakistan: a retrospective review on heavy metal contamination of water, soil, and vegetables. *BioMed research international*, 2014.
8. Lu, S., Wang, J., & Pei, L. (2016). Study on the effects of irrigation with reclaimed water on the content and distribution of heavy metals in soil. *International Journal of Environmental Research and Public Health*, 13(3), 298.
9. Pourrut, B., Shahid, M., Dumat, C., Winterton, P., & Pinelli, E. (2011). Lead uptake, toxicity, and detoxification in plants. *Reviews of environmental contamination and toxicology* volume 213, 113-136.
10. Kumar, B., Smita, K., & Flores, L. C. (2017). Plant mediated detoxification of mercury and lead. *Arabian Journal of Chemistry*, 10, S2335-S2342.
11. Bastam, N., Baninasab, B., & Ghobadi, C. (2013). Improving salt tolerance by exogenous application of salicylic acid in seedlings of pistachio. *Plant growth regulation*, 69, 275-284.
12. Saha, J., Brauer, E. K., Sengupta, A., Popescu, S. C., Gupta, K., & Gupta, B. (2015). Polyamines as redox homeostasis regulators during salt stress in plants. *Frontiers in Environmental Science*, 3, 21.
13. Zhang, J., Jia, W., Yang, J., & Ismail, A. M. (2006). Role of ABA in integrating plant responses to drought and salt stresses. *Field crops research*, 97(1), 111-119.
14. Yang, X., & Lu, C. (2005). Photosynthesis is improved by exogenous glycinebetaine in salt-stressed maize plants. *Physiologia Plantarum*, 124(3), 343-352.
15. Liu, N., Jin, Z., Wang, S., Gong, B., Wen, D., Wang, X., ... & Shi, Q. (2015). Sodic alkaline stress mitigation with exogenous melatonin involves reactive oxygen metabolism and ion homeostasis in tomato. *Scientia Horticulturae*, 181, 18-25.
16. Zahedi, S. M., Hosseini, M. S., Abadía, J., & Marjani, M. (2020). Melatonin foliar sprays elicit salinity stress tolerance and enhance fruit yield and quality in strawberry (*Fragaria × ananassa* Duch.). *Plant Physiology and Biochemistry*, 149, 313-323.
17. Zulfiqar, F., Chen, J., Finnegan, P. M., Younis, A., Nafees, M., Zorrig, W., & Hamed, K. B. (2021). Application of trehalose and salicylic acid mitigates drought stress in sweet basil and improves plant growth. *Plants*, 10(6), 1078.

18. Malik, J., Moosa, A., Zulfiqar, F., Aslam, M. N., Albalawi, M. A., Almowallad, S., ... & Yong, J. W. H. (2024). Biocontrol potential of lipopeptides produced by the novel *Bacillus altitudinis* strain TM22A against postharvest *Alternaria* rot of tomato. *LWT*, *191*, 115541.
19. Hossain, A., Ahmad, Z., Adeel, M., Rahman, M. A., Alam, M. J., Ahmed, S., & Aftab, T. (2021). Emerging roles of osmoprotectants in heavy metal stress tolerance in plants. In *Heavy Metal Toxicity in Plants* (pp. 95-110). CRC Press.
20. Luo, Y., Xie, Y., Li, W., Wei, M., Dai, T., Li, Z., & Wang, B. (2021). Physiological and transcriptomic analyses reveal exogenous trehalose is involved in the responses of wheat roots to high temperature stress. *Plants*, *10*(12), 2644.
21. Morgutti, S., Negrini, N., Pucciariello, C., & Sacchi, G. A. (2019). Role of trehalose and regulation of its levels as a signal molecule to abiotic stresses in plants. In *Plant signaling molecules* (pp. 235-255). Woodhead Publishing.
22. Elbein, A. D., Pan, Y. T., Pastuszak, I., & Carroll, D. (2003). New insights on trehalose: a multifunctional molecule. *Glycobiology*, *13*(4), 17R-27R.
23. Mahmood, A., Moosa, A., Khan, A. U. R., Maqsood, A., Kiani, B., Abbas, C., ... & Khalid, E. (2023). Using essential oils to protect peaches from post-harvest rot caused by *Rhizopus* species.
24. Adeleke, B. S., & Babalola, O. O. (2020). Oilseed crop sunflower (*Helianthus annuus*) as a source of food: Nutritional and health benefits. *Food Science & Nutrition*, *8*(9), 4666-4684.
25. Bassegio, D., Zanotto, M. D., Santos, R. F., Werncke, I., Dias, P. P., & Olivo, M. (2016). Oilseed crop crambe as a source of renewable energy in Brazil. *Renewable and Sustainable Energy Reviews*, *66*, 311-321.
26. Nasim, W., Ahmad, A., Belhouchette, H., Fahad, S., & Hoogenboom, G. (2016). Evaluation of the OILCROP-SUN model for sunflower hybrids under different agro-meteorological conditions of Punjab—Pakistan. *Field crops research*, *188*, 17-30.
27. Awais, M., Wajid, A., Saleem, M. F., Nasim, W., Ahmad, A., Raza, M. A. S., ... & Hussain, J. (2018). Potential impacts of climate change and adaptation strategies for sunflower in Pakistan. *Environmental Science and Pollution Research*, *25*(14), 13719-13730.
28. Ahmad, Z., Abbas, H., Murtaza, T., Khan, A. U. R., Ali, A., Zahid, K., ... & Habib, A. (2023). Assessment of responses of peach cultivars to postharvest pathogen *Botrytis cinerea* and its mitigation using plant essential oils.
29. Lazcano-Ferrat, I., & Lovatt, C. J. (1999). Relationship between relative water content, nitrogen pools, and growth of *Phaseolus vulgaris* L. and *P. acutifolius* A. Gray during water deficit. *Crop Science*, *39*(2), 467-475.
30. Sairam, R. K., & Saxena, D. C. (2000). Oxidative stress and antioxidants in wheat genotypes: possible mechanism of water stress tolerance. *Journal of Agronomy and Crop Science*, *184*(1), 55-61.
31. Velikova, V., Yordanov, I., & Edreva, A. J. P. S. (2000). Oxidative stress and some antioxidant systems in acid rain-treated bean plants: protective role of exogenous polyamines. *Plant science*, *151*(1), 59-66.
32. Mahmood, T., Moosa, A., Ahmad, W., Malik, J., Shafiq, M. U., Yousaf, M., ... & Abbas, G. (2023). Lipopeptides: Powerful Antifungal Compounds Produced by *Bacillus* Species: A Review. *Plant Protection*, *7*(3), 605-614.
33. Kamran, M., Malik, Z., Parveen, A., Huang, L., Riaz, M., Bashir, S., ... & Ali, U. (2020). Ameliorative effects of biochar on rapeseed (*Brassica napus* L.) growth and heavy metal immobilization in soil irrigated with untreated wastewater. *Journal of Plant Growth Regulation*, *39*, 266-281..
34. Giannopolitis, C. N., & Ries, S. K. (1977). Superoxide dismutases: I. Occurrence in higher plants. *Plant physiology*, *59*(2), 309-314.
35. Change, B., & Maehly, A. C. (1955). Assay of catalases and peroxidase. *Methods Enzymol*, *2*, 764-775.
36. Prochazkova, D., Sairam, R. K., Srivastava, G. C., & Singh, D. V. (2001). Oxidative stress and antioxidant activity as the basis of senescence in maize leaves. *Plant Science*, *161*(4), 765-771.
37. Ali, M., Kamran, M., Abbasi, G. H., Saleem, M. H., Ahmad, S., Parveen, A., ... & Fahad, S. (2021). Melatonin-induced salinity tolerance by ameliorating osmotic and oxidative stress in the seedlings of two tomato (*Solanum lycopersicum* L.) cultivars. *Journal of Plant Growth Regulation*, *40*, 2236-2248.

38. Ashraf, A., Bhardwaj, S., Ishtiaq, H., Devi, Y. K., & Kapoor, D. (2021). Lead uptake, toxicity and mitigation strategies in plants. *Plant Archives* (09725210), 21(1).
39. Aslam, M. R., Waris, M., Muhammad, I., Ahmed, M., Khan, Z., Jabeen, Z., ... & Altaf, A. R. (2022). Lead-Immobilization, transformation, and induced toxicity alleviation in sunflower using nanoscale Fe^o/BC: Experimental insights with Mechanistic validations. *Journal of Plant Interactions*, 17(1), 812-823.
40. Shahid, M., Khalid, S., Abbas, G., Shahid, N., Nadeem, M., Sabir, M., ... & Dumat, C. (2015). Heavy metal stress and crop productivity. *Crop production and global environmental issues*, 1-25.
42. Maalik, S., Moosa, A., Zulfiqar, F., Aslam, M. N., & Siddique, K. H. (2023). Endophytic *Bacillus atrophaeus* CHGP13 and salicylic acid inhibit blue mold of lemon by regulating defense enzymes. *Frontiers in Microbiology*, 14, 1184297.
43. Jayasri, M. A., & Suthindhiran, K. (2017). Effect of zinc and lead on the physiological and biochemical properties of aquatic plant *Lemna minor*: its potential role in phytoremediation. *Applied Water Science*, 7, 1247-1253.
44. Abdallah, M. S., Abdelgawad, Z. A., & El-Bassiouny, H. M. S. (2016). Alleviation of the adverse effects of salinity stress using trehalose in two rice varieties. *South African Journal of Botany*, 103, 275-282.
45. Chang, B., Yang, L., Cong, W., Zu, Y., & Tang, Z. (2014). The improved resistance to high salinity induced by trehalose is associated with ionic regulation and osmotic adjustment in *Catharanthus roseus*. *Plant Physiology and Biochemistry*, 77, 140-148.
46. Shu, X., Yin, L., Zhang, Q., & Wang, W. (2012). Effect of Pb toxicity on leaf growth, antioxidant enzyme activities, and photosynthesis in cuttings and seedlings of *Jatropha curcas* L. *Environmental Science and Pollution Research*, 19, 893-902.
47. Fernandez, O., Béthencourt, L., Quero, A., Sangwan, R. S., & Clément, C. (2010). Trehalose and plant stress responses: friend or foe?. *Trends in plant science*, 15(7), 409-417.
48. Bae, H., Herman, E., Bailey, B., Bae, H. J., & Sicher, R. (2005). Exogenous trehalose alters *Arabidopsis* transcripts involved in cell wall modification, abiotic stress, nitrogen metabolism, and plant defense. *Physiologia plantarum*, 125(1), 114-126.
49. Chen, T. H., & Murata, N. (2002). Enhancement of tolerance of abiotic stress by metabolic engineering of betaines and other compatible solutes. *Current opinion in plant biology*, 5(3), 250-257.